

# Package ‘VGAMdata’

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**Title** Data Supporting the 'VGAM' Package

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**Depends** R (>= 3.5.0), methods, stats, VGAM

**Description** Mainly data sets to accompany the VGAM package and the book “Vector Generalized Linear and Additive Models: With an Implementation in R” (Yee, 2015) <[DOI:10.1007/978-1-4939-2818-7](https://doi.org/10.1007/978-1-4939-2818-7)>. These are used to illustrate vector generalized linear and additive models (VGLMs/VGAMs), and associated models (Reduced-Rank VGLMs, Quadratic RR-VGLMs, Row-Column Interaction Models, and constrained and unconstrained ordination models in ecology). This package now contains some old VGAM family functions which have been replaced by newer ones (often because they are now special cases).

**License** GPL-2

**URL** <https://www.stat.auckland.ac.nz/~yee/VGAMdata/>

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**LazyData** yes

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VGAMdata-package	<i>Mainly Data for the VGAM package</i>
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## Description

**VGAMdata** is mainly an assortment of larger data sets which are a useful resource for the **VGAM** package.

## Details

This package mainly contains some larger data sets originally distributed with the **VGAM** package. Ideally both packages can be installed and loaded to be fully functional. The main intent was to limit the size of **VGAM** to a bare essential. Many data sets in my monograph will refer to data sets in either package. Recently, some older or less-used **VGAM** family functions have been shifted into **VGAMdata**, and this is likely to continue in the future.

## Author(s)

Thomas W. Yee, <t.yee@auckland.ac.nz>.

Maintainer: Thomas Yee <t.yee@auckland.ac.nz>.

## References

Yee, T. W. (2015). *Vector Generalized Linear and Additive Models: With an Implementation in R*. New York, USA: *Springer*.

## See Also

[VGAM-package](#).

## Examples

```
# Example 1; xs.nz
head(xs.nz)
summary(xs.nz)

# Example 2; ugss
head(ugss)
summary(ugss)
```

---

airbnb.ac

*Airbnb Accommodation in Two Sardinian Cities*

---

## Description

The `airbnb.ac` data frame has 18159 rows and 9 columns.

## Usage

```
data(airbnb.ac)
```

## Format

This data frame contains the following columns:

**LOS** length of stay, in days.

**discountWeek** Multiplicative factor for the discount for booking one week:  $1 - (\text{published weekly rate}) / (7 \times \text{published nightly rate})$ , e.g., 0.2 means a 20 percent savings off the regular price.

**NumberOfReviews** Number of reviews on the website.

**PriceAvg** Average price per night per person, in Euros.

**Bedrooms** Number of bedrooms.

**Superhost** Logical. Superhost?

**MinimumStay** Minimum stay period, in days.

**MaxGuests** Maximum number of guests.

**City** Character, values are "Alghero" and "Cagliari".

## Details

The data frame comprises Airbnb bookings in two cities located in Sardinia, Italy. The stays were during the whole of 2016. Stays of 30 days or longer and any rows with missing variables were deleted from the original source. Variable LOS exhibits heaping at the values 7 and 14 days.

## Source

The data was obtained with permission from Luca Frigau, University of Cagliari, from <https://www.airbnb.com>.

**See Also**

[gaitdzeta](#), [flamingo](#).

**Examples**

```
## Not run:
mytab <- with(subset(airbnb.ac, City == "Alghero"), table(LOS))
plot(prop.table(mytab), col = "blue", ylab = "Proportion")

## End(Not run)
```

---

bb.de

*Battle of Britain Data (a Luftwaffe subset)*

---

**Description**

Luftwaffe losses during a subset of the Battle of Britain.

**Usage**

```
data(bb.de)
```

**Format**

The format is a 3-dimensional array. The first dimension is the event (in order: shot down or failed to return, written off, seriously damaged), the second dimension is the day, the third is the aircraft type.

**Details**

This is a Battle of Britain data set of Luftwaffe losses during operations 26 August–31 August 1940 continued on to 1–7 September 1940. The aircraft types are prefixed Bf for Messerschmitt (Bayerische Flugzeugwerke), Do for Dornier, He for Heinkel, Ju for Junkers.

Note that p.151 and p.165 of Bowyer (1990) contain tables (during the first week of September) and almost the same data; however, the former is labelled "shot down" whereas the latter is "shot down or failed to return". The latter is used here. Also, there are some other small discrepancies.

Yet to do: add the data available at other dates, and include the RAF data.

**Source**

Bowyer, M. J. F. (1990) *The Battle of Britain: 50 years On*. Patrick Stephens Limited, Northamptonshire, U.K.

**Examples**

```

data(bb.de)
bb.de[, , "Bf109"]

## Not run:
plot(bb.de["sdown", , "Bf109"] ~ as.Date(dimnames(bb.de)[[2]]),
     type = "h", col = "blue", las = 1, lwd = 3,
     ylab = "Frequency", xlab = "1940",
     main = "Numbers shot down (Bf 109)")
abline(h = c(5, 10, 15, 20), lty = "dashed", col = "grey")
points(bb.de["sdown", , "Bf109"] ~ as.Date(dimnames(bb.de)[[2]]), col = "blue")

## End(Not run)

```

---

bd.us

*Births and Deaths of 348 Notable Americans*


---

**Description**

A 12 x 12 table of the Births and Deaths of 348 Notable Americans. The rows and columns are for each month.

**Usage**

```
data(bd.us)
```

**Format**

The format is: chr "bd.us"

**Details**

Rows denote the month of birth; columns for the month of death. These data appear as Table 1 of Phillips and Feldman (1973), who collected the data from Morris (1965). Not all of the 400 people were used because some had not died by that time and other individuals lacked the information.

**Source**

Phillips, D. P. and Feldman, K. A. (1973) A dip in deaths before ceremonial occasions: Some new relationships between social integration and mortality, *American Sociological Review*, **38**, 678–696.

Morris, R. B. (Ed.) (1965) *Four Hundred Notable Americans*. Harper and Row: New York, USA.

**See Also**

[rcim](#).

**Examples**

```
print(bd.us)
sum(bd.us)
rowSums(bd.us)
colSums(bd.us)
```

---

belcap

*BELCAP Dental Data*

---

**Description**

A prospective data set containing the DMFT index of children in Belo Horizonte at the beginning and end of the BELCAP study.

**Usage**

```
data(belcap)
```

**Format**

A data frame with 797 observations on the following 5 variables.

`dmftb` a numeric vector, DMFT-Index at the beginning of the study.

`dmfte` a numeric vector, DMFT-Index at the end of the study.

`gender` a factor with levels 0 = female, 1 = male.

`ethnic` a factor with levels 1 = dark, 2 = white, 3 = black.

`school` the kind of prevention measure. A factor with levels 1 = oral health education, 2 = all four methods, 3 = control group, 4 = enrichment of the school diet with ricebran, 5 = mouthrinse with 0.2% NaF-solution, 6 = oral hygiene.

**Details**

This data set is from the Belo Horizonte Caries Prevention (BELCAP) study. The data is collected from children in Belo Horizonte (Brazil) aged seven years at the start of the study. In order to determine which method(s) were best for preventing tooth decay, six treatments were randomized to six separate schools. The measure used is the decayed, missing and filled teeth (DMFT) index - a well known and important measure of a persons dental health. Only the eight deciduous molars are considered, so the lowest value is 0, and the highest is 8.

**Source**

<https://onlinelibrary.wiley.com/> contains the data file (a supplement of the JRSSA article). Downloaded in January 2014 and formatted into R by J. T. Gray, [jamsgr@gmail.com](mailto:jamsgr@gmail.com).

## References

Bohning, D., D. Ekkehart, P. Schlattmann, L. Mendonca, and U. Kircher (1999). The Zero-Inflated Poisson Model and the Decayed, Missing and Filled Teeth Index in Dental Epidemiology, *Journal of the Royal Statistical Society, A* **162**(2), 195–209.

## Examples

```
data(belcap)
## maybe str(belcap) ; plot(belcap) ...
```

---

Bell

*The Bell Distribution*

---

## Description

Density, and random generation for the Topp-Leone distribution.

## Usage

```
dbell(x, shape, log = FALSE)
rbell(n, shape)
```

## Arguments

x, n	Same as <a href="#">Uniform</a> .
shape	the (shape) parameter, which is positive.
log	Logical. If log = TRUE then the logarithm of the density is returned.

## Details

See [bellff](#), the **VGAM** family function for estimating the parameter  $s$  by maximum likelihood estimation.

## Value

dbell gives the density, rbell generates random deviates. If shape is large then rbell will become computationally expensive.

## See Also

[bell](#).

## Examples

```
## Not run: plot(0:15, dbell(0:15, shape = 1.5), type = "h", col = "blue")
```

bellff

*Bell Distribution Family Function***Description**

Estimating the shape parameter of the Bell distribution by maximum likelihood estimation.

**Usage**

```
bellff(lshape = "loglink", zero = NULL, gshape = expm1(1.6 * ppoints(7)))
```

**Arguments**

lshape, zero, gshape

More information is at [CommonVGAMffArguments](#).

**Details**

The Bell distribution has a probability density function that can be written

$$f(y; s) = \frac{s^y \exp(1 - e^s) B_y}{y!}$$

for  $y = 0(1)\infty$  and shape parameter  $0 < s$ . The mean of  $Y$  is  $\exp(s)s$  (returned as the fitted values). Fisher-scoring is used. This **VGAM** family function handles multiple responses.

The function `bell` returns the first 218 Bell numbers as finite numbers, and returns `Inf` when its argument has a higher value. Hence this **VGAM** family function can only handle low-value counts of less than 219.

**Value**

An object of class "vglmff" (see [vglmff-class](#)). The object is used by modelling functions such as `vglm`, and `vgam`.

**Author(s)**

T. W. Yee

**References**

Castellares, F. and Ferrari, S. L. P. and Lemonte, A. J. (2018). On the Bell distribution and its associated regression model for count data. *Applied Mathematical Modelling*, **56**, 172–185.

**See Also**

[bell](#), [dbell](#), [poissonff](#).

## Examples

```
bdata <- data.frame(y = rbell(1000, shape = loglink(0.5, inverse = TRUE)))
bfit <- vglm(y ~ 1, bellff, data = bdata, trace = TRUE, crit = "coef")
coef(bfit, matrix = TRUE)
Coef(bfit)
```

---

covid19.nz

*COVID-19 in New Zealand: The First Month or So*

---

## Description

The covid19.nz data frame has 69 rows and 3 columns. The number of new cases is tracked daily.

## Usage

```
data(covid19.nz)
```

## Format

This data frame contains the following columns:

**doy** output from `as.Date`, is the day of year.

**newcases** a numeric vector, counts, the number of new cases.

**Day** a numeric vector, 0 for when the first case occurred; incrementally daily after that.

## Details

These were collected from <https://www.nzherald.co.nz/> during the first month or so after the first case.

## Source

The NZ Herald states their source was Johns Hopkins University.

## Examples

```
## Not run: plot(newcases ~ doym, covid19.nz, col = "blue", type = "h",
  las = 1, xlab = "Date")

## End(Not run)
```

---

`crashf.au`*Fatal Crashes on Australian Roads 2010–2012*

---

**Description**

The number of fatal road crashes on Australian roads during 2010–2012. They are cross-classified by time of day (in 6 hour blocks) and day of the week.

**Usage**

```
data(crashf.au)
```

**Format**

A data frame with 4 observations on the following 7 variables.

**Mon, Tue, Wed, Thu, Fri, Sat, Sun** Day of the week.

**Details**

Each cell is the aggregate number of crashes reported in Australia during each six hour time block throughout the years 2010–2012. The rownames are the time period the crashes took place in. Morning is from 3:00am to 8:59am, midday is from 9:00am to 2:59pm, evening is from 3:00pm to 8:59pm and night is from 9:00pm to 2:59am.

**Source**

[http://www.bitre.gov.au/publications/ongoing/files/RDA\\_Summary\\_2012\\_June.pdf](http://www.bitre.gov.au/publications/ongoing/files/RDA_Summary_2012_June.pdf)

**References**

Road Deaths Australia; 2012 Statistical Summary. Department of Infrastructure and Transport, Australian Government; ISSN: 1323–3688

Downloaded by J. T. Gray, April 2014.

**Examples**

```
crashf.au
```

---

`crime.us`*Estimated Crime in 2009 in USA*

---

**Description**

Crime totals and rates, cross-classified by US state, during 2009.

**Usage**

```
data(crime.us)
```

**Format**

A data frame with 50 observations on the following 22 variables.

`State` a character vector. White spaces have been replaced by underscores.

`Population` a numeric vector

`ViolentCrimeTotal` a numeric vector

`Murder` a numeric vector

`Rape` a numeric vector

`Robbery` a numeric vector

`Assault` a numeric vector

`PropertyCrimeTotal` a numeric vector

`Burglary` a numeric vector

`LarcenyTheft` a numeric vector

`MotorVehicleTheft` a numeric vector

`ViolentCrimeRate` a numeric vector

`MurderRate` a numeric vector

`RapeRate` a numeric vector

`RobberyRate` a numeric vector

`AssaultRate` a numeric vector

`PropertyCrimeRate` a numeric vector

`BurglaryRate` a numeric vector

`LarcenyTheftRate` a numeric vector

`MotorVehicleTheftRate` a numeric vector

`stateNumber` a numeric vector, running from 1 to 50.

`abbrev` State name as a character vector

## Details

Each row is a state of the United States of America. The first half of the columns tend to be totals, and the second half are crime rates per 100,000 population.

The data frame was downloaded as a .csv file and edited. The full column names are: State, Population, Violent crime total, Murder and nonnegligent Manslaughter, Forcible rape, Robbery, Aggravated assault, Property crime total, Burglary, Larceny-theft, Motor vehicle theft, Violent Crime rate, Murder and nonnegligent manslaughter rate, Forcible rape rate, Robbery rate, Aggravated assault rate, Property crime rate, Burglary rate, Larceny-theft rate, Motor vehicle theft rate, state Number, abbreviation. Technical details governing the data set are given in the URL.

## Source

<http://www.ucrdatatool.gov>, <http://www.ucrdatatool.gov/Search/Crime/State/RunCrimeOneYearofData.cfm>

## Examples

```
## Not run: # Louisiana is the one outlier
plot(MurderRate ~ stateNumber, crime.us,
     axes = FALSE, type = "h", col = 1:6,
     main = "USA murder rates in 2009 (per 100,000 population)")
axis(1, with(crime.us, abbrev), at = with(crime.us, stateNumber),
     col = 1:6, col.tick = 1:6, cex.lab = 0.5)
axis(2)
## End(Not run)
tail(crime.us[ sort.list(with(crime.us, MurderRate)), ])
```

---

DeLury

*DeLury's Method for Population Size Estimation*

---

## Description

Computes DeLury's method or Leslie's method for estimating a biological population size.

## Usage

```
DeLury(catch, effort, type = c("DeLury", "Leslie"), ricker = FALSE)
```

## Arguments

catch, effort	Catch and effort. These should be numeric vectors of equal length.
type	Character specifying which of the DeLury or Leslie models is to be fitted. The default is the first value.
ricker	Logical. If TRUE then the Ricker (1975) modification is computed.

## Details

This simple function implements the methods of DeLury (1947). These are called the DeLury and Leslie models. Note that there are many assumptions. These include: (i) Catch and effort records are available for a series of consecutive time intervals. The catch for a given time interval, specified by  $t$ , is  $c(t)$ , and the corresponding effort by  $e(t)$ . The *catch per unit effort* (CPUE) for the time interval  $t$  is  $C(t) = c(t)/e(t)$ . Let  $d(t)$  represent the proportion of the population captured during the time interval  $t$ . Then  $d(t) = k(t)e(t)$  so that  $k(t)$  is the proportion of the population captured during interval  $t$  by one unit of effort. Then  $k(t)$  is called the *catchability*, and the *intensity* of effort is  $e(t)$ . Let  $E(t)$  and  $K(t)$  be the total effort and total catch up to interval  $t$ , and  $N(t)$  be the number of individuals in the population at time  $t$ . It is good idea to plot  $\log(C(t))$  against  $E(t)$  for type = "DeLury" and  $C(t)$  versus  $K(t)$  for type = "Leslie".

The other assumptions are as follows.

(ii) The population is closed—the population must be closed to sources of animals such as recruitment and immigration and losses of animals due to natural mortality and emigration.

(iii) Catchability is constant over the period of removals.

(iv) The units of effort are independent, i.e., the individual units of the method of capture (i.e., nets, traps, etc) do not compete with each other.

(v) All fish are equally vulnerable to the method of capture—source of error may include gear saturation and trap-happy or trap-shy individuals.

(vi) Enough fish must be removed to substantially reduce the CPUE.

(vii) The catches may remove less than 2% of the population.

Also, the usual assumptions of simple regression such as

(viii) random sampling,

(ix) the independent variable(s) are measured without error—both catches and effort should be known, not estimated,

(x) a line describes the data,

(xi) the errors are independent and normally distributed.

## Value

A list with the following components.

catch, effort	Catch and effort. Same as the original vectors. These correspond to $c(t)$ and $e(t)$ respectively.
type, ricker	Same as input.
N0	an estimate of the population size at time 0. Only valid if the assumptions are satisfied.
CPUE	Catch Per Unit Effort = $C(t)$ .
K, E	$K(t)$ and $E(t)$ . Only one is computed depending on type.
lmfit	the <code>lm</code> object from the fit of $\log(\text{CPUE})$ on $K$ (when type = "Leslie"). Note that the <code>x</code> component of the object is the model matrix.

**Note**

The data in the example below comes from DeLury (1947), and some plots of his are reproduced. Note that he used log to base 10 whereas natural logs are used here. His plots had some observations obscured by the y-axis!

The DeLury method is not applicable to the data frame [wffc.nc](http://wffc.nc) since the 2008 World Fly Fishing Competition was strictly catch-and-release.

**Author(s)**

T. W. Yee.

**References**

- DeLury, D. B. (1947). On the estimation of biological populations. *Biometrics*, **3**, 145–167.
- Ricker, W. E. (1975). Computation and interpretation of biological statistics of fish populations. *Bull. Fish. Res. Bd. Can.*, **191**, 382–
- Yee, T. W. (2010) VGLMs and VGAMs: an overview for applications in fisheries research. *Fisheries Research*, **101**, 116–126.

**See Also**

[wffc.nc](http://wffc.nc).

**Examples**

```
pounds <- c( 147, 2796, 6888, 7723, 5330, 8839, 6324, 3569, 8120, 8084,
            8252, 8411, 6757, 1152, 1500, 11945, 6995, 5851, 3221, 6345,
            3035, 6271, 5567, 3017, 4559, 4721, 3613, 473, 928, 2784,
            2375, 2640, 3569)
traps <- c( 200, 3780, 7174, 8850, 5793, 9504, 6655, 3685, 8202, 8585,
           9105, 9069, 7920, 1215, 1471, 11597, 8470, 7770, 3430, 7970,
           4740, 8144, 7965, 5198, 7115, 8585, 6935, 1060, 2070, 5725,
           5235, 5480, 8300)
table1 <- DeLury(pounds/1000, traps/1000)

## Not run:
with(table1, plot(1+log(CPUE) ~ E, las = 1, pch = 19, main = "DeLury method",
                xlab = "E(t)", ylab = "1 + log(C(t))", col = "blue"))

## End(Not run)
omitIndices <- -(1:16)
table1b <- DeLury(pounds[omitIndices]/1000, traps[omitIndices]/1000)
## Not run:
with(table1b, plot(1+log(CPUE) ~ E, las = 1, pch = 19, main = "DeLury method",
                 xlab = "E(t)", ylab = "1 + log(C(t))", col = "blue"))
mylmfit <- with(table1b, lmfit)
lines(mylmfit$x[, 2], 1 + predict.lm(mylmfit), col = "red", lty = "dashed")

## End(Not run)
```

```

omitIndices <- -(1:16)
table2 <- DeLury(pounds[omitIndices]/1000, traps[omitIndices]/1000, type = "L")
## Not run:
with(table2, plot(CPUE ~ K, las = 1, pch = 19,
  main = "Leslie method; Fig. III",
  xlab = "K(t)", ylab = "C(t)", col = "blue"))
mylmfit <- with(table2, lmfit)
abline(a = coef(mylmfit)[1], b = coef(mylmfit)[2],
  col = "orange", lty = "dashed")

## End(Not run)

```

---

ecb06.it

*Italian Household Data for 2006 and 2014*


---

## Description

Part of the data collected at two time points (2006 and 2014) by the Bank of Italy, as part of the European Central Banks Eurosystem collection of statistics, within the periodic sample surveys on households, businesses and selected intermediaries.

## Format

Data frame with the following 20 variables:

**ID** a numeric vector, a unique identification number of the household.

**area** a factor with 5 levels, the Italian geographic area or region in which the household lives: NW = North-West, NE = North-East, C = Center, S = South, I = Islands. For users wanting a North-South contrast, this variable might be coded as NC = North and Center (NW, NE and C), SI = South and Islands (S and I).

**sex** a factor with 2 levels, the gender of the head householder: M = Male, F = Female.

**age** a numeric vector, age in years of the head householder.

**marital** a factor with 4 levels, marital status of the head householder: married = Married, single = Single, separated = Separated or divorced, widowed = Widowed.

**education** an ordered factor with 8 levels, the education level of the head householder: none = No education, primaryschool = Primary school, middleschool = Middle school, profschool = Professional school, highschool = High school, bachelors = Bachelors degree, masters = Masters degree, higherdegree = Higher degree.

**job** a factor with 7 levels, the type of job done by the head householder: worker = Worker, employee = Employee, manager = Manager, business = Business person, other = Other kind of independent job, retired = Retired, unemployed = Unemployed.

**occupants** a numeric vector, the number of people living in the same house.

**children** a numeric vector, the number of children of the head householder living with him/her.

**other.children** a numeric vector, the number of children of the head householder not living with the household.

house.owned a numeric vector, the ownership of the house in which the householder lives; 0 = The house is not owned, 1 = The house is owned.

houses a numeric vector, the number of houses owned by the family, including the one in which the family lives.

earners a numeric vector, the number of people in the house who have salary or some kind of earnings.

accounts a numeric vector, the number of bank accounts collectively owned by the household.

ccards a numeric vector, the number of credit cards collectively owned by the household.

tot.income, dep.income, pens.income, self.income, cap.income numeric vectors, the amount of income (in Euros) collectively earned by the household through different activities. The variables can be negative if the household has installments. In order, they are the total amount of income, the amount of income earned through dependent working, the amount of income earned by the household through pensions, the amount of income earned by the household through self-dependent working, the amount of income earned by the household through capital investments.

## Details

The European Central Banks (ECB) Eurosystem requests and helps organize each country within the European Union to routinely collect statistical information via their central banks. These data frames are a subset from data collected by the Bank of Italy. Each year can be considered a cross-sectional study, although there are some people common to each year. Hence the data collectively can be considered partly a longitudinal study too.

## Source

Data was downloaded at <https://www.bancaditalia.it> in May 2016 by Lucia Pilleri.

## References

*Supplements to the Statistical Bulletin, Sample Surveys, Household Income and Wealth in 2006*, New series, Volume XVIII, Number 7–28, January 2008. Banca D'Italia, Centro Stampa, Roma, Pubbl. Mensile, <https://www.bancaditalia.it>.

## Examples

```
data(ecb06.it); data(ecb14.it)
summary(ecb06.it)
summary(ecb14.it)
## Not run:
with(ecb14.it, table(house.owned))
with(ecb14.it, barplot(table(education), col = "lightblue"))

## End(Not run)
```

exam1

*Examination data***Description**

Exam results of 35 students on 18 questions.

**Usage**

```
data(exam1)
```

**Format**

A data frame with 35 observations on the following 18 variables.

**q01, q02, q03, q04, q05, q06** binary response

**q07, q08, q09, q10, q11, q12** binary response

**q13, q14, q15, q16, q17, q18** binary response

**Details**

For each question, a 1 means correct, a 0 means incorrect. A simple Rasch model may be fitted to this dataframe using `rcim` and `binomialff`.

**Source**

Taken from William Revelle's *Short Guide to R*, [http://www.unt.edu/rss/rasch\\_models.htm](http://www.unt.edu/rss/rasch_models.htm), <http://www.personality-project.org/r/>. Downloaded in October 2013.

**Examples**

```
summary(exam1) # The names of the students are the row names

# Fit a simple Rasch model.
# First, remove all questions and people who were totally correct or wrong
exam1.1 <- exam1 [, colMeans(exam1 ) > 0]
exam1.1 <- exam1.1[, colMeans(exam1.1) < 1]
exam1.1 <- exam1.1[rowMeans(exam1.1) > 0, ]
exam1.1 <- exam1.1[rowMeans(exam1.1) < 1, ]
Y.matrix <- rdata <- exam1.1

## Not run: # The following needs: library(VGAM)
rfit <- rcim(Y.matrix, family = binomialff(multiple.responses = TRUE),
            trace = TRUE)

coef(rfit) # Row and column effects
constraints(rfit, matrix = TRUE) # Constraint matrices side-by-side
dim(model.matrix(rfit, type = "vlm")) # 'Big' VLM matrix
```

```
## End(Not run)

## Not run: # This plot shows the (main) row and column effects
par(mfrow = c(1, 2), las = 1, mar = c(4.5, 4.4, 2, 0.9) + 0.1)
saved <- plot(rfit, rcol = "blue", ccol = "orange",
             cylab = "Item effects", rylab = "Person effects",
             rxlab = "", cxlab = "")

names(saved@post) # Some useful output put here
cbind(saved@post$row.effects)
cbind(saved@post$raw.row.effects)
round(cbind(-saved@post$col.effects), dig = 3)
round(cbind(-saved@post$raw.col.effects), dig = 3)
round(matrix(-saved@post$raw.col.effects, ncol = 1, # Rename for humans
            dimnames = list(colnames(Y.matrix), NULL)), dig = 3)

## End(Not run)
```

---

flamingo

*Flamingo Hotel in Sardinia*


---

## Description

The flamingo data frame has 4871 rows and 12 columns.

## Usage

```
data(flamingo)
```

## Format

This data frame contains the following columns:

**LOS** length of stay, in days.

**year** year of the stay.

**arrdate** date of arrival.

**deptime** date of departure.

**seasonindex** seasonality index of the period of the stay, a low value means low season, a high value means very high (peak) season.

**booking** if the guests booked the room through the hotel's internet website (`internet`), the hotel's telephone (`direct`) or a tour operator (`agency`).

**roomtype** the 16 hotel's room types.

**rmtpe3** categorization in 3 groups of roomtype.

**guests** number of guests. Does not include any children; see the two `zz kids` variables below.

**arrangement** arrangement type: Bed and Breakfast (BB), Half Board (HB) and Full Board (FB).

**kids02** number of kids between 0 and 2 years-old.

**kids311** number of kids between 3 and 11 years-old.

**Details**

The Flamingo Hotel is located on the beach in the southern Sardinia, about 40 kilometers from Cagliari. This data concerns stays from early summer 2019 to late summer 2020 with no stays during the winter period in between. Stays longer than 30 days were deleted from the original source. Variable LOS exhibits heaping at the values 7 and 14 days.

**Source**

The data was obtained with permission from Luca Frigau, University of Cagliari.

**See Also**

[gaitdzeta](#), [airbnb.ac](#).

**Examples**

```
## Not run: with(flamingo, spikeplot(LOS, col = "blue", ylab = "Proportion"))
```

---

genpoisson

*Generalized Poisson Regression*

---

**Description**

Estimation of the two-parameter generalized Poisson distribution.

**Usage**

```
genpoisson(llambda = "rhobitlink", ltheta = "loglink",
            ilambda = NULL, itheta = NULL, imethod = 1,
            ishrinkage = 0.95, zero = "lambda")
```

**Arguments**

llambda, ltheta

Parameter link functions for  $\lambda$  and  $\theta$ . See [Links](#) for more choices. The  $\lambda$  parameter lies at least within the interval  $[-1, 1]$ ; see below for more details, and an alternative link is [rhobitlink](#). The  $\theta$  parameter is positive, therefore the default is the log link.

ilambda, itheta

Optional initial values for  $\lambda$  and  $\theta$ . The default is to choose values internally.

imethod

An integer with value 1 or 2 or 3 which specifies the initialization method for the parameters. If failure to converge occurs try another value and/or else specify a value for ilambda and/or itheta.

ishrinkage, zero

See [CommonVGAMffArguments](#) for information.

## Details

This family function is *not* recommended for use; instead try [genpoisson1](#) or [genpoisson2](#). For underdispersion with respect to the Poisson try the GTE (generally-truncated expansion) method described by Yee and Ma (2020).

The generalized Poisson distribution has density

$$f(y) = \theta(\theta + \lambda y)^{y-1} \exp(-\theta - \lambda y)/y!$$

for  $\theta > 0$  and  $y = 0, 1, 2, \dots$ . Now  $\max(-1, -\theta/m) \leq \lambda \leq 1$  where  $m(\geq 4)$  is the greatest positive integer satisfying  $\theta + m\lambda > 0$  when  $\lambda < 0$  [and then  $P(Y = y) = 0$  for  $y > m$ ]. Note the complicated support for this distribution means, for some data sets, the default link for `llambda` will not always work, and some tinkering may be required to get it running.

As Consul and Famoye (2006) state on p.165, the lower limits on  $\lambda$  and  $m \geq 4$  are imposed to ensure that there are at least 5 classes with nonzero probability when  $\lambda$  is negative.

An ordinary Poisson distribution corresponds to  $\lambda = 0$ . The mean (returned as the fitted values) is  $E(Y) = \theta/(1 - \lambda)$  and the variance is  $\theta/(1 - \lambda)^3$ .

For more information see Consul and Famoye (2006) for a summary and Consul (1989) for full details.

## Value

An object of class "vglmff" (see [vglmff-class](#)). The object is used by modelling functions such as [vglm](#), and [vgam](#).

## Warning

Monitor convergence! This family function is fragile. Don't get confused because theta (and not lambda) here really matches more closely with lambda of [dpois](#).

## Note

This family function handles multiple responses. This distribution is potentially useful for dispersion modelling. Convergence problems may occur when `lambda` is very close to 0 or 1. If a failure occurs then you might want to try something like `llambda = extlogitlink(min = -0.9, max = 1)` to handle the LHS complicated constraint, and if that doesn't work, try `llambda = extlogitlink(min = -0.8, max = 1)`, etc.

## Author(s)

T. W. Yee. Easton Huch derived the EIM and it has been implemented in the `weights` slot.

## References

- Consul, P. C. and Famoye, F. (2006). *Lagrangian Probability Distributions*, Boston, USA: Birkhauser.
- Jorgensen, B. (1997). *The Theory of Dispersion Models*. London: Chapman & Hall
- Consul, P. C. (1989). *Generalized Poisson Distributions: Properties and Applications*. New York, USA: Marcel Dekker.
- Yee, T. W. and Ma, C. (2021). Generally-altered, -inflated and -truncated regression, with application to heaped and seeped counts. *Under review*.

**See Also**

[genpoisson1](#), [genpoisson2](#), [poissonff](#), [dpois](#), [dgenpois0](#), [rhobitlink](#), [extlogitlink](#).

**Examples**

```
## Not run:
gdata <- data.frame(x2 = runif(nn <- 500)) # NBD data:
gdata <- transform(gdata, y1 = rnbinom(nn, exp(1), mu = exp(2 - x2)))
fit <- vglm(y1 ~ x2, genpoisson, data = gdata, trace = TRUE)
coef(fit, matrix = TRUE)
summary(fit)
## End(Not run)
```

---

hued

*Harvard University Degrees Conferred by Student Ethnicity*

---

**Description**

A two-way table of counts; there are 7 ethnic groups by 12 degrees.

**Usage**

```
data(hued)
```

**Format**

The format is: chr "hued"

**Details**

The rownames and colnames have been edited. The full names are: Asian/Pacific Islander, Black/Non-Hispanic, Hispanic, International Students, Native American, White/Non-Hispanic, Unknown/Other. The academic year was 2009–2010. GSAS stands for Graduate School of Arts and Sciences. The Other group includes students reported as Two or More Races. See the URL below for more technical details supporting the data.

**Source**

<https://oir.harvard.edu/fact-book>

**See Also**

[huie](#), [huse](#).

**Examples**

```
print(hued)
```

---

huie

*Harvard University International Enrollments*

---

**Description**

A two-way table of counts; there are 12 degrees and 8 areas of the world.

**Usage**

```
data(huie)
```

**Format**

The format is: chr "huie"

**Details**

The rownames and colnames have been edited. The full colnames are: Africa, Asia, Europe, Caribbean and Central and South America, Middle East, North America, Oceania, Stateless.

The data was for the autumn (Fall) of 2010. GSAS stands for Graduate School of Arts and Sciences. See the URL below for more technical details supporting the data.

**Source**

<https://oir.harvard.edu/fact-book>

**See Also**

[hued](#), [huse](#).

**Examples**

```
print(huie)
## maybe str(huie) ; plot(huie) ...
```

---

huse

*Harvard University Numbers of Ladder Faculty by School and Ethnicity*

---

**Description**

A two-way table of counts; there are 14 schools and 5 race/ethnicities.

**Usage**

```
data(huse)
```

**Format**

The format is: chr "huse"

**Details**

Ladder faculty members of Harvard University are cross-classified by their school and their race/ethnicity. This was for the period 2010–1. Ladder Faculty are defined as Assistant Professors or Convertible Instructors, Associate Professors, and Professors that have been appointed in certain Schools.

Abbreviations: FAS = Faculty of Arts and Sciences = Humanities + Social Sciences + Natural Sciences + SEAS, Natural Sciences = Life Sciences + Physical Sciences, SEAS = School of Engineering and Applied Sciences, HBS = Harvard Business School, HMS = Harvard Medical School, HSPH = Harvard School of Public Health, HLS = Harvard Law School, HKS = Harvard Kennedy School, HGSE = Harvard Graduate School of Education, GSD = Graduate School of Design, HDS = Harvard Divinity School, HSDM = Harvard School of Dental Medicine.

See the URL below for many technical details supporting the data. The table was constructed from pp.31–2 from the source.

**Source**

<https://oir.harvard.edu/fact-book>

**References**

*Harvard University Office of the Senior Vice Provost Faculty Development & Diversity: 2010 Annual Report.*

**See Also**

[hued](#), [huie](#).

**Examples**

```
print(huse)
## maybe str(huse) ; plot(huse) ...
```

---

Oalog

*One-Altered Logarithmic Distribution*

---

**Description**

Density, distribution function, quantile function and random generation for the one-altered logarithmic distribution with parameter pobs1.



```
## End(Not run)
```

---

oalog

*One-Altered Logarithmic Distribution*

---

## Description

Fits a one-altered logarithmic distribution based on a conditional model involving a Bernoulli distribution and a 1-truncated logarithmic distribution.

## Usage

```
oalog(lpobs1 = "logitlink", lshape = "logitlink",
      type.fitted = c("mean", "shape", "pobs1", "onempobs1"),
      ipobs1 = NULL, gshape = ppoints(8), zero = NULL)
```

## Arguments

`lpobs1` Link function for the parameter  $p_1$  or  $\phi$ , called `pobs1` or `phi` here. See [Links](#) for more choices.

`lshape` See [logff](#) for details.

`gshape`, `type.fitted` See [CommonVGAMffArguments](#) and [fittedv1m](#) for information.

`ipobs1`, `zero` See [CommonVGAMffArguments](#) for information.

## Details

The response  $Y$  is one with probability  $p_1$ , or  $Y$  has a 1-truncated logarithmic distribution with probability  $1 - p_1$ . Thus  $0 < p_1 < 1$ , which is modelled as a function of the covariates. The one-altered logarithmic distribution differs from the one-inflated logarithmic distribution in that the former has ones coming from one source, whereas the latter has ones coming from the logarithmic distribution too. The one-inflated logarithmic distribution is implemented in the **VGAM** package. Some people call the one-altered logarithmic a *hurdle* model.

The input can be a matrix (multiple responses). By default, the two linear/additive predictors of `oalog` are  $(\text{logit}(\phi), \text{logit}(s))^T$ .

## Value

An object of class `"vg1mff"` (see [vg1mff-class](#)). The object is used by modelling functions such as [vg1m](#), and [vgam](#).

The `fitted.values` slot of the fitted object, which should be extracted by the generic function `fitted`, returns the mean  $\mu$  (default) which is given by

$$\mu = \phi + (1 - \phi)A$$

where  $A$  is the mean of the one-truncated logarithmic distribution. If `type.fitted = "pobs1"` then  $p_1$  is returned.

**Note**

This family function effectively combines [binomialff](#) and [otlog](#) into one family function.

**Author(s)**

T. W. Yee

**See Also**

[Gaitdlog](#), [Oalog](#), [logff](#), [oilog](#), [CommonVGAMffArguments](#), [simulate.vlm](#).

**Examples**

```
## Not run: odata <- data.frame(x2 = runif(nn <- 1000))
odata <- transform(odata, pobs1 = logitlink(-1 + 2*x2, inverse = TRUE),
                  shape = logitlink(-2 + 3*x2, inverse = TRUE))
odata <- transform(odata, y1 = roalog(nn, shape = shape, pobs1 = pobs1),
                  y2 = roalog(nn, shape = shape, pobs1 = pobs1))
with(odata, table(y1))

ofit <- vglm(cbind(y1, y2) ~ x2, oalog, data = odata, trace = TRUE)
coef(ofit, matrix = TRUE)
head(fitted(ofit))
head(predict(ofit))
summary(ofit)

## End(Not run)
```

---

Oapospois

*One-Altered Logarithmic Distribution*

---

**Description**

Density, distribution function, quantile function and random generation for the one-altered positive-Poisson distribution with parameter  $pobs1$ .

**Usage**

```
doapospois(x, lambda, pobs1 = 0, log = FALSE)
poapospois(q, lambda, pobs1 = 0)
qoapospois(p, lambda, pobs1 = 0)
roapospois(n, lambda, pobs1 = 0)
```

**Arguments**

$x$ , $q$ , $n$ , $p$	Same <a href="#">Unif</a> .
$\lambda$ , $\log$	Same as <a href="#">Otpospois</a> .
$pobs1$	Probability of (an observed) one, called <i>pobs1</i> . The default value of $pobs1 = 0$ corresponds to the response having a 1-truncated positive-Poisson distribution.

**Details**

The probability function of  $Y$  is 1 with probability `pobs1`, else a 1-truncated positive-Poisson(`lambda`) distribution.

**Value**

`doapospois` gives the density and `poapospois` gives the distribution function, `qoapospois` gives the quantile function, and `roapospois` generates random deviates.

**Note**

The argument `pobs1` is recycled to the required length, and must have values which lie in the interval  $[0, 1]$ .

**Author(s)**

T. W. Yee

**See Also**

[oapospoisson](#), [Oipospois](#), [Otpospois](#).

**Examples**

```
lambda <- 3; pobs1 <- 0.30; x <- (-1):7
doapospois(x, lambda = lambda, pobs1 = pobs1)
table(roapospois(100, lambda = lambda, pobs1 = pobs1))

## Not run: x <- 0:10
barplot(rbind(doapospois(x, lambda = lambda, pobs1 = pobs1),
              dpospois(x, lambda = lambda)),
        beside = TRUE, col = c("blue", "orange"), cex.main = 0.7, las = 1,
        ylab = "Probability", names.arg = as.character(x),
        main = paste("OAPP(lambda = ", lambda, ", pobs1 = ", pobs1,
                    ") [blue] vs", " PosPoisson(lambda = ", lambda,
                    ") [orange] densities", sep = ""))

## End(Not run)
```

---

oapospoisson

*One-Altered Positive-Poisson Distribution*

---

**Description**

Fits a one-altered positive-Poisson distribution based on a conditional model involving a Bernoulli distribution and a 1-truncated positive-Poisson distribution.

**Usage**

```
oapospoisson(lpobs1 = "logitlink", llambda = "loglink",
             type.fitted = c("mean", "lambda", "pobs1", "onempobs1"),
             ipobs1 = NULL, zero = NULL)
```

**Arguments**

`lpobs1` Link function for the parameter  $p_1$  or  $\phi$ , called `pobs1` or `phi` here. See [Links](#) for more choices.

`llambda` See [pospoisson](#) for details.

`type.fitted` See [CommonVGAMffArguments](#) and [fittedv1m](#) for information.

`ipobs1, zero` See [CommonVGAMffArguments](#) for information.

**Details**

The response  $Y$  is one with probability  $p_1$ , or  $Y$  has a 1-truncated positive-Poisson distribution with probability  $1 - p_1$ . Thus  $0 < p_1 < 1$ , which is modelled as a function of the covariates. The one-altered positive-Poisson distribution differs from the one-inflated positive-Poisson distribution in that the former has ones coming from one source, whereas the latter has ones coming from the positive-Poisson distribution too. The one-inflated positive-Poisson distribution is implemented in the **VGAM** package. Some people call the one-altered positive-Poisson a *hurdle* model.

The input can be a matrix (multiple responses). By default, the two linear/additive predictors of `oapospoisson` are  $(\text{logit}(\phi), \log(\lambda))^T$ .

**Value**

An object of class "vglmff" (see [vglmff-class](#)). The object is used by modelling functions such as [vglm](#), and [vgam](#).

The `fitted.values` slot of the fitted object, which should be extracted by the generic function `fitted`, returns the mean  $\mu$  (default) which is given by

$$\mu = \phi + (1 - \phi)A$$

where  $A$  is the mean of the one-truncated positive-Poisson distribution. If `type.fitted = "pobs1"` then  $p_1$  is returned.

**Note**

This family function effectively combines [binomialff](#) and [otpospoisson](#) into one family function.

**Author(s)**

T. W. Yee

**See Also**

[Oapospois](#), [pospoisson](#), [oipospoisson](#), [CommonVGAMffArguments](#), [simulate.v1m](#).

**Examples**

```
## Not run: odata <- data.frame(x2 = runif(nn <- 1000))
odata <- transform(odata, pobs1 = logitlink(-1 + 2*x2, inverse = TRUE),
                    lambda = loglink( 1 + 1*x2, inverse = TRUE))
odata <- transform(odata, y1 = roapospois(nn, lambda = lambda, pobs1 = pobs1),
                    y2 = roapospois(nn, lambda = lambda, pobs1 = pobs1))
with(odata, table(y1))

ofit <- vglm(cbind(y1, y2) ~ x2, oapospoisson, data = odata, trace = TRUE)
coef(ofit, matrix = TRUE)
head(fitted(ofit))
head(predict(ofit))
summary(ofit)

## End(Not run)
```

Oazeta

*One-Altered Logarithmic Distribution***Description**

Density, distribution function, quantile function and random generation for the one-altered zeta distribution with parameter  $pobs1$ .

**Usage**

```
doazeta(x, shape, pobs1 = 0, log = FALSE)
poazeta(q, shape, pobs1 = 0)
qoazeta(p, shape, pobs1 = 0)
roazeta(n, shape, pobs1 = 0)
```

**Arguments**

$x$ , $q$ , $n$ , $p$	Same <a href="#">Unif.</a>
shape, log	Same as <a href="#">Otzeta</a> .
pobs1	Probability of (an observed) one, called <i>pobs1</i> . The default value of $pobs1 = 0$ corresponds to the response having a 1-truncated zeta distribution.

**Details**

The probability function of  $Y$  is 1 with probability  $pobs1$ , else a 1-truncated zeta distribution.

**Value**

`doazeta` gives the density and `poazeta` gives the distribution function, `qoazeta` gives the quantile function, and `roazeta` generates random deviates.

**Note**

The argument `pobs1` is recycled to the required length, and must have values which lie in the interval  $[0, 1]$ .

**Author(s)**

T. W. Yee

**See Also**

[oazeta](#), [Oizeta](#), [Otzeta](#), [zeta](#).

**Examples**

```
shape <- 1.1; pobs1 <- 0.10; x <- (-1):7
doazeta(x, shape = shape, pobs1 = pobs1)
table(roazeta(100, shape = shape, pobs1 = pobs1))

## Not run: x <- 0:10
barplot(rbind(doazeta(x, shape = shape, pobs1 = pobs1),
              dzeta(x, shape = shape)),
        beside = TRUE, col = c("blue", "orange"), cex.main = 0.7, las = 1,
        ylab = "Probability", names.arg = as.character(x),
        main = paste("OAZ(shape = ", shape, ", pobs1 = ", pobs1,
                    ") [blue] vs", " zeta(shape = ", shape,
                    ") [orange] densities", sep = ""))

## End(Not run)
```

---

oazeta

*One-Altered Zeta Distribution*

---

**Description**

Fits a one-altered zeta distribution based on a conditional model involving a Bernoulli distribution and a 1-truncated zeta distribution.

**Usage**

```
oazeta(lpobs1 = "logitlink", lshape = "loglink",
       type.fitted = c("mean", "shape", "pobs1", "onempobs1"),
       gshape = exp((-4:3)/4), ishape = NULL, ipobs1 = NULL, zero = NULL)
```

**Arguments**

<code>lpobs1</code>	Link function for the parameter $p_1$ or $\phi$ , called <code>pobs1</code> or <code>phi</code> here. See <a href="#">Links</a> for more choices.
<code>lshape</code>	See <a href="#">zeta</a> for details.
<code>type.fitted</code>	See <a href="#">CommonVGAMffArguments</a> and <a href="#">fittedv1m</a> for information.
<code>gshape</code> , <code>ishape</code> , <code>ipobs1</code> , <code>zero</code>	See <a href="#">CommonVGAMffArguments</a> for information.

## Details

The response  $Y$  is one with probability  $p_1$ , or  $Y$  has a 1-truncated zeta distribution with probability  $1 - p_1$ . Thus  $0 < p_1 < 1$ , which is modelled as a function of the covariates. The one-altered zeta distribution differs from the one-inflated zeta distribution in that the former has ones coming from one source, whereas the latter has ones coming from the zeta distribution too. The one-inflated zeta distribution is implemented in the **VGAM** package. Some people call the one-altered zeta a *hurdle* model.

The input can be a matrix (multiple responses). By default, the two linear/additive predictors of oazeta are  $(\text{logit}(\phi), \log(\text{shape}))^T$ .

## Value

An object of class "vglmff" (see [vglmff-class](#)). The object is used by modelling functions such as [vglm](#), and [vgam](#).

The `fitted.values` slot of the fitted object, which should be extracted by the generic function `fitted`, returns the mean  $\mu$  (default) which is given by

$$\mu = \phi + (1 - \phi)A$$

where  $A$  is the mean of the one-truncated zeta distribution. If `type.fitted = "pobs1"` then  $p_1$  is returned.

## Note

This family function effectively combines [binomialff](#) and [otzeta](#) into one family function.

## Author(s)

T. W. Yee

## See Also

[Oazeta](#), [zetaaff](#), [oizeta](#), [otzeta](#), [CommonVGAMffArguments](#), [simulate.vlm](#).

## Examples

```
## Not run: odata <- data.frame(x2 = runif(nn <- 1000))
odata <- transform(odata, pobs1 = logitlink(-1 + 2*x2, inverse = TRUE),
  shape = loglink( 1 + 1*x2, inverse = TRUE))
odata <- transform(odata, y1 = roazeta(nn, shape = shape, pobs1 = pobs1),
  y2 = roazeta(nn, shape = shape, pobs1 = pobs1))
with(odata, table(y1))

ofit <- vglm(cbind(y1, y2) ~ x2, oazeta, data = odata, trace = TRUE)
coef(ofit, matrix = TRUE)
head(fitted(ofit))
head(predict(ofit))
summary(ofit)

## End(Not run)
```

Oilog

*One-Inflated Logarithmic Distribution***Description**

Density, distribution function, quantile function and random generation for the one-inflated logarithmic distribution with parameter `pstr1`.

**Usage**

```
doilog(x, shape, pstr1 = 0, log = FALSE)
poilog(q, shape, pstr1 = 0)
qoilog(p, shape, pstr1 = 0)
roilog(n, shape, pstr1 = 0)
```

**Arguments**

<code>x, q, p, n</code>	Same as <a href="#">Uniform</a> .
<code>shape</code>	Vector of parameters that lie in (0, 1).
<code>pstr1</code>	Probability of a structural one (i.e., ignoring the logarithmic distribution), called $\phi$ . The default value of $\phi = 0$ corresponds to the response having an ordinary logarithmic distribution.
<code>log</code>	Same as <a href="#">Uniform</a> .

**Details**

The probability function of  $Y$  is 1 with probability  $\phi$ , and *Logarithmic*(*prob*) with probability  $1 - \phi$ . Thus

$$P(Y = 1) = \phi + (1 - \phi)P(W = 1)$$

where  $W$  is distributed as a *Logarithmic*(*shape*) random variable. The **VGAM** family function [oilog](#) estimates  $\phi$  by MLE.

**Value**

`doilog` gives the density, `poilog` gives the distribution function, `qoilog` gives the quantile function, and `roilog` generates random deviates.

**Note**

The argument `pstr1` is recycled to the required length, and usually has values which lie in the interval  $[0, 1]$ .

These functions actually allow for the *zero-deflated logarithmic* distribution. Here, `pstr1` is also permitted to lie in the interval  $[-\text{dlog}(1, \text{shape}) / (1 - \text{dlog}(1, \text{shape})), 0]$ . The resulting probability of a unit count is *less than* the nominal logarithmic value, and the use of `pstr1` to stand for the probability of a structural 1 loses its meaning.

When `pstr1` equals  $-\text{dlog}(1, \text{shape}) / (1 - \text{dlog}(1, \text{shape}))$  this corresponds to the 1-truncated logarithmic distribution.

**Author(s)**

T. W. Yee

**See Also**[Gaitdlog](#), [oilog](#), [rlog](#), [logff](#), [Otlog](#).**Examples**

```

shape <- 0.5; pstr1 <- 0.3; x <- (-1):7
(ii <- doilog(x, shape, pstr1 = pstr1))
max(abs(poilog(1:200, shape) -
      cumsum(shape^(1:200) / (-(1:200) * log1p(-shape))))) # Should be 0

## Not run: x <- 0:10
par(mfrow = c(2, 1)) # One-Inflated logarithmic
barplot(rbind(doilog(x, shape, pstr1 = pstr1), dlog(x, shape)),
        beside = TRUE, col = c("blue", "orange"),
        main = paste("OILogff(", shape, ", ", pstr1 = ", pstr1, ") (blue) vs",
                    " Logff(", shape, ") (orange)", sep = ""),
        names.arg = as.character(x))

deflat.limit <- -dlog(1, shape) / plog(1, shape, lower.tail = FALSE)
newpstr1 <- round(deflat.limit, 3) + 0.001 # Inside but near the boundary
barplot(rbind(doilog(x, shape, pstr1 = newpstr1),
              dlog(x, shape)),
        beside = TRUE, col = c("blue", "orange"),
        main = paste("ODLogff(", shape, ", ", pstr1 = ", newpstr1, ") (blue) vs",
                    " Logff(", shape, ") (orange)", sep = ""),
        names.arg = as.character(x))
## End(Not run)

```

---

**oilog***One-inflated Logarithmic Distribution Family Function*

---

**Description**

Fits a 1-inflated logarithmic distribution.

**Usage**

```

oilog(lpstr1 = "logitlink", lshape = "logitlink",
      type.fitted = c("mean", "shape", "pobs1", "pstr1", "onempstr1"),
      ishape = NULL, gpstr1 = ppoints(8), gshape = ppoints(8), zero = NULL)

```

**Arguments**

lpstr1, lshape Link functions. For lpstr1: the same idea as [zipoisson](#) except it applies to a structural 1.

gpstr1, gshape, ishape  
For initial values. See [CommonVGAMffArguments](#) for information.

type.fitted, zero  
See [CommonVGAMffArguments](#) for information.

**Details**

The 1-inflated logarithmic distribution is a mixture distribution of the logarithmic distribution with some probability of obtaining a (structural) 1. Thus there are two sources for obtaining the value 1. This distribution is written here in a way that retains a similar notation to the one-inflated positive-Poisson, i.e., the probability  $P[Y = 1]$  involves another parameter  $\phi$ . See [oipoisson](#).

This family function can handle multiple responses.

**Value**

An object of class "vglmff" (see [vglmff-class](#)). The object is used by modelling functions such as [vglm](#), [rrvglm](#) and [vgam](#).

**Author(s)**

Thomas W. Yee

**See Also**

[Gaitdlog](#), [Oilog](#), [logff](#), [Oizeta](#).

**Examples**

```
## Not run: odata <- data.frame(x2 = runif(nn <- 1000)) # Artificial data
odata <- transform(odata, pstr1 = logitlink(-1 + x2, inverse = TRUE), shape = 0.5)
odata <- transform(odata, y1 = roilog(nn, shape, pstr1 = pstr1))
with(odata, table(y1))
fit1 <- vglm(y1 ~ x2, oilog(zero = "shape"), data = odata, trace = TRUE)
coef(fit1, matrix = TRUE)

## End(Not run)
```

---

Oiposbinom

*One-Inflated Positive Binomial Distribution*


---

### Description

Density, distribution function, quantile function and random generation for the one-inflated positive binomial distribution with parameter `pstr1`.

### Usage

```
doiposbinom(x, size, prob, pstr1 = 0, log = FALSE)
poiposbinom(q, size, prob, pstr1 = 0)
qoiposbinom(p, size, prob, pstr1 = 0)
roiposbinom(n, size, prob, pstr1 = 0)
```

### Arguments

<code>x, p, q, n</code>	Same as <a href="#">Posbinom</a> .
<code>size, prob</code>	Same as <a href="#">Posbinom</a> .
<code>pstr1</code>	Probability of a structural one (i.e., ignoring the positive binomial distribution), called $\phi$ . The default value of $\phi = 0$ corresponds to the response having a positive binomial distribution. However, <code>pstr1</code> can also be negative, in which case it ceases its interpretation as a probability, and this is known as <i>one-deflation</i> .
<code>log</code>	Logical. Return the logarithm of the answer?

### Details

The probability function of  $Y$  is 1 with probability  $\phi$ , and *PosBinomial*(*size*, *prob*) with probability  $1 - \phi$ . Thus

$$P(Y = 1) = \phi + (1 - \phi)P(W = 1)$$

where  $W$  is distributed as a positive *binomial*(*size*, *prob*) random variable.

### Value

`doiposbinom` gives the density, `poiposbinom` gives the distribution function, `qoiposbinom` gives the quantile function, and `roiposbinom` generates random deviates.

### Note

The argument `pstr1` is recycled to the required length, and usually has values which lie in the interval  $[0, 1]$ .

These functions actually allow for the *zero-deflated binomial* distribution. Here, `pstr1` is also permitted to lie in the interval  $[-A, 0]$  for some positive quantity  $A$ . The resulting probability of a unit value is *less than* the nominal positive binomial value, and the use of `pstr1` to stand for the probability of a structural 1 loses its meaning.

If `pstr1` equals  $A$  then this corresponds to the 0- and 1-truncated binomial distribution.

**Author(s)**

T. W. Yee

**See Also**[posbinomial](#), [dbinom](#), [binomialff](#).**Examples**

```

size <- 10; prob <- 0.2; pstr1 <- 0.4; x <- (-1):size
(ii <- doiposbinom(x, size, prob, pstr1 = pstr1))
table(roiposbinom(100, size, prob, pstr1 = pstr1))
round(doiposbinom(x, size, prob, pstr1 = pstr1) * 100) # Should be similar

## Not run: x <- 0:size
par(mfrow = c(2, 1)) # One-Inflated Positive Binomial
barplot(rbind(doiposbinom(x, size, prob, pstr1 = pstr1),
             dposbinom(x, size, prob)),
        beside = TRUE, col = c("blue", "orange"),
        main = paste("OIPB(", size, ",", prob, ", pstr1 = ", pstr1, ") (blue) vs",
                    " PosBinomial(", size, ",", prob, ") (orange)", sep = ""),
        names.arg = as.character(x))

# Zero-deflated Pos Binomial
deflat.limit <- -dposbinom(1, size, prob) / (1 - dposbinom(1, size, prob))
deflat.limit <- size * prob / (1 + (size-1) * prob - 1 / (1-prob)^(size-1))
newpstr1 <- round(deflat.limit, 3) + 0.001 # A little from the boundary
barplot(rbind(doiposbinom(x, size, prob, pstr1 = newpstr1),
             dposbinom(x, size, prob)),
        beside = TRUE, col = c("blue", "orange"),
        main = paste("ODPB(", size, ",", prob, ", pstr1 = ", newpstr1, ") (blue) vs",
                    " PosBinomial(", size, ",", prob, ") (orange)", sep = ""),
        names.arg = as.character(x))
## End(Not run)

```

oiposbinomial

*One-Inflated Positive Binomial Distribution Family Function***Description**

Fits a one-inflated positive binomial distribution by maximum likelihood estimation.

**Usage**

```

oiposbinomial(lpstr1 = "logitlink", lprob = "logitlink",
             type.fitted = c("mean", "prob", "pobs1", "pstr1", "onempstr1"),
             iprob = NULL, gpstr1 = ppoints(9), gprob = ppoints(9),
             multiple.responses = FALSE, zero = NULL)

```

**Arguments**

lpstr1, lprob	Link functions for the parameter $\phi$ and the positive binomial probability $\mu$ parameter. See <a href="#">Links</a> for more choices. See <a href="#">CommonVGAMffArguments</a> also. For the <i>one-deflated</i> model see below.
type.fitted	See <a href="#">CommonVGAMffArguments</a> and <a href="#">fittedvln</a> .
iprob, gpstr1, gprob	For initial values; see <a href="#">CommonVGAMffArguments</a> .
multiple.responses	Logical. See <a href="#">binomialff</a> and <a href="#">posbinomial</a> .
zero	See <a href="#">CommonVGAMffArguments</a> for information.

**Details**

These functions are based on

$$P(Y = y) = \phi + (1 - \phi)N\mu(1 - \mu)^N / (1 - (1 - \mu)^N),$$

for  $y = 1/N$ , and

$$P(Y = y) = (1 - \phi) \binom{N}{Ny} \mu^{Ny} (1 - \mu)^{N(1-y)} / (1 - (1 - \mu)^N).$$

for  $y = 2/N, \dots, 1$ . That is, the response is a sample proportion out of  $N$  trials, and the argument size in [roiposbinom](#) is  $N$  here. Ideally  $N > 2$  is needed. The parameter  $\phi$  is the probability of a structural one, and it satisfies  $0 < \phi < 1$  (usually). The mean of  $Y$  is  $E(Y) = \phi + (1 - \phi)\mu / (1 - (1 - \mu)^N)$  and these are returned as the default fitted values. By default, the two linear/additive predictors for [oiposbinomial\(\)](#) are  $(\text{logit}(\phi), \text{logit}(\mu))^T$ .

**Value**

An object of class "vglmff" (see [vglmff-class](#)). The object is used by modelling functions such as [vglm](#) and [vgam](#).

**Note**

The response variable should have one of the formats described by [binomialff](#), e.g., a factor or two column matrix or a vector of sample proportions with the `weights` argument specifying the values of  $N$ .

To work well, one ideally needs large values of  $N$  and  $\mu$  much greater than 0, i.e., the larger  $N$  and  $\mu$  are, the better. If  $N = 1$  then the model is unidentifiable since the number of parameters is excessive.

Estimated probabilities of a structural one and an observed one are returned, as in [zipoisson](#).

The *one-deflated* positive binomial distribution might be fitted by setting `lpstr1 = "identitylink"`, albeit, not entirely reliably. See [zipoisson](#) for information that can be applied here.

**Author(s)**

T. W. Yee

**See Also**

[roiposbinom](#), [posbinomial](#), [binomialff](#), [rbinom](#).

**Examples**

```
size <- 10 # Number of trials; N in the notation above
nn <- 200
odata <- data.frame(pstr1 = logitlink( 0, inverse = TRUE), # 0.50
                   mubin1 = logitlink(-1, inverse = TRUE), # Mean of usual binomial
                   svec  = rep(size, length = nn),
                   x2    = runif(nn))
odata <- transform(odata,
                   mubin2 = logitlink(-1 + x2, inverse = TRUE))
odata <- transform(odata,
                   y1 = roiposbinom(nn, svec, pr = mubin1, pstr1 = pstr1),
                   y2 = roiposbinom(nn, svec, pr = mubin2, pstr1 = pstr1))
with(odata, table(y1))
fit1 <- vglm(y1 / svec ~ 1, oiposbinomial, data = odata,
             weights = svec, trace = TRUE, crit = "coef")
fit2 <- vglm(y2 / svec ~ x2, oiposbinomial, data = odata,
             weights = svec, trace = TRUE)

coef(fit1, matrix = TRUE)
Coef(fit1) # Useful for intercept-only models
head(fitted(fit1, type = "pobs1")) # Estimate of P(Y = 1)
head(fitted(fit1))
with(odata, mean(y1)) # Compare this with fitted(fit1)
summary(fit1)
```

---

Oipospois

*One-Inflated Positive Poisson Distribution*

---

**Description**

Density, distribution function, quantile function and random generation for the one-inflated positive Poisson distribution with parameter `pstr1`.

**Usage**

```
doipospois(x, lambda, pstr1 = 0, log = FALSE)
poiipospois(q, lambda, pstr1 = 0)
qoiipospois(p, lambda, pstr1 = 0)
roiipospois(n, lambda, pstr1 = 0)
```

**Arguments**

`x`, `p`, `q`, `n`      Same as [Pospois](#).  
`lambda`                    Vector of positive means.

pstr1	Probability of a structural one (i.e., ignoring the positive Poisson distribution), called $\phi$ . The default value of $\phi = 0$ corresponds to the response having a positive Poisson distribution.
log	Logical. Return the logarithm of the answer?

### Details

The probability function of  $Y$  is 1 with probability  $\phi$ , and  $PosPoisson(\lambda)$  with probability  $1 - \phi$ . Thus

$$P(Y = 1) = \phi + (1 - \phi)P(W = 1)$$

where  $W$  is distributed as a positive  $Poisson(\lambda)$  random variate.

### Value

doipospois gives the density, poipospois gives the distribution function, qoipospois gives the quantile function, and roipospois generates random deviates.

### Note

The argument pstr1 is recycled to the required length, and usually has values which lie in the interval  $[0, 1]$ .

These functions actually allow for the *zero-deflated Poisson* distribution. Here, pstr1 is also permitted to lie in the interval  $[-\lambda / (\exp(1) - 1), 0]$ . The resulting probability of a unit count is *less than* the nominal positive Poisson value, and the use of pstr1 to stand for the probability of a structural 1 loses its meaning.

When pstr1 equals  $-\lambda / (\exp(1) - 1)$  this corresponds to the 0- and 1-truncated Poisson distribution.

### Author(s)

T. W. Yee

### See Also

[Pospois](#), [oapospoisson](#), [oipospoisson](#), [otpospoisson](#), [pospoisson](#), [dpois](#), [poissonff](#).

### Examples

```
lambda <- 3; pstr1 <- 0.2; x <- (-1):7
(ii <- doipospois(x, lambda, pstr1 = pstr1))
table(roipospois(100, lambda, pstr1 = pstr1))
round(doipospois(1:10, lambda, pstr1 = pstr1) * 100) # Should be similar

## Not run: x <- 0:10
par(mfrow = c(2, 1)) # One-Inflated Positive Poisson
barplot(rbind(doipospois(x, lambda, pstr1 = pstr1), dpospois(x, lambda)),
        beside = TRUE, col = c("blue", "orange"),
        main = paste("OIPP(", lambda, ", ", pstr1 = ", pstr1, ") (blue) vs",
                    " PosPoisson(", lambda, ") (orange)", sep = ""))
```

```

names.arg = as.character(x))

deflat.limit <- -lambda / (expm1(lambda) - lambda) # 0-deflated Pos Poisson
newpstr1 <- round(deflat.limit, 3) + 0.001 # Inside and near the boundary
barplot(rbind(doipospois(x, lambda, pstr1 = newpstr1),
              dpospois(x, lambda)),
        beside = TRUE, col = c("blue", "orange"),
        main = paste("ODPP(", lambda, ", ", pstr1 = ", newpstr1, ") (blue) vs",
                    " PosPoisson(", lambda, ", ") (orange)", sep = ""),
        names.arg = as.character(x))
## End(Not run)

```

---

oipospoisson

*One-inflated Positive Poisson Distribution Family Function*


---

## Description

Fits a 1-inflated positive Poisson distribution.

## Usage

```

oipospoisson(lpstr1 = "logitlink", llambda = "loglink",
             type.fitted = c("mean", "lambda", "pobs1", "pstr1", "onempstr1"),
             ilambda = NULL, gpstr1 = (1:19)/20, gprobs.y = (1:19)/20,
             imethod = 1, zero = NULL)

```

## Arguments

lpstr1, llambda

For lpstr1: the same idea as [zipoisson](#) except it applies to a structural 1.

ilambda, gpstr1, gprobs.y, imethod

For initial values. See [CommonVGAMffArguments](#) for information.

type.fitted, zero

See [CommonVGAMffArguments](#) for information.

## Details

The 1-inflated positive Poisson distribution is a mixture distribution of the positive (0-truncated) Poisson distribution with some probability of obtaining a (structural) 1. Thus there are two sources for obtaining the value 1. It is similar to a zero-inflated Poisson model, except the Poisson is replaced by a positive Poisson and the 0 is replaced by 1. This distribution is written here in a way that retains a similar notation to the zero-inflated Poisson, i.e., the probability  $P[Y = 1]$  involves another parameter  $\phi$ . See [zipoisson](#).

This family function can handle multiple responses.

## Value

An object of class "vglmff" (see [vglmff-class](#)). The object is used by modelling functions such as [vglm](#), [rrvglm](#) and [vgam](#).

**Warning**

Under- or over-flow may occur if the data is ill-conditioned.

**Author(s)**

Thomas W. Yee

**See Also**

[Oipospois](#), [pospoisson](#), [oapospoisson](#), [otpospoisson](#), [zipoisson](#), [poissonff](#), [simulate.vlm](#).

**Examples**

```
## Not run: set.seed(1)
pdata <- data.frame(x2 = runif(nn <- 1000)) # Artificial data
pdata <- transform(pdata, pstr1 = 0.5, lambda = exp(3 - x2))
pdata <- transform(pdata, y1 = roipospois(nn, lambda, pstr1 = pstr1))
with(pdata, table(y1))
fit1 <- vglm(y1 ~ x2, oipospoisson, data = pdata, trace = TRUE)
coef(fit1, matrix = TRUE)

## End(Not run)
```

---

Oizeta

*One-Inflated Zeta Distribution*

---

**Description**

Density, distribution function, quantile function and random generation for the one-inflated zeta distribution with parameter `pstr1`.

**Usage**

```
doizeta(x, shape, pstr1 = 0, log = FALSE)
poizeta(q, shape, pstr1 = 0)
qoizeta(p, shape, pstr1 = 0)
roizeta(n, shape, pstr1 = 0)
```

**Arguments**

<code>x</code> , <code>q</code> , <code>p</code> , <code>n</code>	Same as <a href="#">Uniform</a> .
<code>shape</code>	Vector of positive shape parameters.
<code>pstr1</code>	Probability of a structural one (i.e., ignoring the zeta distribution), called $\phi$ . The default value of $\phi = 0$ corresponds to the response having an ordinary zeta distribution.
<code>log</code>	Same as <a href="#">Uniform</a> .

**Details**

The probability function of  $Y$  is 1 with probability  $\phi$ , and  $Zeta(shape)$  with probability  $1 - \phi$ . Thus

$$P(Y = 1) = \phi + (1 - \phi)P(W = 1)$$

where  $W$  is distributed as a  $zeta(shape)$  random variable.

**Value**

doizeta gives the density, poizeta gives the distribution function, qoizeta gives the quantile function, and roizeta generates random deviates.

**Note**

The argument `pstr1` is recycled to the required length, and usually has values which lie in the interval  $[0, 1]$ .

These functions actually allow for the *zero-deflated zeta* distribution. Here, `pstr1` is also permitted to lie in the interval  $[-dzeta(1, shape) / (1 - dzeta(1, shape)), 0]$ . The resulting probability of a unit count is *less than* the nominal zeta value, and the use of `pstr1` to stand for the probability of a structural 1 loses its meaning.

When `pstr1` equals  $-dzeta(1, shape) / (1 - dzeta(1, shape))$  this corresponds to the 1-truncated zeta distribution.

**Author(s)**

T. W. Yee

**See Also**

[Zeta](#), [zetaaff](#), [Otzeta](#),

**Examples**

```
shape <- 1.5; pstr1 <- 0.3; x <- (-1):7
(ii <- doizeta(x, shape, pstr1 = pstr1))
max(abs(poizeta(1:200, shape) -
      cumsum(1/(1:200)^(1+shape)) / zeta(shape+1))) # Should be 0

## Not run: x <- 0:10
par(mfrow = c(2, 1)) # One-Inflated zeta
barplot(rbind(doizeta(x, shape, pstr1 = pstr1), dzeta(x, shape)),
        beside = TRUE, col = c("blue", "orange"),
        main = paste("OIZeta(", shape, ", pstr1 = ", pstr1, ") (blue) vs",
                    "Zeta(", shape, ") (orange)", sep = ""),
        names.arg = as.character(x))

deflat.limit <- -dzeta(1, shape) / pzeta(1, shape, lower.tail = FALSE)
newpstr1 <- round(deflat.limit, 3) + 0.001 # Inside but near the boundary
barplot(rbind(doizeta(x, shape, pstr1 = newpstr1),
              dzeta(x, shape)),
```

```

beside = TRUE, col = c("blue", "orange"),
main = paste("0DZeta(", shape, ", pstr1 = ", newpstr1, ") (blue) vs",
            " Zeta(", shape, ") (orange)", sep = ""),
names.arg = as.character(x))
## End(Not run)

```

---

oizeta

*One-inflated Zeta Distribution Family Function*


---

## Description

Fits a 1-inflated zeta distribution.

## Usage

```

oizeta(lpstr1 = "logitlink", lshape = "loglink",
       type.fitted = c("mean", "shape", "pobs1", "pstr1", "onempstr1"),
       ishape = NULL, gpstr1 = ppoints(8), gshape = exp((-3:3) / 4), zero = NULL)

```

## Arguments

lpstr1, lshape For lpstr1: the same idea as [zipoisson](#) except it applies to a structural 1.  
gpstr1, gshape, ishape For initial values. See [CommonVGAMffArguments](#) for information.  
type.fitted, zero See [CommonVGAMffArguments](#) for information.

## Details

The 1-inflated zeta distribution is a mixture distribution of the zeta distribution with some probability of obtaining a (structural) 1. Thus there are two sources for obtaining the value 1. This distribution is written here in a way that retains a similar notation to the zero-inflated Poisson, i.e., the probability  $P[Y = 1]$  involves another parameter  $\phi$ . See [zipoisson](#).

This family function can handle multiple responses.

## Value

An object of class "vglmff" (see [vglmff-class](#)). The object is used by modelling functions such as [vglm](#), [rrvglm](#) and [vgam](#).

## Warning

Under- or over-flow may occur if the data is ill-conditioned. Lots of data is needed to estimate the parameters accurately. Usually, probably the shape parameter is best modelled as intercept-only.

## Author(s)

Thomas W. Yee

**See Also**

[Oizeta](#), [zetaff](#), [oazeta](#), [otzeta](#), [diffzeta](#), [zeta](#), [Oizipf](#).

**Examples**

```
## Not run: odata <- data.frame(x2 = runif(nn <- 1000)) # Artificial data
odata <- transform(odata, pstr1 = logitlink(-1 + x2, inverse = TRUE), shape = exp(-0.5))
odata <- transform(odata, y1 = roizeta(nn, shape, pstr1 = pstr1))
with(odata, table(y1))
fit1 <- vglm(y1 ~ x2, oizeta(zero = "shape"), data = odata, trace = TRUE)
coef(fit1, matrix = TRUE)

## End(Not run)
```

---

Oizipf

*One-Inflated Zipf Distribution*


---

**Description**

Density, distribution function, quantile function and random generation for the one-inflated Zipf distribution with parameter `pstr1`.

**Usage**

```
doizipf(x, N, shape, pstr1 = 0, log = FALSE)
poizipf(q, N, shape, pstr1 = 0)
qoizipf(p, N, shape, pstr1 = 0)
roizipf(n, N, shape, pstr1 = 0)
```

**Arguments**

<code>x</code> , <code>q</code> , <code>p</code> , <code>n</code>	Same as <a href="#">Uniform</a> .
<code>N</code> , <code>shape</code>	See <a href="#">Zipf</a> .
<code>pstr1</code>	Probability of a structural one (i.e., ignoring the Zipf distribution), called $\phi$ . The default value of $\phi = 0$ corresponds to the response having an ordinary Zipf distribution.
<code>log</code>	Same as <a href="#">Uniform</a> .

**Details**

The probability function of  $Y$  is 1 with probability  $\phi$ , and  $Zipf(N, s)$  with probability  $1 - \phi$ . Thus

$$P(Y = 1) = \phi + (1 - \phi)P(W = 1)$$

where  $W$  is distributed as a  $Zipf(N, s)$  random variable. The **VGAM** family function [oizeta](#) estimates the two parameters of this model by Fisher scoring.

**Value**

doizipf gives the density, poizipf gives the distribution function, qoizipf gives the quantile function, and roizipf generates random deviates.

**Note**

The argument pstr1 is recycled to the required length, and usually has values which lie in the interval  $[0, 1]$ .

These functions actually allow for the *zero-deflated Zipf* distribution. Here, pstr1 is also permitted to lie in the interval  $[-dzipf(1, N, s) / (1 - dzipf(1, N, s)), 0]$ . The resulting probability of a unit count is *less than* the nominal zipf value, and the use of pstr1 to stand for the probability of a structural 1 loses its meaning.

When pstr1 equals  $-dzipf(1, N, s) / (1 - dzipf(1, N, s))$  this corresponds to the 1-truncated zipf distribution.

**Author(s)**

T. W. Yee

**See Also**

[oizeta](#), [Zipf](#), [zipf](#), [Oizeta](#).

**Examples**

```
N <- 10; shape <- 1.5; pstr1 <- 0.3; x <- (-1):N
(ii <- doizipf(x, N, shape, pstr1 = pstr1))

## Not run: x <- 0:10
par(mfrow = c(2, 1)) # One-Inflated zipf
barplot(rbind(doizipf(x, N, shape, pstr1 = pstr1), dzipf(x, N, shape)),
        beside = TRUE, col = c("blue", "orange"),
        main = paste("OIZipf(", N, ", ", shape, ", pstr1 = ", pstr1, ") (blue) vs",
                    " Zipf(", N, ", ", shape, ") (orange)", sep = ""),
        names.arg = as.character(x))

deflat.limit <- -dzipf(1, N, shape) / (1 - dzipf(1, N, shape))
newpstr1 <- round(deflat.limit, 3) + 0.001 # Inside but near the boundary
barplot(rbind(doizipf(x, N, shape, pstr1 = newpstr1),
              dzipf(x, N, shape)),
        beside = TRUE, col = c("blue", "orange"),
        main = paste("ODZipf(", N, ", ", shape, ", pstr1 = ", newpstr1, ") (blue) vs",
                    " Zipf(", N, ", ", shape, ") (orange)", sep = ""),
        names.arg = as.character(x))
## End(Not run)
```

oizipf

*One-inflated Zipf Distribution Family Function***Description**

Fits a 1-inflated Zipf distribution.

**Usage**

```
oizipf(N = NULL, lpstr1 = "logitlink", lshape = "loglink",
       type.fitted = c("mean", "shape", "pobs1", "pstr1", "onempstr1"),
       ishape = NULL, gpstr1 = ppoints(8), gshape = exp((-3:3) / 4), zero = NULL)
```

**Arguments**

**N** Same as [zipf](#).

**lpstr1, lshape** For lpstr1: the same idea as [zipoisson](#) except it applies to a structural 1.

**gpstr1, gshape, ishape** For initial values. See [CommonVGAMffArguments](#) for information.

**type.fitted, zero** See [CommonVGAMffArguments](#) for information.

**Details**

The 1-inflated Zipf distribution is a mixture distribution of the Zipf distribution with some probability of obtaining a (structural) 1. Thus there are two sources for obtaining the value 1. This distribution is written here in a way that retains a similar notation to the zero-inflated Poisson, i.e., the probability  $P[Y = 1]$  involves another parameter  $\phi$ . See [zipoisson](#).

This family function can handle multiple responses.

**Value**

An object of class "vglmff" (see [vglmff-class](#)). The object is used by modelling functions such as [vglm](#), [rrvglm](#) and [vgam](#).

**Warning**

Under- or over-flow may occur if the data is ill-conditioned. Lots of data is needed to estimate the parameters accurately. Usually, probably the shape parameter is best modelled as intercept-only.

**Author(s)**

Thomas W. Yee

**See Also**

[Oizipf](#), [zipf](#), [Oizeta](#).

**Examples**

```
## Not run: odata <- data.frame(x2 = runif(nn <- 1000)) # Artificial data
odata <- transform(odata, pstr1 = logitlink(-1 + x2, inverse = TRUE),
                  myN = 10,
                  shape = exp(-0.5))
odata <- transform(odata, y1 = roizipf(nn, N = myN, s = shape, pstr1 = pstr1))
with(odata, table(y1))
fit1 <- vglm(y1 ~ x2, oizipf(zero = "shape"), data = odata, trace = TRUE)
coef(fit1, matrix = TRUE)

## End(Not run)
```

---

oly12

2012 Summer Olympics: Individuals Data

---

**Description**

Individual data for the Summer 2012 Olympic Games.

**Usage**

```
data(oly12)
```

**Format**

A data frame with 10384 observations on the following 14 variables.

Name The individual competitor's name.

Country Country.

Age A numeric vector, age in years.

Height A numeric vector, height in m.

Weight A numeric vector, weight in kg.

Sex A factor with levels F and M.

DOB A Date, date of birth.

PlaceOB Place of birth.

Gold Numeric vector, number of such medals won.

Silver Similar to Gold.

Bronze Similar to Gold.

Total A numeric vector, total number of medals.

Sport A factor with levels Archery, Athletics, Athletics, Triathlon, Badminton, etc.

Event The sporting event.

**Details**

This data set represents a very small modification of a .csv spreadsheet from the source below. Height has been converted to meters, and date of birth is of a "Date" class (see [as.Date](#)). A few non-ASCII characters have been replaced by some ASCII sequence (yet to be fixed up properly).

Some competitors share the same name. Some errors in the data are likely to exist.

**Source**

Downloaded from <http://www.guardian.co.uk/sport/series/london-2012-olympics-data> in 2013-03; more recently it has changed to <https://www.theguardian.com/sport/series/london-2012-olympics-data>.

**Examples**

```
data(oly12)
mtab <- with(oly12, table(Country, Gold))
(mtab <- head(sort(mtab[, "1"] + 2 * mtab[, "2"], decreasing = TRUE), 10))

## Not run:
barplot(mtab, col = "gold", cex.names = 0.8, names = abbreviate(names(mtab)),
        beside = TRUE, main = "2012 Summer Olympic Final Gold Medal Count",
        ylab = "Gold medal count", las = 1, sub = "Top 10 countries")

## End(Not run)
```

---

 Otlog

*One-truncated Logarithmic Distribution*


---

**Description**

Density, distribution function, quantile function, and random generation for the one-truncated logarithmic distribution.

**Usage**

```
dotlog(x, shape, log = FALSE)
potlog(q, shape, log.p = FALSE)
qotlog(p, shape)
rotlog(n, shape)
```

**Arguments**

x, q	Vector of quantiles. For the density, it should be a vector with integer values $> 1$ in order for the probabilities to be positive.
p	vector of probabilities.
n	number of observations. Same as in <a href="#">runif</a> .

shape            The parameter value  $c$  described in in [logff](#). Here it is called shape because  $0 < c < 1$  is the range.

log, log.p       Logical. If `log.p = TRUE` then all probabilities  $p$  are given as  $\log(p)$ .

### Details

The one-truncated logarithmic distribution is a logarithmic distribution but with the probability of a one being zero. The other probabilities are scaled to add to unity. Some more details are given in [logff](#).

### Value

`dotlog` gives the density, `potlog` gives the distribution function, `qotlog` gives the quantile function, and `rotlog` generates random deviates.

### Note

Given some response data, the **VGAM** family function `otlog` estimates the parameter shape. Function `potlog()` suffers from the problems that `plog` sometimes has.

### Author(s)

T. W. Yee

### See Also

[Gaitdlog](#), [otlog](#), [rlog](#), [Oilog](#).

### Examples

```
dotlog(1:20, 0.5)
rotlog(20, 0.5)

## Not run:  shape <- 0.8; x <- 1:10
plot(x, dotlog(x, shape = shape), type = "h", ylim = 0:1,
     sub = "shape=0.8", las = 1, col = "blue", ylab = "Probability",
     main = "1-truncated logarithmic distribution: blue=PMF; orange=CDF")
lines(x + 0.1, potlog(x, shape = shape), col = "orange", lty = 3, type = "h")
## End(Not run)
```

---

otlog

*One-truncated Logarithmic Distribution*

---

### Description

Estimating the (single) parameter of the 1-truncated logarithmic distribution.

**Usage**

```
otlog(lshape = "logitlink", gshape = ppoints(8), zero = NULL)
```

**Arguments**

lshape, gshape, zero  
Same as [logff](#).

**Details**

The 1-truncated logarithmic distribution is a logarithmic distribution but with the probability of a one being zero. The other probabilities are scaled to add to unity. Some more details can be found at [logff](#). Multiple responses are permitted.

**Value**

An object of class "vglmff" (see [vglmff-class](#)). The object is used by modelling functions such as [vglm](#), and [vgam](#).

**Author(s)**

T. W. Yee

**See Also**

[Gaitdlog](#), [Otlog](#), [logff](#), [oalog](#), [oilog](#), [simulate.vlm](#).

**Examples**

```
## Not run:
odata <- data.frame(y1 = rotlog(n = 1000, shape = logitlink(1/3, inverse = TRUE)))
ofit <- vglm(y1 ~ 1, otlog, data = odata, trace = TRUE, crit = "c")
coef(ofit, matrix = TRUE)
Coef(ofit)
with(odata,
     hist(y1, shape = TRUE, breaks = seq(0.5, max(y1) + 0.5, by = 1),
          border = "blue"))
x <- seq(1, with(odata, max(y1)), by = 1)
with(odata, lines(x, dotlog(x, Coef(ofit)[1]), col = "orange", type = "h", lwd = 2))
## End(Not run)
```

---

`Otpospois`*One-truncated Positive-Poisson Distribution*

---

**Description**

Density, distribution function, quantile function, and random generation for the one-truncated positive-Poisson distribution.

**Usage**

```
dotpospois(x, lambda, log = FALSE)
potpospois(q, lambda, log.p = FALSE)
qotpospois(p, lambda)
rotpospois(n, lambda)
```

**Arguments**

`x`, `q`, `p`, `n`      Same as [Pospois](#).  
`lambda`, `log`, `log.p`  
                         Same as [Pospois](#).

**Details**

The one-truncated positive-Poisson is a Poisson distribution but with the probability of a one and a zero being 0. That is, its support is 2, 3, .... The other probabilities are scaled to add to unity. Some more details are given in [pospoisson](#).

**Value**

`dotpospois` gives the density, `potpospois` gives the distribution function, `qotpospois` gives the quantile function, and `rotpospois` generates random deviates.

**Note**

Given some response data, the **VGAM** family function [otpospoisson](#) estimates the parameter `lambda`.

**Author(s)**

T. W. Yee

**See Also**

[otpospoisson](#), [Pospois](#), [Oipospois](#).

**Examples**

```

dotpospois(1:20, 0.5)
rotpospois(20, 0.5)

## Not run: lambda <- 4; x <- 1:10
plot(x, dotpospois(x, lambda = lambda), type = "h", ylim = 0:1,
      sub = "lambda=4", las = 1, col = "blue", ylab = "Probability",
      main = "1-truncated positive-Poisson distribution: blue=PMF; orange=CDF")
lines(x + 0.1, potpospois(x, lambda = lambda), col = "orange", lty = 3, type = "h")
## End(Not run)

```

---

otpospoisson	<i>One-truncated Poisson Distribution</i>
--------------	---

---

**Description**

Estimating the (single) parameter of the 1-truncated positive Poisson distribution.

**Usage**

```

otpospoisson(llambda = "loglink",
             type.fitted = c("mean", "lambda", "prob0", "prob1"),
             ilambda = NULL, imethod = 1, zero = NULL)

```

**Arguments**

llambda, type.fitted, ilambda  
                                   Same as [pospoisson](#).  
 imethod, zero   Same as [pospoisson](#).

**Details**

The 1-truncated positive Poisson distribution has support on 2, 3, ... It is a Poisson distribution but with the probability of a one or zero being 0. The other probabilities are scaled to add to unity. Some more details can be found at [pospoisson](#). Multiple responses are permitted.

**Value**

An object of class "vglmff" (see [vglmff-class](#)). The object is used by modelling functions such as [vglm](#), and [vgam](#).

**Author(s)**

T. W. Yee

**See Also**

[Otpospois](#), [oipospoisson](#), [simulate.vlm](#).

## Examples

```
## Not run:
odata <- data.frame(y1 = rotpospois(n = 1000, lambda = loglink(1, inverse = TRUE)))
ofit <- vglm(y1 ~ 1, otpospoisson, data = odata, trace = TRUE, crit = "c")
coef(ofit, matrix = TRUE)
Coef(ofit)
with(odata,
      hist(y1, prob = TRUE, breaks = seq(0.5, max(y1) + 0.5, by = 1),
           border = "blue"))
x <- seq(1, with(odata, max(y1)), by = 1)
with(odata, lines(x, dotpospois(x, Coef(ofit)[1]), col = "orange", type = "h", lwd = 2))
## End(Not run)
```

---

Otzeta

*One-truncated Zeta Distribution*

---

## Description

Density, distribution function, quantile function, and random generation for the one-truncated zeta distribution.

## Usage

```
dotzeta(x, shape, log = FALSE)
potzeta(q, shape, log.p = FALSE)
qotzeta(p, shape)
rotzeta(n, shape)
```

## Arguments

<code>x</code> , <code>q</code> , <code>p</code> , <code>n</code>	Same as in <a href="#">runif</a> .
<code>shape</code>	The positive shape parameter described in in <a href="#">zetaff</a> . Here it is called <code>shape</code> because it is positive.
<code>log</code> , <code>log.p</code>	Same as in <a href="#">runif</a> .

## Details

The one-truncated zeta distribution is a zeta distribution but with the probability of a one being zero. The other probabilities are scaled to add to unity. Some more details are given in [zetaff](#).

## Value

`dotzeta` gives the density, `potzeta` gives the distribution function, `qotzeta` gives the quantile function, and `rotzeta` generates random deviates.

## Note

Given some response data, the **VGAM** family function [otzeta](#) estimates the parameter `shape`.

**Author(s)**

T. W. Yee

**See Also**[Otzeta](#), [zetaff](#), [Oizeta](#).**Examples**

```

dotzeta(1:20, 0.5)
rotzeta(20, 0.5)

## Not run:  shape <- 0.8; x <- 1:10
plot(x, dotzeta(x, shape = shape), type = "h", ylim = 0:1,
      sub = "shape=0.8", las = 1, col = "blue", ylab = "Probability",
      main = "1-truncated zeta distribution: blue=PMF; orange=CDF")
lines(x + 0.1, potzeta(x, shape = shape), col = "orange", lty = 3, type = "h")
## End(Not run)

```

otzeta

*One-truncated Zeta Distribution Family Function***Description**

Estimates the parameter of the 1-truncated zeta distribution.

**Usage**

```
otzeta(lshape = "loglink", ishape = NULL, gshape = exp((-4:3)/4), zero = NULL)
```

**Arguments**

lshape, ishape, gshape, zero  
 Same as [zetaff](#).

**Details**

The 1-truncated zeta distribution is the ordinary zeta distribution but with the probability of one being 0. Thus the other probabilities are scaled up (i.e., divided by  $1 - P[Y = 1]$ ). The mean is returned by default as the fitted values. More details can be found at [zetaff](#). Multiple responses are handled.

**Value**

An object of class "vglmff" (see [vglmff-class](#)). The object is used by modelling functions such as [vglm](#), and [vgam](#).

**Author(s)**

T. W. Yee

**See Also**[Otzeta](#), [zetaff](#), [oizeta](#), [diffzeta](#), [zeta](#), [dzeta](#), [hzeta](#), [zipf](#).**Examples**

```
## Not run: odata <- data.frame(x2 = runif(nn <- 1000)) # Artificial data
odata <- transform(odata, shape = loglink(-0.25 + x2, inverse = TRUE))
odata <- transform(odata, y1 = rotzeta(nn, shape))
with(odata, table(y1))
ofit <- vglm(y1 ~ x2, otzeta, data = odata, trace = TRUE, crit = "coef")
coef(ofit, matrix = TRUE)

## End(Not run)
```

pirates1

---

*Personal data of the executed pirates associated with Bartholomew Roberts*

---

**Description**

The age, names and habitation of 52 pirates who were found guilty of piracy and executed, after the ships associated with Bartholomew Roberts were captured.

**Usage**

```
data(pirates1)
```

**Format**

A data frame with the following 3 variables.

age a numeric vector, their age in years at the time of trial. Bartholomew Roberts himself was 39 years old at his death.

name character.

habitation character.

**Details**

According to Wiki, in February 1722 Captain Ogle was sent by the British Government to find and capture the notorious pirate Bartholomew Roberts (real name: John Roberts, but also known later as Black Bart). When his warship caught up with the *Royal Fortune* he attacked and Bartholomew Roberts was the first to fall, followed by 2 others. The remaining pirates surrendered soon afterwards. A total of 272 men were captured, and of these, 65 were black, and they were sold into

slavery. The remainder were taken to Cape Coast Castle, apart from those who died on the voyage back. The trial was held in April, 1722, and 54 were condemned to death, of whom 52 were hanged and two were reprieved. Of those executed, their personal data (name, age, habitation) were recorded.

### Source

Pages 248–249 of Johnson, Captain Charles, (1955) (Editor: Arthur L. Hayward). *A General History of the Robberies and Murders of the Most Notorious Pirates*, London: Routledge and Kegan Paul Ltd. This edition was first published in 1926. The earliest manuscript of the book dates back to 1724.

This data was entered into R by Lucia Pilleri.

### See Also

[pirates2](#).

### Examples

```
summary(pirates1)
```

---

pirates2	<i>Personal data of the crew of the ship Ranger, associated with the pirate Edward Low</i>
----------	--

---

### Description

A data frame containing the age, name, birthplace and verdict of 35 members of a pirate ship associated with Edward Low, who were taken to trial on 10th to 12th July, 1723.

### Usage

```
data(pirates2)
```

### Format

The variables age and name are analogous to [pirates1](#). The variable guilty is binary and 1 means yes, 0 means not guilty. Guilty crew members were executed except for two: John Brown and Patrick Cunningham; they were respited for one year and recommended to the King's favour.

### Details

Starting on the 10th July, 1723, the crew of the *Ranger* were judged. The captain of the ship was Charles Harris, and this ship was one of two pirate ships under Captain Edward Low. Their personal data (name, age, place of birth) and verdicts are recorded in Johnson (1955). This data was constructed from pp.295–296 of that book and includes those who were not found guilty (and therefore were not executed). The execution of the 25 men were performed on 19 July near Newport, Rhode Island, USA. The notorious pirate Edward Low himself was brought to trial in 1724 under different circumstances and was hanged in Martinique.

**Source**

Same as [pirates1](#). This data was entered into R by Lucia Pilleri.

**See Also**

[pirates1](#).

**Examples**

```
summary(pirates2)
```

---

 Posbinom

*Positive-Binomial Distribution*


---

**Description**

Density, distribution function, quantile function and random generation for the positive-binomial distribution.

**Usage**

```
dposbinom(x, size, prob, log = FALSE)
pposbinom(q, size, prob)
qposbinom(p, size, prob)
rposbinom(n, size, prob)
```

**Arguments**

<code>x, q</code>	vector of quantiles.
<code>p</code>	vector of probabilities.
<code>n</code>	number of observations. Fed into <a href="#">runif</a> .
<code>size</code>	number of trials. It is the $N$ symbol in the formula given in <a href="#">posbinomial</a> and should be positive.
<code>prob</code>	probability of success on each trial. Should be in $(0, 1)$ .
<code>log</code>	See <a href="#">dbinom</a> .

**Details**

The positive-binomial distribution is a binomial distribution but with the probability of a zero being zero. The other probabilities are scaled to add to unity. The mean therefore is

$$\mu / (1 - (1 - \mu)^N)$$

where  $\mu$  is the argument `prob` above. As  $\mu$  increases, the positive-binomial and binomial distributions become more similar. Unlike similar functions for the binomial distribution, a zero value of `prob` is not permitted here.

**Value**

dposbinom gives the density, pposbinom gives the distribution function, qposbinom gives the quantile function, and rposbinom generates random deviates.

**Note**

These functions are or are likely to be deprecated. Use [Gaitdbinom](#) instead.

For dposbinom(), if arguments size or prob equal 0 then a NaN is returned.

The family function [posbinomial](#) estimates the parameters by maximum likelihood estimation.

**Author(s)**

T. W. Yee.

**See Also**

[posbinomial](#), [dposbern](#), [Gaitdbinom](#), [zabinomial](#), [zibinomial](#), [Binomial](#).

**Examples**

```

prob <- 0.2; size <- 10
table(y <- rposbinom(n = 1000, size, prob))
mean(y) # Sample mean
size * prob / (1 - (1 - prob)^size) # Population mean

(ii <- dposbinom(0:size, size, prob))
cumsum(ii) - pposbinom(0:size, size, prob) # Should be 0s
table(rposbinom(100, size, prob))

table(qposbinom(runif(1000), size, prob))
round(dposbinom(1:10, size, prob) * 1000) # Should be similar

## Not run: barplot(rbind(dposbinom(x = 0:size, size, prob),
                        dbinom(x = 0:size, size, prob)),
                  beside = TRUE, col = c("blue", "green"),
                  main = paste("Positive-binomial(", size, ", ",
                               prob, ") (blue) vs",
                               " Binomial(", size, ", ",
                               prob, ") (green)", sep = ""),
                  names.arg = as.character(0:size), las = 1)
## End(Not run)

# Simulated data example
nn <- 1000; sizeval1 <- 10; sizeval2 <- 20
pdata <- data.frame(x2 = seq(0, 1, length = nn))
pdata <- transform(pdata, prob1 = logitlink(-2 + 2 * x2, inverse = TRUE),
                  prob2 = logitlink(-1 + 1 * x2, inverse = TRUE),
                  sizev1 = rep(sizeval1, len = nn),
                  sizev2 = rep(sizeval2, len = nn))
pdata <- transform(pdata, y1 = rposbinom(nn, size = sizev1, prob = prob1),
                  y2 = rposbinom(nn, size = sizev2, prob = prob2))
with(pdata, table(y1))

```

```
with(pdata, table(y2))
# Multiple responses
fit2 <- vglm(cbind(y1, y2) ~ x2, posbinomial(multiple.responses = TRUE),
            trace = TRUE, data = pdata, weight = cbind(sizev1, sizev2))
coef(fit2, matrix = TRUE)
```

---

 Posnegbin

---

*Positive-Negative Binomial Distribution*


---

### Description

Density, distribution function, quantile function and random generation for the positive-negative binomial distribution.

### Usage

```
dposnegbin(x, size, prob = NULL, munb = NULL, log = FALSE)
pposnegbin(q, size, prob = NULL, munb = NULL,
            lower.tail = TRUE, log.p = FALSE)
qposnegbin(p, size, prob = NULL, munb = NULL)
rposnegbin(n, size, prob = NULL, munb = NULL)
```

### Arguments

`x, q` vector of quantiles.

`p` vector of probabilities.

`n` number of observations. Fed into [runif](#).

`size, prob, munb, log`  
 Same arguments as that of an ordinary negative binomial distribution (see [dnbinom](#)).  
 Some arguments have been renamed slightly.  
 Short vectors are recycled. The parameter  $1/\text{size}$  is known as a dispersion parameter; as `size` approaches infinity, the negative binomial distribution approaches a Poisson distribution.  
 Note that `prob` must lie in  $(0, 1)$ , otherwise a NaN is returned.

`log.p, lower.tail`  
 Same arguments as that of an ordinary negative binomial distribution (see [pnbinom](#)).

### Details

The positive-negative binomial distribution is a negative binomial distribution but with the probability of a zero being zero. The other probabilities are scaled to add to unity. The mean therefore is

$$\mu/(1 - p(0))$$

where  $\mu$  the mean of an ordinary negative binomial distribution.

**Value**

dposnegbin gives the density, pposnegbin gives the distribution function, qposnegbin gives the quantile function, and rposnegbin generates  $n$  random deviates.

**Note**

These functions are or are likely to be deprecated. Use [Gaitdnbinom](#) instead.

**Author(s)**

T. W. Yee

**References**

Welsh, A. H., Cunningham, R. B., Donnelly, C. F. and Lindenmayer, D. B. (1996). Modelling the abundances of rare species: statistical models for counts with extra zeros. *Ecological Modelling*, **88**, 297–308.

**See Also**

[Gaitdnbinom](#), [posnegbinomial](#), [zanegbinomial](#), [zinegbinomial](#), [rnbinom](#).

**Examples**

```
munb <- 5; size <- 4; n <- 1000
table(y <- rposnegbin(n, munb = munb, size = size))
mean(y) # sample mean
munb / (1 - (size / (size + munb))^size) # population mean
munb / pnbinom(0, mu = munb, size = size, lower.tail = FALSE) # same as before

x <- (-1):17
(ii <- dposnegbin(x, munb = munb, size = size))
max(abs(cumsum(ii) - pposnegbin(x, munb = munb, size = size))) # Should be 0

## Not run:
x <- 0:10
barplot(rbind(dposnegbin(x, munb = munb, size = size),
              dnbinom(x, mu = munb, size = size)),
        beside = TRUE, col = c("blue", "green"),
        main = paste("dposnegbin(munb = ", munb, ", size = ", size, ") (blue) vs",
                    " dnbinom(mu = ", munb, ", size = ", size, ") (green)", sep = ""),
        names.arg = as.character(x))
## End(Not run)

# Another test for pposnegbin()
nn <- 5000
mytab <- cumsum(table(rposnegbin(nn, munb = munb, size = size))) / nn
myans <- pposnegbin(sort(as.numeric(names(mytab))), munb = munb, size = size)
max(abs(mytab - myans)) # Should be 0
```

---

Pospois

*Positive-Poisson Distribution*

---

### Description

Density, distribution function, quantile function and random generation for the positive-Poisson distribution.

### Usage

```
dpospois(x, lambda, log = FALSE)
ppospois(q, lambda)
qpospois(p, lambda)
rpospois(n, lambda)
```

### Arguments

x, q	vector of quantiles.
p	vector of probabilities.
n	number of observations. Fed into <a href="#">runif</a> .
lambda	vector of positive means (of an ordinary Poisson distribution). Short vectors are recycled.
log	logical.

### Details

The positive-Poisson distribution is a Poisson distribution but with the probability of a zero being zero. The other probabilities are scaled to add to unity. The mean therefore is

$$\lambda/(1 - \exp(-\lambda)).$$

As  $\lambda$  increases, the positive-Poisson and Poisson distributions become more similar. Unlike similar functions for the Poisson distribution, a zero value of `lambda` returns a NaN.

### Value

`dpospois` gives the density, `ppospois` gives the distribution function, `qpospois` gives the quantile function, and `rpospois` generates random deviates.

### Note

These functions are or are likely to be deprecated. Use [Gaitdpois](#) instead.

The family function [pospoisson](#) estimates  $\lambda$  by maximum likelihood estimation.

### Author(s)

T. W. Yee

**See Also**

[Gaitdpois](#), [pospoisson](#), [zapoisson](#), [zipoisson](#), [rpois](#).

**Examples**

```
lambda <- 2; y = rpospois(n = 1000, lambda)
table(y)
mean(y) # Sample mean
lambda / (1 - exp(-lambda)) # Population mean

(ii <- dpospois(0:7, lambda))
cumsum(ii) - ppospois(0:7, lambda) # Should be 0s
table(rpospois(100, lambda))

table(qpospois(runif(1000), lambda))
round(dpospois(1:10, lambda) * 1000) # Should be similar

## Not run: x <- 0:7
barplot(rbind(dpospois(x, lambda), dpois(x, lambda)),
        beside = TRUE, col = c("blue", "orange"),
        main = paste("Positive Poisson(", lambda, ") (blue) vs",
                    " Poisson(", lambda, ") (orange)", sep = ""),
        names.arg = as.character(x), las = 1, lwd = 2)
## End(Not run)
```

---

 prison.us

*US Prison Data*


---

**Description**

Number of prisoners in each North American state, and the populations of those states from years 1977 to 2010

**Usage**

```
data(prison.us)
```

**Format**

A data frame with 34 observations on the following 103 variables.

**Year** a numeric vector, the year

**AL.num, AL.pop** numeric vectors

**AK.num, AK.pop, AZ.num** numeric vectors

**AZ.pop, AR.num, AR.pop** numeric vectors

**CA.num, CA.pop, CO.num** numeric vectors

**CO.pop, CT.num, CT.pop** numeric vectors

**DE.num, DE.pop, FL.num** numeric vectors  
**FL.pop, GA.num, GA.pop** numeric vectors  
**HI.num, HI.pop, ID.num** numeric vectors  
**ID.pop, IL.num, IL.pop** numeric vectors  
**IN.num, IN.pop, IA.num** numeric vectors  
**IA.pop, KS.num, KS.pop** numeric vectors  
**KY.num, KY.pop, LA.num** numeric vectors  
**LA.pop, ME.num, ME.pop** numeric vectors  
**MD.num, MD.pop, MA.num** numeric vectors  
**MA.pop, MI.num, MI.pop** numeric vectors  
**MN.num, MN.pop, MS.num** numeric vectors  
**MS.pop, MO.num, MO.pop** numeric vectors  
**MT.num, MT.pop, NE.num** numeric vectors  
**NE.pop, NV.num, NV.pop** numeric vectors  
**NH.num, NH.pop, NJ.num** numeric vectors  
**NJ.pop, NM.num, NM.pop** numeric vectors  
**NY.num, NY.pop, NC.num** numeric vectors  
**NC.pop, ND.num, ND.pop** numeric vectors  
**OH.num, OH.pop, OK.num** numeric vectors  
**OK.pop, OR.num, OR.pop** numeric vectors  
**PA.num, PA.pop, RI.num** numeric vectors  
**RI.pop, SC.num, SC.pop** numeric vectors  
**SD.num, SD.pop, TN.num** numeric vectors  
**TN.pop, TX.num, TX.pop** numeric vectors  
**UT.num, UT.pop, VT.num** numeric vectors  
**VT.pop, VA.num, VA.pop** numeric vectors  
**WA.num, WA.pop, WV.num** numeric vectors  
**WV.pop, WI.num, WI.pop** numeric vectors  
**WY.num, WY.pop** numeric vectors  
**US.pop, US.num** numeric vectors, overall counts for the whole country

### Details

This is a data set of the number of prisoners in each American state and the populations of those states, from 1977 to 2010. The number of prisoners are taken from December 31st, while the populations are estimates taken from July 1st based on the previous Census, except for pop.1980, which uses exact census data from 1980/04/01.

Warning: a scatterplot of `US.pop` shows a discontinuity around 2000.

### Source

The prisoner data was compiled from: Bureau of Justice Statistics, <http://www.bjs.gov/index.cfm>. Downloaded in September 2013 and formatted into R by J. T. Gray, [jamsgr@gmail.com](mailto:jamsgr@gmail.com).

The population data was compiled from: United States Census Bureau, <http://www.census.gov/popest/data>. Downloaded in September 2013 by J. T. Gray. This site may have become stale.

### Examples

```
summary(prison.us)
## Not run: # This plot shows a discontinuity around 2000.
plot(US.pop / 1e6 ~ Year, prison.us, main = "US population (millions)",
     las = 1, type = "b", col = "blue")
## End(Not run)
```

---

profs.nz

*Professors of Statistics in New Zealand*

---

### Description

This data set contains information on about 22 past or present professors of statistics in New Zealand universities.

### Usage

```
data(profs.nz)
```

### Format

A data frame with 22 observations on the following 7 variables.

`pubtotal` a numeric vector, the total number of publications.

`cites` a numeric vector, the number of citations.

`initials` character, first and middle and surname initials.

`Surname` character, the surname.

`firstyear` a numeric vector, the earliest indexed publication.

`ID` a numeric vector, the unique MR Author ID for each professor.

`pub1stAuthor` a numeric vector, the total number of publications which are first authored by the person.

`ARPtotal` a numeric vector, the total number of author/related publications.

`institution` character, with values "MU", "UA", "UC", "UO", "UW", "VU", the university affiliation. The abbreviations are for: Massey University, University of Auckland, University of Canterbury, University of Otago, University of Waikato and Victoria University Wellington.

**Details**

This data set contains information taken from the MathSciNet database on professors of statistics (and some related fields) affiliated with New Zealand universities.

In the future the following names may be added: C. F. Ansley, P. C. B. Phillips, B. S. Weir, C. S. Withers.

**Source**

The data is compiled from <https://mathscinet.ams.org/mathscinet/> by J. T. Gray in April 2014.

**Examples**

```

profs.nz
profs.nz[order(with(profs.nz, pubtotal), decreasing = TRUE), ]
## Not run:
plot(pub1stAuthor / pubtotal ~ pubtotal,
     main = "Professors of Statistics in NZ",
     xlab = "Number of publications in MathSciNet",
     ylab = "Proportion of first-authored papers",
     data = profs.nz, col = "blue", las = 1, type = "n")
with(profs.nz, text(pubtotal, y = pub1stAuthor / pubtotal,
                  labels = initials, col = "blue", las = 1))

## End(Not run)

```

---

rugby

*Wins, Losses and Draws Between the Top 10 Rugby Teams*


---

**Description**

The number of wins, losses and draws for each of the top 10 rugby teams against each other

**Usage**

```

data(rugby)
data(rugby.ties)

```

**Format**

The format is as two matrices.

**Details**

The first matrix is of the number of games won by each team against each of the other teams. The other matrix is the number of draws (ties) between each team. This data is current as of 2013-10-07.

**Source**

The match statistics are compiled from <http://www.rugbydata.com/> on 2013-10-07 by J. T. Gray, [jamsgr@gmail.com](mailto:jamsgr@gmail.com).

The top ten teams are determined by the International Rugby Board world rankings, <https://www.world.rugby/>.

**Examples**

```
data(rugby); data(rugby.ties)
rugby
rugby.ties
```

---

SardiniaHotels

*Data from hotels in Sardinia, Italy*

---

**Description**

This data set contains information and satisfaction scores appearing on the TripAdvisor website between the years 2008 and 2016 regarding hotels in Sardinia, Italy.

The satisfaction data refer to the reputation of hotel located along Sardinian coasts, as expressed by clients with respect to different services (e.g., breakfast, restaurant, swimming pool) offered by the hotel.

**Usage**

```
data(SardiniaHotels)
```

**Format**

A data frame with 518 rows and 43 columns (variables). Each row refers to a single hotel.

The following variables are included in the dataset:

`municipality` a factor, the municipality where the hotel is located.

`stars` an ordered factor with levels:

`1OR2stars` for 1 star or 2 star hotels,

`3stars` 3 star hotels,

`residence`,

`4stars`, 4 star hotels,

`5starsORresort`, 5 star hotels or resorts.

`area` a factor with levels related to the area of the Sardinian coast where each single hotel is located:

`AlgheroSassari`, `CagliariVillasimius`, `CostaSmeralda`, `DorgaliOrosei`, `Gallura`, `NurraAnglona`, `Ogliastra`, `Olbia`, `OristanoBosa`, `PulaChia`, `Sarabus`, `Sulcis`.

`seaLocation` a factor with levels `yes` (if the hotel is located close to the sea) and `no` (otherwise).

`excellent` a numeric vector, the number of people that expressed the highest level of satisfaction.

- good a numeric vector, the number of people that expressed a good level of satisfaction.
- average a numeric vector, the number of people that expressed an average level of satisfaction.
- bad a numeric vector, the number of people that expressed a bad level of satisfaction.
- poor a numeric vector, the number of people that expressed the lowest level of satisfaction.
- family a numeric vector, the number of people travelling with family.
- couple a numeric vector, the number of people travelling with their partner.
- single a numeric vector, the number of people travelling alone.
- business a numeric vector, the number of people travelling for work.
- MarMay a numeric vector, the number of people travelling during the period March to May.
- JunAug a numeric vector, the number of people travelling during the period June to August.
- SepNov a numeric vector, the number of people travelling during the period September to November.
- DecFeb a numeric vector, the number of people travelling during the period December to February.
- location a numeric vector, the satisfaction score expressed by tourists towards the location.
- sleepQuality a numeric vector, the satisfaction score expressed by tourists towards the sleep quality.
- room a numeric vector, the satisfaction score expressed by tourists towards the comfort and quality of the room.
- services a numeric vector, the satisfaction score expressed by tourists towards the quality of the services.
- priceQualityRate a numeric vector, the satisfaction score expressed by tourists towards ratio between price and quality.
- cleaning a numeric vector, the satisfaction score expressed by tourists towards level of room and hotel cleaning.
- bt1 a factor with levels breakfast, cleaning, location, overall, price, restaurant, room, services, staff, structure and Wi-Fi.  
It expresses the 1st most used word in reviews for a hotel.
- ratebt1 a factor with levels -1 (if the satisfaction score expressed in bt1 is prevalently negative) and 1 (if the satisfaction score expressed in bt1 is prevalently positive).
- bt2 a factor with levels breakfast, cleaning, location, overall, price, restaurant, room, services, staff, structure and Wi-Fi.  
It expresses the 2nd most used word in reviews for a hotel.
- ratebt2 a factor with levels -1 (if the satisfaction score expressed in bt2 is prevalently negative) and 1 (if the satisfaction score expressed in bt2 is prevalently positive).
- bt3 similar to bt1 and bt2, but with a corresponding different ranking.
- bt4 similar to bt1 and bt2, but with a corresponding different ranking.
- bt5 similar to bt1 and bt2, but with a corresponding different ranking.
- bt6 similar to bt1 and bt2, but with a corresponding different ranking.
- bt7 similar to bt1 and bt2, but with a corresponding different ranking.
- bt8 similar to bt1 and bt2, but with a corresponding different ranking.

bt9 similar to bt1 and bt2, but with a corresponding different ranking.  
bt10 similar to bt1 and bt2, but with a corresponding different ranking.  
ratebt3 similar to ratebt1 and ratebt2, but with a corresponding different ranking.  
ratebt4 similar to ratebt1 and ratebt2, but with a corresponding different ranking.  
ratebt5 similar to ratebt1 and ratebt2, but with a corresponding different ranking.  
ratebt6 similar to ratebt1 and ratebt2, but with a corresponding different ranking.  
ratebt7 similar to ratebt1 and ratebt2, but with a corresponding different ranking.  
ratebt8 similar to ratebt1 and ratebt2, but with a corresponding different ranking.  
ratebt9 similar to ratebt1 and ratebt2, but with a corresponding different ranking.  
ratebt10 similar to ratebt1 and ratebt2, but with a corresponding different ranking.

### Details

These data were manually collected during March–June 2016 by students of the class of "Statistics for Tourism" at the University of Cagliari, Italy (Bachelor's degree in Tourism Economics and Management), under the supervision of Prof. Claudio Conversano and Dr. Giulia Contu.

Many of the variables fall into several natural groups, e.g., [municipality, stars, area, seaLocation]; [excellent, good, average, bad, poor]; [MarMay, JunAug, SepNov, DecFeb]; [family, couple, single, business]; [location,...cleaning]; [bt1,...bt10]; [ratebt1,...ratebt10].

### Source

TripAdvisor, <https://www.tripadvisor.it/>.

### Examples

```
data(SardiniaHotels)
summary(SardiniaHotels)
```

---

students.tw

*Taiwanese students answer a multiple response question*

---

### Description

This data is a subset from a survey of 49609 first-year college students in Taiwan collected in the year 2003 about their preferences for college study.

### Usage

```
data(students.tw)
```

**Format**

A data frame with 49609 observations on the following 12 response variables. For binary variables, a "1" means yes, and "0" means no. See below for exact wording (translated from the original Chinese).

ID a numeric vector, a unique identification number for each student in the survey.

read Read Chinese and foreign classics.

t.travel Travel around Taiwan.

conference Present academic papers in conferences.

act.leader Lead large-scale activities.

team Be on a school team.

stu.leader Be a student association leader.

intern Participate internship programs.

love Fall in love.

sex Have sexual experience.

o.travel Travel around the world.

friends Make many friends.

other Other experience which is not included in the survey.

**Details**

This data frame is a subset of a larger data set where any student with any missing value was deleted. The remaining data set contains of 32792 students. Unfortunately, other variables such as age and sex were not made available.

Each student was asked the following multiple response question.

Question : What kind of experience do you expect to receive during the period of college study? (Select at least one response)

1. Read Chinese and foreign classics
2. Travel around Taiwan
3. Present academic papers in conferences
4. Lead large-scale activities
5. Be on a school team
6. Be a student association leader
7. Participate internship programs
8. Fall in love
9. Have sexual experience
10. Travel around the world
11. Make many friends
12. Other

**Source**

Originally, the data set for was downloaded from a survey center of Academia Sinica <https://srda.sinica.edu.tw/news>. It now seems unavailable.

**References**

Wang, H. and Huang, W. H. (2013) Bayesian Ranking Responses in Multiple Response Questions. *Journal of the Royal Statistical Society, Series A*, (to appear).

Help from Viet Hoang Quoc is gratefully acknowledged.

**Examples**

```
data(students.tw)
summary(students.tw)

with(students.tw, table(love, sex))
## Not run:
plot(jitter(sex) ~ jitter(love), data = students.tw, col = "blue",
      main = "Taiwanese students")

## End(Not run)
```

---

Tikuv

*A Short-tailed Symmetric Distribution*


---

**Description**

Density, cumulative distribution function, quantile function and random generation for the short-tailed symmetric distribution of Tiku and Vaughan (1999).

**Usage**

```
dtikuv(x, d, mean = 0, sigma = 1, log = FALSE)
ptikuv(q, d, mean = 0, sigma = 1, lower.tail = TRUE, log.p = FALSE)
qtikuv(p, d, mean = 0, sigma = 1, lower.tail = TRUE, log.p = FALSE, ...)
rtikuv(n, d, mean = 0, sigma = 1, Smallno = 1.0e-6)
```

**Arguments**

x, q	vector of quantiles.
p	vector of probabilities.
n	number of observations. Same as in <a href="#">runif</a> .
d, mean, sigma	arguments for the parameters of the distribution. See <a href="#">tikuv</a> for more details. For <a href="#">rtikuv</a> , arguments mean and sigma must be of length 1.
Smallno	Numeric, a small value used by the rejection method for determining the lower and upper limits of the distribution. That is, $ptikuv(L) < Smallno$ and $ptikuv(U) > 1 - Smallno$ where L and U are the lower and upper limits respectively.

... Arguments that can be passed into [uniroot](#).  
 log Logical. If log = TRUE then the logarithm of the density is returned.  
 lower.tail, log.p Same meaning as in [pnorm](#) or [qnorm](#).

### Details

See [tikuv](#) for more details.

### Value

`dtikuv` gives the density, `ptikuv` gives the cumulative distribution function, `qtikuv` gives the quantile function, and `rtikuv` generates random deviates.

### Author(s)

T. W. Yee and Kai Huang

### See Also

[tikuv](#).

### Examples

```
## Not run: par(mfrow = c(2, 1))
x <- seq(-5, 5, len = 401)
plot(x, dnorm(x), type = "l", col = "black", ylab = "", las = 1,
      main = "Black is standard normal, others are dtikuv(x, d)")
lines(x, dtikuv(x, d = -10), col = "orange")
lines(x, dtikuv(x, d = -1 ), col = "blue")
lines(x, dtikuv(x, d = 1 ), col = "green")
legend("topleft", col = c("orange","blue","green"), lty = rep(1, len = 3),
      legend = paste("d =", c(-10, -1, 1)))

plot(x, pnorm(x), type = "l", col = "black", ylab = "", las = 1,
      main = "Black is standard normal, others are ptikuv(x, d)")
lines(x, ptikuv(x, d = -10), col = "orange")
lines(x, ptikuv(x, d = -1 ), col = "blue")
lines(x, ptikuv(x, d = 1 ), col = "green")
legend("topleft", col = c("orange","blue","green"), lty = rep(1, len = 3),
      legend = paste("d =", c(-10, -1, 1)))
## End(Not run)

probs <- seq(0.1, 0.9, by = 0.1)
ptikuv(qtikuv(p = probs, d = 1), d = 1) - probs # Should be all 0
```

tikuv

*Short-tailed Symmetric Distribution Family Function***Description**

Fits the short-tailed symmetric distribution of Tiku and Vaughan (1999).

**Usage**

```
tikuv(d, lmean = "identitylink", lsigma = "loglink", isigma = NULL,
      zero = "sigma")
```

**Arguments**

<code>d</code>	The $d$ parameter. It must be a single numeric value less than 2. Then $h = 2 - d > 0$ is another parameter.
<code>lmean, lsigma</code>	Link functions for the mean and standard deviation parameters of the usual univariate normal distribution (see <b>Details</b> below). They are $\mu$ and $\sigma$ respectively. See <a href="#">Links</a> for more choices.
<code>isigma</code>	Optional initial value for $\sigma$ . A NULL means a value is computed internally.
<code>zero</code>	A vector specifying which linear/additive predictors are modelled as intercept-only. The values can be from the set {1,2}, corresponding respectively to $\mu$ , $\sigma$ . If <code>zero = NULL</code> then all linear/additive predictors are modelled as a linear combination of the explanatory variables. For many data sets having <code>zero = 2</code> is a good idea. See <a href="#">CommonVGAMffArguments</a> for information.

**Details**

The short-tailed symmetric distribution of Tiku and Vaughan (1999) has a probability density function that can be written

$$f(y) = \frac{K}{\sqrt{2\pi}\sigma} \left[ 1 + \frac{1}{2h} \left( \frac{y - \mu}{\sigma} \right)^2 \right]^2 \exp \left( -\frac{1}{2} (y - \mu)^2 / \sigma^2 \right)$$

where  $h = 2 - d > 0$ ,  $K$  is a function of  $h$ ,  $-\infty < y < \infty$ ,  $\sigma > 0$ . The mean of  $Y$  is  $E(Y) = \mu$  and this is returned as the fitted values.

**Value**

An object of class "vglmff" (see [vglmff-class](#)). The object is used by modelling functions such as [vglm](#), and [vgam](#).

**Warning**

Under- or over-flow may occur if the data is ill-conditioned, e.g., when  $d$  is very close to 2 or approaches  $-\text{Inf}$ .

**Note**

The density function is the product of a univariate normal density and a polynomial in the response  $y$ . The distribution is bimodal if  $d > 0$ , else is unimodal. A normal distribution arises as the limit as  $d$  approaches  $-\infty$ , i.e., as  $h$  approaches  $\infty$ . Fisher scoring is implemented. After fitting the value of  $d$  is stored in @misc with component name  $d$ .

**Author(s)**

Thomas W. Yee

**References**

- Akkaya, A. D. and Tiku, M. L. (2008). Short-tailed distributions and inliers. *Test*, **17**, 282–296.
- Tiku, M. L. and Vaughan, D. C. (1999). A family of short-tailed symmetric distributions. *Technical report, McMaster University, Canada*.

**See Also**

[dtikuv](#), [uninormal](#).

**Examples**

```
m <- 1.0; sigma <- exp(0.5)
tdata <- data.frame(y = rtikuv(n = 1000, d = 1, m = m, s = sigma))
tdata <- transform(tdata, sy = sort(y))
fit <- vglm(y ~ 1, tikuv(d = 1), data = tdata, trace = TRUE)
coef(fit, matrix = TRUE)
(Cfit <- Coef(fit))
with(tdata, mean(y))
## Not run: with(tdata, hist(y, prob = TRUE))
lines(dtikuv(sy, d = 1, m = Cfit[1], s = Cfit[2]) ~ sy, data = tdata, col = "orange")
## End(Not run)
```

---

trap0

*Trout Data at the Te Whaiiau Trap on Lake Otamangakau*

---

**Description**

Rainbow and brown trout trapped at the Te Whaiiau Trap at Lake Otamangakau in the central North Island of New Zealand. The data were collected by the Department of Conservation.

**Usage**

```
data(trap0)
```

## Format

A data frame with 1226 observations on the following 15 variables.

**Date** Date as a class "Date" variable.

**BFTW, BMTW, RFTW, RMTW** numeric vectors, the number of fish trapped daily. B/R is for brown/rainbow trout. F/M is for female/male. TW is for the Te Whaiau trap location (there was another trap just off the Tongariro River).

**MinAT, MaxAT** numeric vectors, daily minimum and maximum ambient temperatures in Celsius.

**Rain** numeric vector, daily rainfall that has been scaled between 0 (none) and 100 (flooding situation).

**LevelTW** numeric vector, water level of the stream that has been scaled between 0 (none) and 100 (flooding situation). In a flooding situation it is possible that some fish going upstream were not caught.

**Year, Month, Day** numeric vectors, extracted from Date.

**doy** a numeric vector, Julian day of year. The value 1 means 1st of January, and so on up to 365.

**f.Year** a factor vector, the year as a factor.

**fict.Year** similar to Date but a fictional year is used for all the data. This allows all the data to be plotted along one calendar year.

## Details

These are the daily numbers of fish trapped at the Te Whaiau trap near Lake Otamangakau, during the winter months when spawning is at its peak. These fish were all going upstream. There are two species of trout, split up by males and females, in the data set. The first is brown trout (*Salmo trutta*) and the second is rainbow trout (*Oncorhynchus mykiss*). Information on the movement patterns of brown and rainbow trout in Lake Otamangakau and Lake Te Whaiau can be found in Dedual et al. (2000).

Brown trout are more sedentary compared with rainbow trout, and spawning activities of brown trout occur between May and June whilst peak spawning of rainbow trout occurs between July and August. Furthermore, brown trout have been observed avoiding water above 19 degrees Celsius and optimum temperatures for growth are between 10–15 degrees for brown trout and 16.5–17.2 degrees for rainbow trout.

See also [lake0](#).

## Source

Many thanks to Dr Michel Dedual (<http://www.doc.govt.nz>) for making this data available. Help from Simeon Pattenwise is also acknowledged.

## References

Dedual, M. and Maxwell, I. D. and Hayes, J. W. and Strickland, R. R. (2000). Distribution and movements of brown (*Salmo trutta*) and rainbow trout (*Oncorhynchus mykiss*) in Lake Otamangakau, central North Island, New Zealand. *New Zealand Journal of Marine and Freshwater Research*, **34**: 615–627.

**Examples**

```
data("trap0")
summary(trap0)
```

---

 tube10
 

---



---

*London Underground (Tube) Passenger Counts for November 2010*


---

**Description**

The data set contains counts of the number of passengers entering London Underground stations (also known as *the Tube*) on a typical day of November 2010 in quarter-hour time blocks.

**Usage**

```
data("tube10")
```

**Format**

A data frame with 100 observations on the following 270 variables.

ActonTown a numeric vector  
 Aldgate a numeric vector  
 AldgateEast a numeric vector  
 Alperton a numeric vector  
 Amersham a numeric vector  
 Angel a numeric vector  
 Archway a numeric vector  
 ArnosGrove a numeric vector  
 Arsenal a numeric vector  
 BakerStreet a numeric vector  
 Balham a numeric vector  
 BankAndMonument a numeric vector  
 Barbican a numeric vector  
 Barking a numeric vector  
 Barkingside a numeric vector  
 BaronsCourt a numeric vector  
 Bayswater a numeric vector  
 Becontree a numeric vector  
 BelsizePark a numeric vector  
 Bermondsey a numeric vector  
 BethnalGreen a numeric vector

Blackfriars a numeric vector  
BlackhorseRoad a numeric vector  
BondStreet a numeric vector  
Borough a numeric vector  
BostonManor a numeric vector  
BoundsGreen a numeric vector  
BowRoad a numeric vector  
BrentCross a numeric vector  
Brixton a numeric vector  
BromleyByBow a numeric vector  
BuckhurstHill a numeric vector  
BurntOak a numeric vector  
CaledonianRoad a numeric vector  
CamdenTown a numeric vector  
CanadaWater a numeric vector  
CanaryWharf a numeric vector  
CanningTown a numeric vector  
CannonStreet a numeric vector  
CanonsPark a numeric vector  
ChalfontAndLatimer a numeric vector  
ChalkFarm a numeric vector  
ChanceryLane a numeric vector  
CharingCross a numeric vector  
Chesham a numeric vector  
Chigwell a numeric vector  
ChiswickPark a numeric vector  
Chorleywood a numeric vector  
ClaphamCommon a numeric vector  
ClaphamNorth a numeric vector  
ClaphamSouth a numeric vector  
Cockfosters a numeric vector  
Colindale a numeric vector  
ColliersWood a numeric vector  
CoventGarden a numeric vector  
Croxley a numeric vector  
DagenhamEast a numeric vector  
DagenhamHeathway a numeric vector

Debden a numeric vector  
DollisHill a numeric vector  
EalingBroadway a numeric vector  
EalingCommon a numeric vector  
EarlsCourt a numeric vector  
EastActon a numeric vector  
EastFinchley a numeric vector  
EastHam a numeric vector  
EastPutney a numeric vector  
Eastcote a numeric vector  
Edgware a numeric vector  
EdgwareRoadBak a numeric vector  
EdgwareRoadCir a numeric vector  
ElephantAndCastle a numeric vector  
ElmPark a numeric vector  
Embankment a numeric vector  
Epping a numeric vector  
Euston a numeric vector  
EustonSquare a numeric vector  
Fairlop a numeric vector  
Farringdon a numeric vector  
FinchleyCentral a numeric vector  
FinchleyRoad a numeric vector  
FinsburyPark a numeric vector  
FulhamBroadway a numeric vector  
GantsHill a numeric vector  
GloucesterRoad a numeric vector  
GoldersGreen a numeric vector  
GoldhawkRoad a numeric vector  
GoodgeStreet a numeric vector  
GrangeHill a numeric vector  
GreatPortlandStreet a numeric vector  
GreenPark a numeric vector  
Greenford a numeric vector  
Gunnersbury a numeric vector  
Hainault a numeric vector  
HammersmithDis a numeric vector

HammersmithHC a numeric vector  
Hampstead a numeric vector  
HangerLane a numeric vector  
Harlesden a numeric vector  
HarrowAndWealdstone a numeric vector  
HarrowOnTheHill a numeric vector  
HattonCross a numeric vector  
HeathrowTerminals123 a numeric vector  
HeathrowTerminal4 a numeric vector  
HeathrowTerminal5 a numeric vector  
HendonCentral a numeric vector  
HighBarnet a numeric vector  
HighStreetKensington a numeric vector  
HighburyAndIslington a numeric vector  
Highgate a numeric vector  
Hillingdon a numeric vector  
Holborn a numeric vector  
HollandPark a numeric vector  
HollowayRoad a numeric vector  
Hornchurch a numeric vector  
HounslowCentral a numeric vector  
HounslowEast a numeric vector  
HounslowWest a numeric vector  
HydeParkCorner a numeric vector  
Ickenham a numeric vector  
Kennington a numeric vector  
KensalGreen a numeric vector  
KensingtonOlympia a numeric vector  
KentishTown a numeric vector  
Kenton a numeric vector  
KewGardens a numeric vector  
Kilburn a numeric vector  
KilburnPark a numeric vector  
KingsCrossStPancras a numeric vector  
Kingsbury a numeric vector  
Knightsbridge a numeric vector  
LadbrokeGrove a numeric vector

LambethNorth a numeric vector  
LancasterGate a numeric vector  
LatimerRoad a numeric vector  
LeicesterSquare a numeric vector  
Leyton a numeric vector  
Leytonstone a numeric vector  
LiverpoolStreet a numeric vector  
LondonBridge a numeric vector  
Loughton a numeric vector  
MaidaVale a numeric vector  
ManorHouse a numeric vector  
MansionHouse a numeric vector  
MarbleArch a numeric vector  
Marylebone a numeric vector  
MileEnd a numeric vector  
MillHillEast a numeric vector  
MoorPark a numeric vector  
Moorgate a numeric vector  
Morden a numeric vector  
MorningsonCrescent a numeric vector  
Neasden a numeric vector  
NewburyPark a numeric vector  
NorthActon a numeric vector  
NorthEaling a numeric vector  
NorthGreenwich a numeric vector  
NorthHarrow a numeric vector  
NorthWembley a numeric vector  
Northfields a numeric vector  
Northolt a numeric vector  
NorthwickPark a numeric vector  
Northwood a numeric vector  
NorthwoodHills a numeric vector  
NottingHillGate a numeric vector  
Oakwood a numeric vector  
OldStreet a numeric vector  
Osterley a numeric vector  
Oval a numeric vector

OxfordCircus a numeric vector  
Paddington a numeric vector  
ParkRoyal a numeric vector  
ParsonsGreen a numeric vector  
Perivale a numeric vector  
PiccadillyCircus a numeric vector  
Pimlico a numeric vector  
Pinner a numeric vector  
Plaistow a numeric vector  
PrestonRoad a numeric vector  
PutneyBridge a numeric vector  
QueensPark a numeric vector  
Queensbury a numeric vector  
Queensway a numeric vector  
RavenscourtPark a numeric vector  
RaynersLane a numeric vector  
Redbridge a numeric vector  
RegentsPark a numeric vector  
Richmond a numeric vector  
Rickmansworth a numeric vector  
RodingValley a numeric vector  
RoyalOak a numeric vector  
Ruislip a numeric vector  
RuislipGardens a numeric vector  
RuislipManor a numeric vector  
RussellSquare a numeric vector  
SevenSisters a numeric vector  
ShepherdsBushCen a numeric vector  
ShepherdsBushHC a numeric vector  
SloaneSquare a numeric vector  
Snaresbrook a numeric vector  
SouthEaling a numeric vector  
SouthHarrow a numeric vector  
SouthKensington a numeric vector  
SouthKenton a numeric vector  
SouthRuislip a numeric vector  
SouthWimbledon a numeric vector

SouthWoodford a numeric vector  
Southfields a numeric vector  
Southgate a numeric vector  
Southwark a numeric vector  
StJameessPark a numeric vector  
StJohnsWood a numeric vector  
StPauls a numeric vector  
StamfordBrook a numeric vector  
Stanmore a numeric vector  
StepneyGreen a numeric vector  
Stockwell a numeric vector  
StonebridgePark a numeric vector  
Stratford a numeric vector  
SudburyHill a numeric vector  
SudburyTown a numeric vector  
SwissCottage a numeric vector  
Temple a numeric vector  
TheydonBois a numeric vector  
TootingBec a numeric vector  
TootingBroadway a numeric vector  
TottenhamCourtRoad a numeric vector  
TottenhamHale a numeric vector  
TotteridgeAndWhetstone a numeric vector  
TowerHill a numeric vector  
TufnellPark a numeric vector  
TurnhamGreen a numeric vector  
TurnpikeLane a numeric vector  
Upminster a numeric vector  
UpminsterBridge a numeric vector  
Upney a numeric vector  
UptonPark a numeric vector  
Uxbridge a numeric vector  
Vauxhall a numeric vector  
Victoria a numeric vector  
WalthamstowCentral a numeric vector  
Wanstead a numeric vector  
WarrenStreet a numeric vector

WarwickAvenue a numeric vector  
Waterloo a numeric vector  
Watford a numeric vector  
WembleyCentral a numeric vector  
WembleyPark a numeric vector  
WestActon a numeric vector  
WestBrompton a numeric vector  
WestFinchley a numeric vector  
WestHam a numeric vector  
WestHampstead a numeric vector  
WestHarrow a numeric vector  
WestKensington a numeric vector  
WestRuislip a numeric vector  
WestbournePark a numeric vector  
Westminster a numeric vector  
WhiteCity a numeric vector  
Whitechapel a numeric vector  
WillesdenGreen a numeric vector  
WillesdenJunction a numeric vector  
Wimbledon a numeric vector  
WimbledonPark a numeric vector  
WoodGreen a numeric vector  
WoodLane a numeric vector  
Woodford a numeric vector  
WoodsidePark a numeric vector  
Total a numeric vector; the total over all stations.  
mins24 a numeric vector; minutes on a 24 hour clock, e.g., 0 is midnight, 120 is 2am.

### Details

Each cell contains the number of passengers entering a station during a quarter-hour period of time on a typical day during November 2010. The column names of the data frame are the station names and the most of the rows are the start time of each quarter-hour time block given in 24 hour time, e.g., 2215 means 10:15pm to 10:29pm. The last four rows are "Total", "A.M.Peak", "Interpeak", "P.M.Peak".

The data is adjusted to remove the effect of abnormal circumstances that many affect passenger numbers such as industrial action.

### Source

The data came from the UK Government Transport for London website <https://www.tfl.gov.uk>. Downloaded in December 2013 and formatted into R by J. T. Gray (and slightly edited by T. W. Yee).

**Examples**

```
## Not run:
data(tube10)
waterloo <- tube10[1:(4*24), "Waterloo"] # Omit the totals and the peaks
barplot(unlist(waterloo))
barplot(log10(1 + unlist(waterloo)), col = "lightblue",
        ylab = "log10(1+.)", las = 1)

## End(Not run)
```

---

 ugss

---

*Undergraduate Statistics Students Lifestyle Questionnaire*


---

**Description**

About 800 students studying undergraduate statistics were asked many lifestyle questions.

**Usage**

```
data(ugss)
```

**Format**

A data frame with 804 observations on the following 29 variables.

sex Gender, a factor, (female or male)

age age in years, a numeric vector

eyes eye colour, a factor, (blue, brown, green, hazel or other)

piercings Number of body piercings, a numeric vector

pierced Any body piercings? a factor, (Yes or No)

tattoos Number of tattoos, a numeric vector

tattooed Any tattoos? a factor, (Yes or No)

glasses Wears glasses etc.? a factor, (Yes or No)

sleep Average number of hours of sleep per night, a numeric vector

study Average number of hours of study per week, a numeric vector

tv Average number of hours watching TV per week, a numeric vector

movies Number of movies seen at a cinema during the last 3 months, a numeric vector

movies3m Seen movies in last 3 months? a factor, (Yes or No)

sport Favourite sport, a factor, about 19 of them

entertainment Favourite entertainment, a factor, about 15 of them

fruit Favourite fruit a factor, about 13 of them

income Average income during semester per week, a numeric vector

rent Amount spent on rent or room and board per week, a numeric vector  
 clothes Average amount spent on clothes per month, a numeric vector  
 hair Average cost to get a hair-cut, a numeric vector  
 tobacco Average amount spent on tobacco per week, a numeric vector  
 smokes Smokes? a factor, (Yes or No)  
 alcohol Average amount spent on alcohol per week, a numeric vector  
 buy.alcohol Buys (purchases) alcohol? a factor, (Yes or No)  
 sendtxt Average number text messages sent per day, a numeric vector.  
 receivetxt Average number text messages received per day, a numeric vector.  
 txts Uses text messaging? a factor, (Yes or No)  
 country Country of birth, a factor, about 54 of them  
 status Student status, a factor, (International, NZ.Citizen, NZ.Resident)

### Details

This data was collected online and anonymously in 2010. The respondents were students studying an undergraduate statistics course at a New Zealand university. Possibly there are duplicate students (due to failing and re-enrolling). All monies are in NZD. Note the data has had minimal checking. Most numerical variables tend to have measurement error, and all of them happen to be all integer-valued.

### Examples

```
summary(ugss)
```

---

vtinpat	<i>Vermont Hospital Inpatient Data</i>
---------	--

---

### Description

Information on inpatients discharged from hospitals in Vermont, USA, 2012.

### Usage

```
data(vtinpat)
```

### Format

A data frame with 52206 observations on the following 7 variables.

hospital a factor with levels 1 = Northwestern Medical Center, 2 = North Country Hospital and Health Center, 3 = Northeastern Vermont Regional Hospital, 4 = Copley Hospital, 5 = Fletcher Allen Health Care, 6 = Central Vermont Hospital, 8 = Rutland Regional Medical Center, 9 = Porter Medical Center, 10 = Gifford Memorial Hospital, 11 = Mount Ascutney Hospital and Health Center, 12 = Springfield Hospital, 14 = Grace Cottage Hospital, 15 = Brattleboro Memorial Hospital, 16 = Southwestern Vermont Medical Center

`admit` a factor with levels 1 = Emergency, 2 = Urgent, 3 = Elective, 4, Newborn, 5 = Trauma

`age.group` a factor with levels 1 = Under 1, 2 = 1-17, 3 = 18-24, 4 = 25-29, 5 = 30-34, 6 = 35-39, 7 = 40-44, 8 = 45-49, 9 = 50-54, 10 = 55-59, 11 = 60-64, 12 = 65-69, 13 = 70-74, 14 = 75+

`sex` a factor with levels 1 = Male, 2 = Female

`discharge` a factor with levels 1 = To another medical facility, 2 = home, 3 = against medical advice, 4 = Died, 5 = To court or law enforcement, 6 = still a patient

`diagnosis` a factor with levels 1 = Brain And C.N.S., 2 = Eye, 3 = Ear, Nose & Throat, 4 = Respiratory, 5 = Heart & Circulatory, 6 = Digestive, 7 = Liver & Pancreas, 8 = Musculoskeletal, 9 = Skin and Breast, 10 = Endocrine, 11 = Kidney & Urinary, 12 = Male Reproductive, 13 = Female Reproductive, 14 = Pregnancy, Childbirth, 15 = Neonatal, 16 = Spleen & Blood, 17 = Lymphatic, 18 = Infection, 19 = Mental Illness, 20 = Substance Abuse, 21 = Injury, Toxic Effects, 22 = Burns, 23 = Other, 24 = Trauma, 25 = H.I.V.

`los` a numeric vector, number of days spent in hospital

### Details

This data set contains details on inpatients discharged from hospitals in Vermont, USA, in 2012 as part of the Vermont Uniform Hospital Discharge Data Set. The Vermont Department of Financial Regulation administers this program and the Vermont Department of Health manages the data set.

### Source

Vermont department of Health, <https://www.healthvermont.gov/stats/systems> formatted into R by J. T. Gray in mid-2014.

### Examples

```
summary(vtinpat)
```

---

wffc

*2008 World Fly Fishing Championships Data*

---

### Description

Capture records of the 2008 FIPS-MOUCHE World Fly Fishing Championships held in Rotorua, New Zealand during 22–30 March 2008.

### Usage

```
data(wffc)
```

## Format

A data frame with 4267 observations on the following 8 variables. Each row is a recorded capture.

`length` a numeric vector; length of fish in mm.

`water` a factor with levels Waihou, Waimakariri, Whanganui, Otamangakau, Rotoaira. These are known as Sectors IV, V, I, II, III respectively, and are also represented by the variable `sector`.

`session` a numeric vector; a value from the set 1,2,...,6. These are time ordered, and there were two sessions per competition day.

`sector` a numeric vector; a value from the set 1,2,...,5. Ideally these should be converted to a factor.

`beatboat` a numeric vector; beat or boat number, a value from the set 1,2,...,19. Ideally these should be converted to a factor. For a river though, they are contiguous whereas on a lake it is less so.

`comid` a numeric vector; the competitor's ID number. Uniquely identifies each competitor. These ID numbers actually correspond to their rankings in the individual level.

`iname` a character vector; the individual competitor's name.

`country` a character vector; what country the competitors represented. The countries represented were Australia (AUS), Canada (CAN), Croatia (CRO), Czech Republic (CZE), England (ENG), Finland (FIN), France (FRA), Holland (NED), Ireland (IRE), Italy (ITA), Japan (JPN), Malta (MAL), New Zealand (NZL), Poland (POL), Portugal (POR), South Africa (RSA), Slovakia (SVK), USA (USA), Wales (WAL).

## Details

Details may be obtained at Yee (2010) and Yee (2014). Here is a brief summary. The three competition days were 28–30 March. Each session was fixed at 9.00am–12.00pm and 2.30–5.30pm daily. One of the sessions was a rest session. Each of 19 teams had 5 members, called A, B, C, D and E (there was a composite team, actually). The scoring system allocated 100 points to each eligible fish (minimum length was 18 cm) and 20 points for each cm of its length (rounded up to the nearest centimeter). Thus a 181mm or 190mm fish was worth 480 points. Each river was divided into 19 contiguous downstream beats labelled 1,2,...,19. Each lake was fished by 9 boats, each with two competitors except for one boat which only had one. Each competitor was randomly assigned to a beat/boat.

Competitors were ranked according to their placings at each sector-session combination, and then these placings were summed. Those with the minimum total placings were the winners, thus it was not necessarily those who had the maximum points who won. For example, in Session 1 at the Waihou River, each of the 19 competitors was ranked 1 (best) to 19 (worst) according to the point system. This is the “placing” for that session. These placings were added up over the 5 sessions to give the “total placings”.

All sectors have naturally wild Rainbow trout (*Oncorhynchus mykiss*) while Lake Otamangakau and the Whanganui River also holds Brown trout (*Salmo trutta*). Only these two species were targeted. The species was not recorded electronically, however a post-analysis of the paper score sheets from the two lakes showed that, approximately, less than 5 percent were Brown trout. It may be safely assumed that all the Waihou and Waimakariri fish were Rainbow trout. The gender of the fish were also not recorded electronically, and anyway, distinguishing between male and female was very difficult for small fish.

Although species and gender data were supposed to have been collected at the time of capture the quality of these variables is rather poor and furthermore they were not recorded electronically.

Note that some fish may have been caught more than once, hence these data do not represent individual fish but rather recorded captures.

Note also that a few internal discrepancies may be found within and between the data frames `wffc`, `wffc.nc`, `wffc.indiv`, `wffc.teams`. This is due to various reasons, such as competitors being replaced by reserves when sick, fish that were included or excluded upon the local judge's decision, competitors who fished two hours instead of three by mistake, etc. The data has already been cleaned of errors and internal inconsistencies but a few may remain.

### Source

This data frame was adapted from the WFFC's spreadsheet. Special thanks goes to Paul Dewar, Jill Mandeno, Ilkka Pirinen, and the other members of the Organising Committee of the 28th FIPS-Mouche World Fly Fishing Championships for access to the data. The assistance and feedback of Colin Shepherd is gratefully acknowledged.

### References

Yee, T. W. (2010). VGLMs and VGAMs: an overview for applications in fisheries research. *Fisheries Research*, **101**, 116–126.

Yee, T. W. (2014). Scoring rules, and the role of chance: analysis of the 2008 World Fly Fishing Championships. *Journal of Quantitative Analysis in Sports*. **10**, 397–409.

### See Also

`wffc.indiv`, `wffc.teams`, `wffc.nc`, `wffc.P1`, `lake0`.

### Examples

```
summary(wffc)
with(wffc, table(water, session))

# Obtain some simple plots
waihou <- subset(wffc, water == "Waihou")
waimak <- subset(wffc, water == "Waimakariri")
whang <- subset(wffc, water == "Whanganui")
otam <- subset(wffc, water == "Otamangakau")
roto <- subset(wffc, water == "Rotoaira")
minlength <- min(wffc[, "length"])
maxlength <- max(wffc[, "length"])
nwater <- c("Waihou" = nrow(waihou), "Waimakariri" = nrow(waimak),
           "Whanganui" = nrow(whang), "Otamangakau" = nrow(otam),
           "Rotoaira" = nrow(roto))

## Not run:
par(mfrow = c(2, 3), las = 1)
# Overall distribution of length
with(wffc, boxplot(length/10 ~ water, ylim = c(minlength, maxlength)/10,
                  border = "blue", main = "Length (cm)", cex.axis = 0.5))
```

```

# Overall distribution of LOG length
with(wffc, boxplot(length/10 ~ water, ylim = c(minlength, maxlength)/10,
  border = "blue", log = "y", cex.axis = 0.5,
  main = "Length (cm) on a log scale"))

# Overall distribution of number of captures
pie(nwater, border = "blue", main = "Proportion of captures",
  labels = names(nwater), density = 10, col = 1:length(nwater),
  angle = 85+30* 1:length(nwater))

# Overall distribution of number of captures
with(wffc, barplot(nwater, main = "Number of captures", cex.names = 0.5,
  col = "lightblue"))

# Overall distribution of proportion of number of captures
with(wffc, barplot(nwater / sum(nwater), cex.names = 0.5, col = "lightblue",
  main = "Proportion of captures"))

# An interesting lake
with(roto, hist(length/10, xlab = "Fish length (cm)", col = "lightblue",
  breaks = seq(18, 70, by = 3), prob = TRUE, ylim = c(0, 0.08),
  border = "blue", ylab = "", main = "Lake Rotoaira", lwd = 2))

## End(Not run)

```

---

wffc.indiv

2008 World Fly Fishing Championships (Individual results) Data

---

## Description

Individual competitors' results of the 2008 FIPS-MOUCHE World Fly Fishing Championships held in Rotorua, New Zealand during 22–30 March 2008.

## Usage

```
data(wffc.indiv)
```

## Format

A data frame with 99 observations on the following 8 variables. Some of these variable are described in [wffc](#).

`totalPlacings` a numeric vector; these are the summed placings over the 5 sessions.

`points` a numeric vector.

`noofcaptures` a numeric vector.

`longest` a numeric vector.

`individual` a numeric vector; did the competitor fish in a team or as an individual? (one team was made of composite countries due to low numbers).

`country` a character vector.

`iname` a character vector.

`comid` a numeric vector.

## Details

This data frame gives the individual results of the competition. See also [wffc](#) and [wffc.teams](#) for more details and links.

## References

Yee, T. W. (2010). VGLMs and VGAMs: an overview for applications in fisheries research. *Fisheries Research*, **101**, 116–126.

## Examples

```
summary(wffc.indiv)
```

---

wffc.nc

*2008 World Fly Fishing Championships (Number of captures) Data*

---

## Description

Number of captures in the 2008 FIPS-MOUCHE World Fly Fishing Championships held in Rotorua, New Zealand during 22–30 March 2008.

## Usage

```
data(wffc.nc)
```

## Format

A data frame with 475 observations on the following 7 variables. Most of these variable are described in [wffc](#). Each row is sorted by sector, session and beat.

sector a numeric vector.

session a numeric vector.

beatboat a numeric vector.

numbers a numeric vector.

comid a numeric vector.

iname a character vector.

country a character vector.

## Details

This data frame was obtained by processing [wffc](#). The key variable is numbers, which is sector-session-beat specific.

Note that some fish may have been caught more than once, hence these data do not represent individual fish.

**References**

Yee, T. W. (2010). VGLMs and VGAMs: an overview for applications in fisheries research. *Fisheries Research*, **101**, 116–126.

**See Also**

[DeLury, lake0.](#)

**Examples**

```
xtabs( ~ sector + session, wffc.nc)
```

---

wffc.points

*Point System for the 2008 World Fly Fishing Championships*


---

**Description**

Point system for the 2008 World Fly Fishing Championships: current and some proposals.

**Usage**

```
wffc.P1(length, c1 = 100, min.eligible = 0.18, ppm = 2000)
wffc.P2(length, c1 = 100, min.eligible = 0.18, ppm = 2000,
        c.quad = 12700)
wffc.P3(length, c1 = 100, min.eligible = 0.18, ppm = 2000)
wffc.P1star(length, c1 = 100, min.eligible = 0.18, ppm = 2000)
wffc.P2star(length, c1 = 100, min.eligible = 0.18, ppm = 2000,
            c.quad = 12700)
wffc.P3star(length, c1 = 100, min.eligible = 0.18, ppm = 2000)
```

**Arguments**

length	Length of the fish, in meters. Numeric vector.
c1	Points added to each eligible fish.
min.eligible	The 2008 WFFC regulations stipulated that the smallest eligible fish was 0.180 m, which is 180 mm.
ppm	Points per meter of length of the fish.
c.quad	Constants for the quadratic terms. The defaults mean that a fish twice the minimum legal size will award about 50 percent more points compared to <code>wffc.P1()</code> and <code>wffc.P1star()</code> . See below for examples.

## Details

The official website contains a document with the official rules and regulations of the competition. The function `wffc.P1()` implements the current WFFC point system, and is ‘discrete’ in that fish lengths are rounded up to the nearest centimeter (provided it is greater or equal to `min.eligible` m). `wffc.P1star()` is a continuous version of it in that it is piecewise linear with two pieces and it is discontinuous at `min.eligible`.

The function `wffc.P2()` is a new proposal which rewards catching bigger fish. It is based on a quadratic polynomial. `wffc.P2star()` is a continuous version of it.

The function `wffc.P3()` is another new proposal which rewards catching bigger fish. Named a *cumulative linear proposal*, its slope is ppm between `min.eligible` and  $2 * \text{min.eligible}$ , its slope is  $2 * \text{ppm}$  between  $2 * \text{min.eligible}$  and  $3 * \text{min.eligible}$ , its slope is  $3 * \text{ppm}$  between  $3 * \text{min.eligible}$  and  $4 * \text{min.eligible}$ , etc. One adds the usual `c1` to each eligible fish. `wffc.P3star()` is a continuous version of `wffc.P3()`.

The function `wffc.P4()` is another new proposal which rewards catching bigger fish. Named a *cumulative linear proposal*, its slope is ppm between `min.eligible` and  $2 * \text{min.eligible}$ , its slope is  $2 * \text{ppm}$  between  $2 * \text{min.eligible}$  and  $1.5 * \text{min.eligible}$ , its slope is  $3 * \text{ppm}$  between  $1.5 * \text{min.eligible}$  and  $2 * \text{min.eligible}$ , etc. One adds the usual `c1` to each eligible fish. `wffc.P4star()` is a continuous version of `wffc.P4()`.

## Value

A vector with the number of points.

## Note

`wffc.P2` and `wffc.P2star` may change in the future, as well as possibly `wffc.P3` and `wffc.P3star` and `wffc.P4` and `wffc.P4star`.

## Author(s)

T. W. Yee.

## References

Yee, T. W. (2014). Scoring rules, and the role of chance: analysis of the 2008 World Fly Fishing Championships. *Journal of Quantitative Analysis in Sports*. **10**, 397–409.

## See Also

[wffc](#).

## Examples

```
## Not run: fishlength <- seq(0.0, 0.72, by = 0.001)
plot(fishlength, wffc.P2star(fishlength), type = "l", col = "blue",
     las = 1, lty = "dashed", lwd = 2, las = 1, cex.main = 0.8,
     xlab = "Fish length (m)", ylab = "Competition points",
     main = "Current (red) and proposed (blue and green) WFFC point system")
lines(fishlength, wffc.P1star(fishlength), type = "l", col = "red", lwd = 2)
```

```

lines(fishlength, wffc.P3star(fishlength), type = "l", col = "darkgreen",
      lwd = 2, lty = "dashed")
lines(fishlength, wffc.P4star(fishlength), type = "l", col = "orange",
      lwd = 2, lty = "dashed")
abline(v = (1:4) * 0.18, lty = "dotted")
abline(h = (1:13) * wffc.P1star(0.18), lty = "dotted")
## End(Not run)

# Successive slopes:
(wffc.P1star((2:8)*0.18) - wffc.P1star((1:7)*0.18)) / (0.18 * 2000)
(wffc.P3star((2:8)*0.18) - wffc.P3star((1:7)*0.18)) / (0.18 * 2000)
(wffc.P4star((2:8)*0.18) - wffc.P4star((1:7)*0.18)) / (0.18 * 2000)

# More successive slopes:
MM2 <- 0.18 / 2
ind1 <- 2:12
(wffc.P4star((ind1)*MM2) - wffc.P4star((ind1-1)*MM2)) / (MM2 * 2000)

# About 50 percent more points:
wffc.P2 (2 * 0.18) / wffc.P1 (2 * 0.18)
wffc.P2star(2 * 0.18) / wffc.P1star(2 * 0.18)

```

---

wffc.teams

2008 World Fly Fishing Championships (Team results) Data

---

## Description

Team results of the 2008 FIPS-MOUCHE World Fly Fishing Championships held in Rotorua, New Zealand during 22–30 March 2008.

## Usage

```
data(wffc.teams)
```

## Format

A data frame with 18 observations on the following 5 variables. Some of these variable are described in [wffc](#).

country a character vector.

totalPlacings a numeric vector; these are the summed placings over the 5 sessions and 5 team members.

points a numeric vector; see [wffc](#).

noofcaptures a numeric vector.

longestfish a numeric vector.

**Details**

This data frame gives the team results of the competition. See also [wffc](#) and [wffc.indiv](#) for more details and links.

**Examples**

```
wffc.teams
```

---

```
xs.nz
```

*Cross-sectional Data from the New Zealand Population*

---

**Description**

A cross-sectional data set of a workforce company, plus another health survey, in New Zealand during the 1990s,

**Usage**

```
data(xs.nz)
```

**Format**

A data frame with 10529 observations on the following 59 variables. For binary variables, a "1" or TRUE means yes, and "0" or FALSE means no. Also, "D" means don't know, and "-" means not applicable. The pregnancy questions were administered to women only.

`regnum` a numeric vector, a unique registration number. This differs from their original registration number, and the rows are sorted by their new registration number.

`study1` a logical vector, Study 1 (workforce) or Study 2?

`age` a numeric vector, age in years.

`sex` a factor with levels F and M.

`pulse` a numeric vector, beats per minute.

`sbp` a numeric vector, systolic blood pressure (mm Hg).

`dbp` a numeric vector, diastolic blood pressure (mm Hg).

`cholest` a numeric vector, cholesterol (mmol/L).

`height` a numeric vector, in m.

`weight` a numeric vector, in kg.

`fh.heartdisease` a factor with levels 0, 1, D. Has a family history of heart disease (heart attack, angina, or had a heart bypass operation) within the immediate family (brother, sister, father or mother, blood relatives only)? Note that D means: do not know.

`fh.age` a factor, following from `fh.heartdisease`, if yes, how old was the family member when it happened (if more than one family member, give the age of the youngest person)?

`fh.cancer` a factor with levels 0, 1, D. Has a family history of cancer within the immediate family (blood relatives only)? Note that D means: do not know.

heartattack a numeric vector, have you ever been told by a doctor that you have had a heart attack ("coronary")?

stroke a numeric vector, have you ever been told by a doctor that you have had a stroke?

diabetes a numeric vector, have you ever been told by a doctor that you have had diabetes?

hypertension a numeric vector, have you ever been told by a doctor that you have had high blood pressure (hypertension)?

highchol a numeric vector, have you ever been told by a doctor that you have had high cholesterol?

asthma a numeric vector, have you ever been told by a doctor that you have had asthma?

cancer a numeric vector, have you ever been told by a doctor that you have had cancer?

acne a numeric vector, have you ever received treatment from a doctor for acne (pimples)?

sunburn a numeric vector, have you ever received treatment from a doctor for sunburn?

smokepassive a numeric vector, on average, how many hours each week (at work and at home) would you spend near someone who is smoking? (put "0" if none)

smokeever a numeric vector, have you ever smoked tailor-made or roll-you-own cigarettes once a week or more? A 1 means yes and 0 means no.

smokenow a numeric vector, do you smoke tailor-made or roll-you-own cigarettes now? A 1 means yes and 0 means no.

smokeagequit a factor, if no to smokenow, how old were you when you stopped smoking? Using `as.numeric(as.character(smokeagequit))` will work for those values which are not `as.character(smokeagequit) == "-"`.

smokeyears a numeric vector, if yes to smokeever, for how many years altogether have you smoked tailor-made or roll-you-own cigarettes?

smoketailormade a numeric vector, how many tailor-made cigarettes do you smoke each day?

smokeweekpack a numeric vector, how many packets of *roll-your-own* tobacco do you use each week? (put "0" if none)

smokepacketsize a numeric vector, what size packets of *roll-your-own* tobacco do you usually buy? ("0" means don't smoke *roll-your-owns*, else 25g or 30g or 35g or 50g)

drinkmonth a numeric vector, do you drink alcohol once a month or more?

drinkfreqweek a numeric vector, if yes to drinkmonth, about how often do you drink alcohol (days per week)? Note: 0.25 is once a month, 0.5 is once every two weeks, 1 is once a week, 2.5 is 2-3 days a week, 4.5 is 4-5 days a week, 6.5 is 6-7 days a week.  
Further note: 1 can, small bottle or handle of beer or home brew = 1 drink, 1 quart bottle of beer = 2 drinks, 1 jug of beer = 3 drinks, 1 flagon/peter of beer = 6 drinks, 1 glass of wine, sherry = 1 drink, 1 bottle of wine = 6 drinks, 1 double nip of spirits = 1 drink.

drinkweek a numeric vector, how many drinks per week, on average. This is the average daily amount of drinks multiplied by the frequency of drinking per week. See `drinkfreqweek` on what constitutes a 'drink'.

drinkmaxday a numeric vector, in the last three months, what is the largest number of drinks that you had on any one day? Warning: some values are considered unrealistically excessive.

eggs a numeric vector, how many eggs do you eat a week (raw, boiled, scrambled, poached, or in quiche)?

chocbiscuits a numeric vector, how many chocolate biscuits do you usually eat in a week?

- pregnant a factor, have you ever been pregnant for more than 5 months?
- pregfirst a factor, if yes to pregnant, how old were you when your first baby was born (or you had a miscarriage after 5 months)?
- preglast a factor, how old were you when your last baby was born (or you had a miscarriage after 5 months)?
- babies numeric, how many babies have you given birth to?
- moody a numeric vector, does your mood often go up or down?
- miserable a numeric vector, do you ever feel 'just miserable' for no reason?
- hurt a numeric vector, are your feelings easily hurt?
- fedup a numeric vector, do you often feel 'fed up'?
- nervous a numeric vector, would you call yourself a nervous person?
- worrier a numeric vector, are you a worrier?
- worry a numeric vector, do you worry about awful things that might happen?
- tense a numeric vector, would you call yourself tense or 'highly strung'?
- embarrassed a numeric vector, do you worry too long after an embarrassing experience?
- nerves a numeric vector, do you suffer from 'nerves'?
- nofriend a numeric vector, do you have a friend or family member that you can talk to about problems or worries that you may have? The value 1 effectively means "no", i.e., s/he has no friend or friends.
- depressed a numeric vector, in your lifetime, have you ever had two weeks or more when nearly every day you felt sad or depressed?
- exervig a numeric vector, how many hours per week would you do any vigorous activity or exercise either at work or away from work that makes you breathe hard and sweat? Values here ought to be less than 168.
- exermod a numeric vector, how many hours per week would you do any moderate activity or exercise such as brisk walking, cycling or mowing the lawn? Values here ought to be less than 168.
- feethour a numeric vector, on an average work day, how long would you spend on your feet, either standing or moving about?
- ethnicity a factor with 4 levels, what ethnic group do you belong to? European = European (NZ European or British or other European), Maori = Maori, Polynesian = Pacific Island Polynesian, Other = Other (Chinese, Indian, Other).
- sleep a numeric vector, how many hours do you usually sleep each night?
- snore a factor with levels 0, 1, D. Do you usually snore? Note that D means: do not know.
- cat a numeric vector, do you have a household pet cat?
- dog a numeric vector, do you have a household pet dog?
- hand a factor with levels right = right, left = left, both = either. Are you right-handed, left-handed, or no preference for left or right?
- numhouse an ordered factor with 4 levels: 1 = 1, 2 = 2, 3 = 3, 4+ = four or more; how many people (including yourself) usually live in your house?

marital a factor with 4 levels: single = single, married = married or living with a partner, separated = separated or divorced, widowed = widowed.

educ an ordered factor with 4 levels: primary = Primary school, secondary = High school/secondary school, polytechnic = Polytechnic or similar, university = University. What was the highest level of education you received?

## Details

The data frame is a subset of the entire data set which was collected from a confidential self-administered questionnaire administered in a large New Zealand workforce observational study conducted during 1992–3. The data were augmented by a second study consisting of retirees. The data can be considered a reasonable representation of the white male New Zealand population in the early 1990s. There were physical, lifestyle and psychological variables that were measured. The psychological variables were headed "Questions about your feelings".

Although some data cleaning was performed and logic checks conducted, anomalies remain. Some variables, of course, are subject to a lot of measurement error and bias. It is conceivable that some participants had poor reading skills! In particular, the smoking variables contain a small percentage of conflicting values, and when NAs are taken into account then there would be several different ways the data might be cleaned. If `smokeever == 0` then strictly speaking, only `smokepassive` is the other variable—the other smoking variables should either be NA or 0.

## Warning

More variables may be added in the future and these may be placed in any column position. Therefore references such as `xs.nz[, 12]` are dangerous. Also, variable names may change in the future as well as their format or internal structure, e.g., factor versus numeric.

## Note

More error checking are needed for the pregnancy and smoking variables.

## Source

Originally, Clinical Trials Research Unit, University of Auckland, New Zealand, <http://www.ctr.u.auckland.ac.nz>. Originally much of the error checking and formatting was performed by Stephen Vander Hoorn. Lately (2014), more changes and error checks were made to the data by James T. Gray.

## References

MacMahon, S., Norton, R., Jackson, R., Mackie, M. J., Cheng, A., Vander Hoorn, S., Milne, A., McCulloch, A. (1995). Fletcher Challenge-University of Auckland Heart & Health Study: design and baseline findings. *New Zealand Medical Journal*, **108**, 499–502.

## See Also

[chest.nz](#).

**Examples**

```
data(xs.nz)
summary(xs.nz)
```

---

yip88

*Zero-Inflated Poisson Distribution (Yip (1988) algorithm)*


---

**Description**

Fits a zero-inflated Poisson distribution based on Yip (1988).

**Usage**

```
yip88(link = "loglink", n.arg = NULL, imethod = 1)
```

**Arguments**

link	Link function for the usual $\lambda$ parameter. See <a href="#">Links</a> for more choices.
n.arg	The total number of observations in the data set. Needed when the response variable has all the zeros deleted from it, so that the number of zeros can be determined.
imethod	Details at <a href="#">CommonVGAMffArguments</a> .

**Details**

The method implemented here, Yip (1988), maximizes a *conditional* likelihood. Consequently, the methodology used here deletes the zeros from the data set, and is thus related to the positive Poisson distribution (where  $P(Y = 0) = 0$ ).

The probability function of  $Y$  is 0 with probability  $\phi$ , and  $\text{Poisson}(\lambda)$  with probability  $1 - \phi$ . Thus

$$P(Y = 0) = \phi + (1 - \phi)P(W = 0)$$

where  $W$  is  $\text{Poisson}(\lambda)$ . The mean,  $(1 - \phi)\lambda$ , can be obtained by the extractor function `fitted` applied to the object.

This family function treats  $\phi$  as a scalar. If you want to model both  $\phi$  and  $\lambda$  as a function of covariates, try [zipoisson](#).

**Value**

An object of class "vglmff" (see [vglmff-class](#)). The object is used by modelling functions such as [vglm](#), [rrvglm](#) and [vgam](#).

**Warning**

Under- or over-flow may occur if the data is ill-conditioned. Yip (1988) only considered  $\phi$  being a scalar and not modelled as a function of covariates. To get around this limitation, try [zipoisson](#).

Inference obtained from `summary.vglm` and `summary.vgam` may or may not be correct. In particular, the p-values, standard errors and degrees of freedom may need adjustment. Use simulation on artificial data to check that these are reasonable.

**Note**

The data may be inputted in two ways. The first is when the response is a vector of positive values, with the argument `n` in `yip88` specifying the total number of observations. The second is simply include all the data in the response. In this case, the zeros are trimmed off during the computation, and the `x` and `y` slots of the object, if assigned, will reflect this.

The estimate of  $\phi$  is placed in the `misc` slot as `@misc$pstr0`. However, this estimate is computed only for intercept models, i.e., the formula is of the form  $y \sim 1$ .

**Author(s)**

Thomas W. Yee

**References**

Yip, P. (1988). Inference about the mean of a Poisson distribution in the presence of a nuisance parameter. *The Australian Journal of Statistics*, **30**, 299–306.

Angers, J-F. and Biswas, A. (2003). A Bayesian analysis of zero-inflated generalized Poisson model. *Computational Statistics & Data Analysis*, **42**, 37–46.

**See Also**

[zipoisson](#), [Zipois](#), [zapoisson](#), [pospoisson](#), [poissonff](#), [dzipois](#).

**Examples**

```
phi <- 0.35; lambda <- 2 # Generate some artificial data
y <- rzipois(n <- 1000, lambda, phi)
table(y)

# Two equivalent ways of fitting the same model
fit1 <- vglm(y ~ 1, yip88(n = length(y)), subset = y > 0)
fit2 <- vglm(y ~ 1, yip88, trace = TRUE, crit = "coef")
(true.mean <- (1-phi) * lambda)
mean(y)
head(fitted(fit1))
fit1@misc$pstr0 # The estimate of phi

# Compare the ZIP with the positive Poisson distribution
pp <- vglm(y ~ 1, pospoisson, subset = y > 0, crit = "c")
coef(pp)
Coef(pp)
coef(fit1) - coef(pp) # Same
head(fitted(fit1) - fitted(pp)) # Different

# Another example (Angers and Biswas, 2003) -----
abdata <- data.frame(y = 0:7, w = c(182, 41, 12, 2, 2, 0, 0, 1))
abdata <- subset(abdata, w > 0)

yy <- with(abdata, rep(y, w))
fit3 <- vglm(yy ~ 1, yip88(n = length(yy)), subset = yy > 0)
```

```
fit3@misc$pstr0 # Estimate of phi (they get 0.5154 with SE 0.0707)
coef(fit3, matrix = TRUE)
Coef(fit3) # Estimate of lambda (they get 0.6997 with SE 0.1520)
head(fitted(fit3))
mean(yy) # Compare this with fitted(fit3)
```

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