Package ‘combiroc’

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Title  Selection and Ranking of Omics Biomarkers Combinations Made Easy
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Description Provides functions and a workflow to easily and powerfully calculating specificity, sensitivity and ROC curves of biomarkers combinations. Allows to rank and select multi-markers signatures as well as to find the best performing sub-signatures. The method used was first published as a Shiny app and described in Mazzara et al. (2017) <doi:10.1038/srep45477> and further described in Bombaci & Rossi (2019) <doi:10.1007/978-1-4939-9164-8_16>.
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classify

Classify data.frames using glm(link='binomial') models.

Description

A function that applies the previously calculated models to an unclassified dataset and classifies the samples.

Usage

classify(
  unclassified_data,
  Models,
  Metrics,
  Positive_class = 1,
  Negative_class = 0
)

Arguments

unclassified_data
  a data.frame returned by load_unclassified_data().

Models
  a list of glm() objects returned by roc_reports().

Metrics
  a list of data.frame objects containing ROC metrics, returned by roc_reports().

Positive_class
  a numeric or a character that specifies the label of the samples that will be classified as positives.

Negative_class
  a numeric or a character that specifies the label of the samples that will be classified as negatives.
**Details**
This function can classify dataset loaded with `load_unclassified_data()` that MUST contain all the markers of the classified dataset used to train the models (the one loaded with `load_data()`).

**Value**
a data.frame containing the predicted class of each sample, for each marker/combination in Models

**Examples**
demo_data # combiroc built-in demo data (proteomics data from Zingaretti et al. 2012 - PMC3518104)
demo_unclassified_data # combiroc built-in unclassified demo data

combs <- combi(data= demo_data, signalthr=450, combithr=1) # compute combinations

reports <- roc_reports(data= demo_data, markers_table= combs,
                       selected_combinations= c(1,11,15),
                       single_markers=c('Marker1', 'Marker2'), case_class='A') # train logistic
                       # regression models

# To classify new samples with logistic regression models.

classified_data <- classify(unclassified_data= demo_unclassified_data, Models= reports$Models,
                            Metrics= reports$Metrics, Positive_class=1, Negative_class=0)

classified_data # show samples classified using Logistic regression models

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**combi**  
*Compute combinations.*

**Description**
A function that computes the marker combinations and counts their corresponding positive samples for each class (once thresholds are selected).

**Usage**
```r
combi(data, signalthr = 0, combithr = 1, max_length = NULL)
```

**Arguments**
- **data** a data.frame returned by `load_data()`.
- **signalthr** a numeric that specifies the value above which a marker expression is considered positive in a given sample. Since the target of the analysis is the identification of marker combinations capable to correctly classify samples, the user should choose a signalthr that:
• Positively selects most samples belonging to the case class, which must be above signalthr.
• Negatively selects most control samples, which must be below signalthr.

combithr   a numeric that specifies the necessary number of positively expressed markers (>= signalthr), in a given combination, to consider that combination positively expressed in a sample.

max_length   an integer that specifies the max combination length that is allowed

Details
This function counts how many samples are 'positive' for each combination. A sample, to be considered positive for a given combination, must have a value higher than a given signal threshold (signalthr) for at least a given number of markers composing that combination (combithr).

Value
a data.frame containing how many samples of each class are "positive" for each combination.

Examples

demo_data # combiroc built-in demo data (proteomics data from Zingaretti et al. 2012 - PMC3518104)

# To compute the marker combinations and count their corresponding positive samples for each class.
combs <- combi(data= demo_data, signalthr=450, combithr=1) # count as positive the samples with # value >= 450 for at least 1 marker # in the combination

Description
Easily and Powerfully Calculates Specificity, Sensitivity and ROC Curves of Biomarkers Combinations. In the following sections there is a brief summary of the package content.

data loading and reshaping
• load_data(): to check and load data.
• load_unclassified_data(): to check and load unclassified data.
• combiroc_long(): to reshape data in long format.

distribution inspection
• markers_distribution(): to show distribution of intensity values for all the markers both singularly and all together.
combinatorial analysis

• combi(): to compute marker combinations.
• se_sp(): to compute sensitivity and specificity of each combination.
• ranked_combs(): to rank combinations.

logistic regression training and fitting

• roc_reports(): to train logistic regression and compute ROC.
• classify(): to apply the previously calculated models to an unclassified dataset and classifies the samples.

markers/combinations correspondence

• show_markers(): to show the composition of combinations
• combs_with(): to show all combinations with given markers.

built-in demo datasets

• demo_data: proteomics data from Zingaretti et al. 2012 - PMC3518104)
• demo_unclassified_data: dataset obtained by randomly picking 20 samples from demo_data without their classification.

---

combiroc_long

Reshape combiroc data in long format.

Description

A function that simply wraps dyplr::pivot_longer() to reshape data in long format.

Usage

combiroc_long(data)

Arguments

data a data.frame returned by load_data().

Details

This function returns the data in long format (with 'Markers' and 'Values' columns)

Value

a data.frame in long format
combs_with

Examples

demo_data # combiroc built-in demo data (proteomics data from Zingaretti et al. 2012 - PMC3518104)

# To reshape demo_data in long format
demo_data_long <- combiroc_long(data = demo_data)

demo_data # combiroc built-in demo data (proteomics data from Zingaretti et al. 2012 - PMC3518104)
combs <- combi(data= demo_data, signalthr=450, combithr=1) # compute combinations
# To show all the combinations with given markers.
combs_with(markers = c('Marker1', 'Marker2'), markers_table = combs)

---

**Combs_with**

Show combinations with given markers.

---

**Description**

A function to find all the combinations containing all the markers of interest.

**Usage**

combs_with(markers, markers_table)

**Arguments**

- **markers**: a character vector containing one or more markers of interest.
- **markers_table**: a data.frame with ranked combination, reporting: SE, SP, number of markers composing the combination and the score (returned by ranked_combs()).

**Value**

a numeric vector containing the numbers corresponding to the combinations containing all the selected markers.

**Examples**

demo_data # combiroc built-in demo data (proteomics data from Zingaretti et al. 2012 - PMC3518104)
combs <- combi(data= demo_data, signalthr=450, combithr=1) # compute combinations
# To show all the combinations with given markers.
combs_with(markers = c('Marker1', 'Marker2'), markers_table = combs)
**demo_data**  
*Combiroc built-in demo data (proteomics data from Zingaretti et al. 2012 - PMC3518104)*

**Description**

A dataset containing signal intensity values of a 5-marker signatures for Autoimmune Hepatitis (AIH). Samples have been clinically diagnosed as “abnormal” (class A) or "normal" (class B).

**Usage**

`demo_data`

**Format**

A data frame with 170 rows and 7 variables:

- **Patient.ID**  the ID of samples
- **Class**  the class of the samples: A-Healthy, B-AIH
- **Marker1**  the signal intensity value of Marker1
- **Marker2**  the signal intensity value of Marker2
- **Marker3**  the signal intensity value of Marker3
- **Marker4**  the signal intensity value of Marker4
- **Marker5**  the signal intensity value of Marker5

**demo_unclassified_data**  
*Combiroc built-in unclassified demo data*

**Description**

A dataset containing signal intensity values of a 5-marker signatures for Autoimmune Hepatitis (AIH). This dataset has been obtained by randomly picking 20 samples from demo_data without their classification.

**Usage**

`demo_unclassified_data`
Format

A data frame with 20 rows and 7 variables:

- **Patient.ID** the ID of samples
- **Marker1** the signal intensity value of Marker1
- **Marker2** the signal intensity value of Marker2
- **Marker3** the signal intensity value of Marker3
- **Marker4** the signal intensity value of Marker4
- **Marker5** the signal intensity value of Marker5

load_data

Load CombiROC data.

Description

A customized read.table() function that checks the conformity of the dataset format, and only if all checks are passed, loads it.

Usage

load_data(data, sep = ";", na.strings = "")

Arguments

- **data** the name of the file which the data are to be read from.
- **sep** the field separator character.
- **na.strings** a character vector of strings which are to be interpreted as NA values.

Details

The dataset to be analysed should be in text format, which can be comma, tab or semicolon separated:

- The 1st column must contain patient/sample IDs as characters.
- The 2nd column must contain the class to which each sample belongs.
- The classes must be exactly 2 and they must be written in character format.
- From the 3rd column on, the dataset must contain numerical values that represent the signal corresponding to the markers abundance in each sample (marker-related columns).
- Marker-related columns can be called 'Marker1, Marker2, Marker3, ...' or can be called directly with the gene/protein name, but "-" is not allowed in the column name. Only if all the checks are passed, it reorders alphabetically the marker-related columns depending on marker names (necessary for a proper computation of combinations), and it forces "Class" as 2nd column name.
Value

A data frame (data.frame) containing a representation of the data in the file.

Examples

demo_data # combiroc built-in demo data (proteomics data from Zingaretti et al. 2012 - PMC3518104)

# save a data.frame as a csv to be load by combiroc package
file= tempfile()
write.csv2(demo_data, file = file, row.names = FALSE)

#To load a csv file if correctly formatted

demo_data <- load_data(data = file, sep = ';', na.strings = '')

load_unclassified_data

Load unclassified data.

Description

A function to load datasets not yet classified. It’s analogue to load_data() since it loads the same data
type and performs the same format checks, with the exception of "Class" column that in unclassified
data is missing.

Usage

load_unclassified_data(data, sep = ";", na.strings = "")

Arguments

data the name of the file which the data are to be read from.
sep the field separator character.
na.strings a character vector of strings which are to be interpreted as NA values.

Details

The unclassified dataset to be loaded should be in text format, which can be comma, tab or semi-
colon separated:

- The 1st column must contain unique patient/sample IDs.
- From the 2nd column on, the dataset must contain numerical values that represent the signal
corresponding to the markers abundance in each sample (marker-related columns).
- Marker-related columns must be called with the same name of the dataset previously loaded
with load_data(). Only if all the checks are passed, it reorders alphabetically the marker-
related columns depending on marker names (necessary for a proper computation of combi-
nations), and it forces "Class" as 2nd column name.
Value

a data frame (data.frame) containing a representation of the data in the file.

Examples

demo_unclassified_data # combiroc built-in unclassified demo data

# save a data.frame as a csv to be load by combiroc package
file= tempfile()
write.csv2(demo_unclassified_data, file = file, row.names = FALSE)

# To load an unclassified dataset.
demo_unclassified_data <- load_unclassified_data(data= file ,
sep = ";", na.strings="" )

markers_distribution  Show distribution of intensity values for all the markers both singularly and all together.

Description

A function that takes as input data in long format, and shows how the signal intensity value of markers are distributed.

Usage

markers_distribution(  
data_long,  
min_SE = 40,  
min_SP = 80,  
x_lim = NULL,  
y_lim = NULL,  
boxplot_lim = NULL,  
signalthr_prediction = FALSE,  
case_class  
)

Arguments

data_long a data.frame in long format returned by combiroc_long()
min_SE a numeric that specifies the min value of SE that a threshold must have to be included in $Coord.
min_SP a numeric that specifies the min value of SP that a threshold must have to be included in $Coord.
x_lim a numeric setting the max values of x that will be visualized in the density plot (zoom only, no data loss).
markers_distribution

y_lim          a numeric setting the max values of y that will be visualized in the density plot (zoom only, no data loss).
boxplot_lim    a numeric setting the max values of y that will be visualized in the boxplot (zoom only, no data loss).
signalthr_prediction
                a boolean that specifies if the density plot will also show the "suggested signal threshold".
case_class     a character that specifies which of the two classes of the dataset is the case class.

Details

This function returns a named list containing the following objects:

- "Density_plot": a density plot showing the distribution of the signal intensity values for both the classes.
- "Density_summary": a data.frame showing a summary statistics of the distributions.
- "ROC": a ROC curve showing how many real positive samples would be found positive (SE) and how many real negative samples would be found negative (SP) in function of signal threshold. NB: these SE and SP are refereed to the signal intensity threshold considering all the markers together; it is NOT equal to the SE/SP of a single marker/combination found with se_sp().
- "Coord": a data.frame that contains the coordinates of the above described "ROC" (threshold, SP and SE) that have at least a min SE (40 by default) and a min SP (80 by default).
- "Boxplot": a boxplot showing the distribution of the signal intensity values of each marker singularly, for both the classes.

In case of lack of a priori known threshold the user can set set signalthr_prediction= TRUE. In this way the function provides a "suggested signal threshold" that corresponds to the median of the singnal threshold values (in "Coord") at which SE/SP are grater or equal to their set minimal values (min_SE and min_SP), and it adds this threshold on the "Density_plot" object as a dashed black line. The use of the median allows to pick a threshold whose SE/SP are not too close to the limits (min_SE and min_SP), but it is recommended to always inspect "Coord" and choose the most appropriate signal threshold by considering SP, SE and Youden index.

Value

a named list containing 'Coord' and 'Density_summary' data.frames, and 'ROC', 'Boxplot' and 'Density_plot' plot objects.

Examples

demo_data # combiroc built-in demo data (proteomics data from Zingaretti et al. 2012 - PMC3518104)
demo_data_long <- combiroc_long(data = demo_data) # long format data
# To visualize the distribution of the expression of each marker.

distributions <- markers_distribution(data_long = demo_data_long, 
  boxplot_lim = 1500, y_lim = 0.001, 
  x_lim = 3000, signalthr_prediction = FALSE, 
  case_class = 'A', min_SE = 40, min_SP = 80)

distributions$Density_plot # density plot

distributions$Density_summary # summary statistics of density plot

distributions$ROC # ROC showing signal threshold range ensuring min SE and/or SP

distributions$Coord # ROC values

distributions$Boxplot # Boxplot

ranked_combs

**Rank combinations.**

**Description**

A function to rank combinations by a Youden index and select them if they have a min SE and/or SP.

**Usage**

ranked_combs(data, combo_table, case_class, min_SE = 0, min_SP = 0)

**Arguments**

- **data** a data.frame returned by load_data().
- **combo_table** a data.frame with SE, SP and number of composing markers for each combination (returned by se_sp()).
- **case_class** a character that specifies which of the two classes of the dataset is the case class.
- **min_SE** a numeric that specifies the min value of SE that a combination must have to be filtered-in.
- **min_SP** a numeric that specifies the min value of SP that a combination must have to be filtered-in.

**Details**

This function is meant to help the user in finding the best combinations (in the first rows) and allows also (not mandatory) the SE/SP-dependent filtering of combinations.

**Value**

a named list containing:

- Stable, a data.frame with ranked combination, reporting: SE, SP, number of markers composing the combination and the score.
- $bubble_chart, a dot plot showing the selected 'gold' combinations
roc_reports

Examples

demo_data # combiroc built-in demo data (proteomics data from Zingaretti et al. 2012 - PMC3518104)
combs <- combi(data= demo_data, signalthr=450, combithr=1) # compute combinations
combs_SE_SP <- se_sp(data=demo_data, combinations_table=combs) # compute SE and SP # of each combination

# To rank combinations by Youden index and filter-out the ones with SE < min_SE and SP < min_SP
rc <- ranked_combs(data= demo_data, combo_table= combs_SE_SP,
                    case_class='A', min_SE=40, min_SP=80)
rc$table # to visualize the selected gold combinations through a data.frame
rc$bubble_chart # to visualize the selected gold combinations through a data.frame

roc_reports

Train logistic regression and compute ROC.

Description

A function to compute General Linear Model (binomial) and the corresponding ROC curves for each selected combination.

Usage

roc_reports(
  data,
  markers_table,
  selected_combinations = NULL,
  single_markers = NULL,
  case_class
)

Arguments

data a data.frame returned by load_data().
markers_table a data.frame with combinations and corresponding positive samples counts, obtained with combi().
selected_combinations a numeric vector that specifies the combinations of interest.
single_markers a character vector that specifies the single markers of interest.
case_class a character that specifies which of the two classes of the dataset is the case class.
Details

This function trains a logistic regression model for each combination and returns a named list containing 3 objects:

- "Plot": a ggplot object with the ROC curves of the selected combinations.
- "Metrics": a data.frame with the metrics of the roc curves (AUC, opt. cutoff, etc ...).
- "Models": the list of models (glm() objects) that have been computed and then used to classify the samples (in which you can find the model equation for each selected combination).

Value

a named list containing 3 objects: "Plot", "Metrics" and "Models".

Examples

demo_data # combiroc built-in demo data (proteomics data from Zingaretti et al. 2012 - PMC3518104)
combs <- combi(data= demo_data, signalthr=450, combithr=1) # compute combinations

# To train logistic regression models on each selected combinations and each selected marker, and compute corresponding ROCs.
reports <- roc_reports(data= demo_data, markers_table= combs,
                        selected_combinations= c(1,11,15),
                        single_markers=c('Marker1', 'Marker2'), case_class='A')

reports$Plot # Shows the ROC curves
reports$Metrics # Shows the ROC metrics
reports$Models # show models
reports$reports$Models$`Combination 11` # show model trained with Combination 11

se_sp (Compute sensitivity and specificity of each combination)

Description

A function to compute sensitivity and specificity of each combination for each class.

Usage

se_sp(data, combinations_table)

Arguments

data a data.frame returned by load_data().
combinations_table a data.frame containing how many samples of each class are "positive" for each combination (returned by combi()).
show_markers

Details

This function calculate SE and SP for each combination. The SE of a given combination (capability to find real positives/cases) corresponds to the SE of the case class, while its SP (capability to exclude real negatives/controls) corresponds to the SP of the control class.

Value

data.frame with SE, SP and number of composing markers for each combination.

Examples

demo_data # combiroc built-in demo data (proteomics data from Zingaretti et al. 2012 - PMC3518104)
combs <- combi(data= demo_data, signalthr=450, combithr=1) # compute combinations

# To compute sensitivity and specificity of each combination
combs_SE_SP <- se_sp(data=demo_data, combinations_table=combs)

show_markers

Description

A function to show the composition of combinations of interest.

Usage

show_markers(markers_table, selected_combinations)

Arguments

  markers_table  a data.frame with combinations returned by combi().
  selected_combinations  a numeric vector that specifies the combinations of interest.

Value

a data.frame containing the selected combinations and their composing markers.

Examples

demo_data # combiroc built-in demo data (proteomics data from Zingaretti et al. 2012 - PMC3518104)
combs <- combi(data= demo_data, signalthr=450, combithr=1) # compute combinations

# To show the composition of combinations of interest.
show_markers(markers_table = combs, selected_combinations = c(1,11))
single_markers_statistics

*Provide statistics for each marker.*

**Description**

A function that computes the statistics and a scatter-plot for each marker.

**Usage**

`single_markers_statistics(data_long)`

**Arguments**

- `data_long` a data.frame in long format returned by `combiroc_long()`.

**Details**

This function computes the main statistics of the signal values distribution of each marker in both classes. In addition it also shows the values through scatter plots.

**Value**

a list object containing:

- 'Statistics': a dataframe containing the main statistics for each marker in each class.
- 'Plots': a named list of scatter plots showing signal intensity values.

**Examples**

demo_data # combiroc built-in demo data (proteomics data from Zingaretti et al. 2012 – PMC3518104)

data_long <- combiroc_long(demo_data) # reshape data in long format
 sms <- single_markers_statistics(data_long)

sms$Statistics # to visualize the statistics of each single marker
sms$Plots[[1]] # to visualize the scatterplot of the first marker
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