Package ‘RxODE’

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Title Facilities for Simulating from ODE-Based Models
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Description Facilities for running simulations from ordinary differential equation (‘ODE’) models, such as pharmacometrics and other compartmental models. A compilation manager translates the ODE model into C, compiles it, and dynamically loads the object code into R for improved computational efficiency. An event table object facilitates the specification of complex dosing regimens (optional) and sampling schedules. NB: The use of this package requires both C and Fortran compilers, for details on their use with R please see Section 6.3, Appendix A, and Appendix D in the ‘R Administration and Installation’ manual. Also the code is mostly released under GPL. The ‘VODE’ and ‘LSODA’ are in the public domain. The information is available in the inst/COPYRIGHTS.

BugReports https://github.com/nlmixrdevelopment/RxODE/issues/
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.rxFinalizeInner

Description

Finalize inner RxODE based on symengine saved info

Usage

.rxFinalizeInner(.s, sum.prod = FALSE, optExpression = TRUE)

Arguments

.s Symengine/RxODE object
sum.prod A boolean determining if RxODE should use more numerically stable sums/products.
optExpression Optimize the model text for computer evaluation.

Value

Nothing

Author(s)

Matthew L Fidler
.rxFinalizePred

Finalize RxODE pred based on symengine saved info

Description

Finalize RxODE pred based on symengine saved info

Usage

.rxFinalizePred(.s, sum.prod = FALSE, optExpression = TRUE)

Arguments

.s Symengine/RxODE object
sum.prod A boolean determining if RxODE should use more numerically stable sums/products.
optExpression Optimize the model text for computer evaluation.

Value

Nothing

Author(s)

Matthew L Fidler

.rxGenEBE

Generate pieces for EBE only problem

Description

Generate pieces for EBE only problem

Usage

.rxGenEBE(
  obj,
  predfn,
  pkpars = NULL,
  errfn = NULL,
  init = NULL,
  pred.minus.dv = TRUE,
  sum.prod = FALSE,
  optExpression = TRUE,
  theta = FALSE,
  addProp = c("combined2", "combined1")
)

**Arguments**

- **obj**: RxODE object
- **predfn**: Prediction function
- **pkpars**: Pk Pars function
- **errfn**: Error function
- **init**: Initialization parameters for scaling.
- **pred.minus.dv**: Boolean stating if the FOCEi objective function is based on PRED-DV (like NONMEM). Default TRUE.
- **sum.prod**: A boolean determining if RxODE should use more numerically stable sums/products.
- **optExpression**: Optimize the model text for computer evaluation.
- **theta**: Calculate THETA derivatives instead of ETA derivatives. By default FALSE
- **addProp**: one of "combined1" and "combined2"; These are the two forms of additive+proportional errors supported by monolix/nonmem:
  - combined1: \( \text{transform}(y) = \text{transform}(f) + (a + b \cdot f^c) \cdot \epsilon \)
  - combined2: \( \text{transform}(y) = \text{transform}(f) + (a^2 + b^2 \cdot f^{2c}) \cdot \epsilon \)

**Value**

RxODE/symengine environment

**Author(s)**

Matthew L. Fidler

---

**.rxGenEtaS**

*Generate the ETA sensitivities for FO related methods*

**Description**

Generate the ETA sensitivities for FO related methods

**Usage**

```r
.rxGenEtaS(
  obj,
  predfn,
  pkpars = NULL,
  errfn = NULL,
  init = NULL,
  promoteLinSens = TRUE,
  theta = FALSE,
  addProp = c("combined2", "combined1")
)
```
Arguments

- **obj**: RxODE object
- **predfn**: Prediction function
- **pkpars**: Pk Pars function
- **errfn**: Error function
- **init**: Initialization parameters for scaling.
- **promoteLinSens**: Promote solved linear compartment systems to sensitivity-based solutions.
- **theta**: Calculate THETA derivatives instead of ETA derivatives. By default FALSE
- **addProp**: one of "combined1" and "combined2"; These are the two forms of additive+proportional errors supported by monolix/nonmem:
  - combined1: transform(y) = transform(f) + (a + b*f^c)*eps
  - combined2: transform(y) = transform(f) + (a^2 + b^2*f^(2c))*eps

Value

RxODE/symengine environment

Author(s)

Matthew L. Fidler

Description

Generate FOCE without interaction

Usage

```r
.rxGenFoce(
  obj,
  predfn,
  pkpars = NULL,
  errfn = NULL,
  init = NULL,
  pred.minus.dv = TRUE,
  sum.prod = FALSE,
  optExpression = TRUE,
  promoteLinSens = TRUE,
  theta = FALSE,
  addProp = c("combined2", "combined1")
)
```
### Arguments

- **obj**: RxODE object
- **predfn**: Prediction function
- **pkpars**: Pk Pars function
- **errfn**: Error function
- **init**: Initialization parameters for scaling.
- **pred.minus.dv**: Boolean stating if the FOCEi objective function is based on PRED-DV (like NONMEM). Default TRUE.
- **sum.prod**: A boolean determining if RxODE should use more numerically stable sums/products.
- **optExpression**: Optimize the model text for computer evaluation.
- **promoteLinSens**: Promote solved linear compartment systems to sensitivity-based solutions.
- **theta**: Calculate THETA derivatives instead of ETA derivatives. By default FALSE
- **addProp**: one of "combined1" and "combined2"; These are the two forms of additive+proportional errors supported by monolix/nonmem:
  - combined1: transform(y) = transform(f) + (a + b * f^c) * eps
  - combined2: transform(y) = transform(f) + (a^2 + b^2 * f^(2c)) * eps

### Value

RxODE/symengine environment

### Author(s)

Matthew Fidler

---

`.rxGenFocei` Generate pieces for FOCEi inner problem

### Description

Generate pieces for FOCEi inner problem

### Usage

```r
.rxGenFocei(
  obj,
  predfn,
  pkpars = NULL,
  errfn = NULL,
  init = NULL,
  pred.minus.dv = TRUE,
  sum.prod = FALSE,
  optExpression = TRUE,
```
promoteLinSens = TRUE,
theta = FALSE,
addProp = c("combined2", "combined1")
)

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>obj</td>
<td>RxODE object</td>
</tr>
<tr>
<td>predfn</td>
<td>Prediction function</td>
</tr>
<tr>
<td>pkpars</td>
<td>Pk Pars function</td>
</tr>
<tr>
<td>errfn</td>
<td>Error function</td>
</tr>
<tr>
<td>init</td>
<td>Initialization parameters for scaling.</td>
</tr>
<tr>
<td>pred.minus.dv</td>
<td>Boolean stating if the FOCEi objective function is based on PRED-DV (like NONMEM). Default TRUE.</td>
</tr>
<tr>
<td>sum.prod</td>
<td>A boolean determining if RxODE should use more numerically stable sums/products.</td>
</tr>
<tr>
<td>optExpression</td>
<td>Optimize the model text for computer evaluation.</td>
</tr>
<tr>
<td>promoteLinSens</td>
<td>Promote solved linear compartment systems to sensitivity-based solutions.</td>
</tr>
<tr>
<td>theta</td>
<td>Calculate THETA derivatives instead of ETA derivatives. By default FALSE</td>
</tr>
<tr>
<td>addProp</td>
<td>one of &quot;combined1&quot; and &quot;combined2&quot;; These are the two forms of additive+proportional errors supported by monolix/nonmem: combined1: transform(y)=transform(f)+(a+b*f^c)<em>eps combined2: transform(y)=transform(f)+(a^2+b^2</em>f^(2c))*eps</td>
</tr>
</tbody>
</table>

Value

RxODE/symengine environment

Author(s)

Matthew L. Fidler

Description

Get the state information for the current model

Usage

.rxGenFunState(obj)

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>obj</td>
<td>RxODE model that can take rxModelVars</td>
</tr>
</tbody>
</table>
Value

ccharacter vector of initial preserved states (state), extra states (statef) and dvid translation information (dvid). This is used in generating the final RxODE model.

Author(s)

Matthew Fidler

Generate the $d(\text{err})/d(\text{eta})$ values for FO related methods

Usage

\[.\text{rxGenHdEta}(\text{obj}, \text{predfn}, \text{pkpars} = \text{NULL}, \text{errfn} = \text{NULL}, \text{init} = \text{NULL}, \text{pred.minus.dv} = \text{TRUE}, \text{promoteLinSens} = \text{TRUE}, \text{theta} = \text{FALSE}, \text{addProp} = \text{c("combined2", "combined1")})\]

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>obj</td>
<td>RxODE object</td>
</tr>
<tr>
<td>predfn</td>
<td>Prediction function</td>
</tr>
<tr>
<td>pkpars</td>
<td>Pk Pars function</td>
</tr>
<tr>
<td>errfn</td>
<td>Error function</td>
</tr>
<tr>
<td>init</td>
<td>Initialization parameters for scaling.</td>
</tr>
<tr>
<td>pred.minus.dv</td>
<td>Boolean stating if the FOCEi objective function is based on PRED-DV (like NONMEM). Default TRUE.</td>
</tr>
<tr>
<td>promoteLinSens</td>
<td>Promote solved linear compartment systems to sensitivity-based solutions.</td>
</tr>
<tr>
<td>theta</td>
<td>Calculate THETA derivatives instead of ETA derivatives. By default FALSE</td>
</tr>
<tr>
<td>addProp</td>
<td>one of &quot;combined1&quot; and &quot;combined2&quot;; These are the two forms of additive+proportional errors supported by monolix/nonmem: combined1: transform(y)=transform(f)+(a+b*f^c)<em>eps combined2: transform(y)=transform(f)+(a^2+b^2</em>f^2c)*eps</td>
</tr>
</tbody>
</table>
Value
RxODE/symengine environment

Author(s)
Matthew L. Fidler

Usage
.rxGenPred(obj, predfn, errfn, init, addProp = c("combined2", "combined1"))

Arguments

.obj RxODE object
.pedfn Prediction function
.errfn Error function
.init Initialization parameters for scaling.
.addProp one of "combined1" and "combined2"; These are the two forms of additive+proportional errors supported by monolix/nonmem:
combined1: \( \text{transform}(y) = \text{transform}(f) + (a + b \cdot f^c) \cdot \varepsilon \)
combined2: \( \text{transform}(y) = \text{transform}(f) + (a^2 + b^2 \cdot f^{2c}) \cdot \varepsilon \)

Value
A list of (1) RxODE model variables augmented with pred/error information and (2) extra error variables created.

Author(s)
Matthew L Fidler
.rxRmPrint

Description
Remove print statements

Usage
.rxRmPrint(x)

Arguments
x RxODE lines to remove

Value
RxODE with print lines removed.

Author(s)
Matthew L. Fidler

.rxWinRtoolsPath Setup Rtools path

Description
Setup Rtools path

Usage
.rxWinRtoolsPath(rm.rtools = TRUE, retry = FALSE)

Arguments
rm.rtools Remove the Rtools from the current path specs.
retry Should you retry to find Rtools? If NA, don’t throw an error if it isn’t found.

Author(s)
Matthew L. Fidler
.rxWithOptions

**Description**

Temporarily set options then restore them while running code

**Usage**

```
.rxWithOptions(ops, code)
```

**Arguments**

- `ops` list of options that will be temporarily set for the code
- `code` The code to run during the sink

**Value**

value of code

**Examples**

```
.rxWithOptions(list(digits = 21), {
    print(pi)
})
```

print(pi)

---

.dxWithWd

**Description**

Temporarily set options then restore them while running code

**Usage**

```
.rxWithWd(wd, code)
```

**Arguments**

- `wd` working directory to temporarily set the system to while evaluating the code
- `code` The code to run during the sink
.setWarnIdSort

Value

value of code

Examples

```
.rxWithWd(tempdir(), {
  getwd()
})
getwd()
```

Description

Turn on/off warnings for ID sorting.

Usage

```
.setWarnIdSort(warnIdSort = TRUE)
```

Arguments

- `warnIdSort` Boolean for if the sorting warning is turned on or off.

Value

Nothing

Author(s)

Matthew Fidler
**add.dosing**

*Add dosing to eventTable*

**Description**

This adds a dosing event to the event table. This is provided for piping syntax through magrittr. It can also be accessed by `eventTable$add.dosing(...)`

**Usage**

```r
add.dosing(
  eventTable,
  dose,
  nbr.doses = 1L,
  dosing.interval = 24,
  dosing.to = 1L,
  rate = NULL,
  amount.units = NA_character_,
  start.time = 0,
  do.sampling = FALSE,
  time.units = NA_character_,
  ...
)
```

**Arguments**

- `eventTable`  
  eventTable object; When accessed from object it would be eventTable$eventTable
- `dose`  
  numeric scalar, dose amount in `amount.units`
- `nbr.doses`  
  integer, number of doses;
- `dosing.interval`  
  required numeric scalar, time between doses in `time.units`, defaults to 24 of `time.units="hours"`;  
- `dosing.to`  
  integer, compartment the dose goes into (first compartment by default);
- `rate`  
  for infusions, the rate of infusion (default is NULL, for bolus dosing);
- `amount.units`  
  optional string indicating the dosing units. Defaults to `NA` to indicate as per the original EventTable definition.
- `start.time`  
  required dosing start time;
- `do.sampling`  
  logical, should observation sampling records be added at the dosing times? Defaults to `FALSE`.
- `time.units`  
  optional string indicating the time units. Defaults to "hours" to indicate as per the original EventTable definition.
- `...`  
  Other parameters passed to `et()`.
Value

eventTable with updated dosing (note the event table will be updated anyway)

Author(s)

Matthew L. Fidler
Matthew L Fidler, Wenping Wang

References


See Also

eventTable, add.sampling, add.dosing, et, etRep, etRbind, RxODE

Examples

library(RxODE)
library(units)

## Model from RxODE tutorial
mod1 <- RxODE(
  KA=2.94E-01;
  CL=1.86E+01;
  V2=4.02E+01;
  Q=1.05E+01;
  V3=2.97E+02;
  Kin=1;
  Kout=1;
  EC50=200;
  C2 = centr/V2;
  C3 = peri/V3;
  d/dt(depot) =-KA*depot;
  d/dt(centr) = KA*depot - CL*C2 - Q*C2 + Q*C3;
  d/dt(peri) = Q*C2 - Q*C3;
  d/dt(eff) = Kin - Kout*(1-C2/(EC50+C2))*eff;
)

## These are making the more complex regimens of the RxODE tutorial

## bid for 5 days
bid <- et(timeUnits="hr") %>%
et(amt=10000,ii=12,until=set_units(5, "days"))

## qd for 5 days
qd <- et(timeUnits="hr") %>%
et(amt=20000, ii=24, until=set_units(5, "days"))

## bid for 5 days followed by qd for 5 days
et <- seq(bid,qd) %>% et(seq(0,11*24,length.out=100));
bidQd <- rxSolve(mod1, et)
plot(bidQd, C2)

## Now Infusion for 5 days followed by oral for 5 days
## note you can dose to a named compartment instead of using the compartment number
infusion <- et(timeUnits = "hr") %>%
et(amt=10000, rate=5000, ii=24, until=set_units(5, "days"), cmt="centr")
qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(5, "days"), cmt="depot")
et <- seq(infusion,qd)
infusionQd <- rxSolve(mod1, et)
plot(infusionQd, C2)

## 2wk-on, 1wk-off
qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")
et <- seq(qd, set_units(1,"weeks"), qd) %>%
    add.sampling(set_units(seq(0, 5.5,by=0.005),weeks))
wkOnOff <- rxSolve(mod1, et)
plot(wkOnOff, C2)

## You can also repeat the cycle easily with the rep function
qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")
et <- etRep(qd, times=4, wait=set_units(1,"weeks")) %>%
    add.sampling(set_units(seq(0, 12.5,by=0.005),weeks))
repCycle4 <- rxSolve(mod1, et)
plot(repCycle4, C2)
add.sampling

Add sampling to eventTable

Description
This adds a dosing event to the event table. This is provided for piping syntax through magrittr. It can also be accessed by eventTable$add.sampling()

Usage
add.sampling(eventTable, time, time.units = NA)

Arguments
- eventTable: An eventTable object. When accessed from object it would be eventTable$
- time: a vector of time values (in time.units).
- time.units: an optional string specifying the time units. Defaults to the units specified when the EventTable was initialized.

Value
eventTable with updated sampling. (Note the event table will be updated even if you don’t reassign the eventTable)

Author(s)
Matthew L Fidler, Wenping Wang

References

See Also
eventTable, add.sampling, add.dosing, et, etRep, etRbind, RxODE

Examples

library(RxODE)
library(units)

## Model from RxODE tutorial
mod1 <- RxODE({
KA=2.94E-01;
CL=1.86E+01;
V2=4.02E+01;
Q=1.05E+01;
V3=2.97E+02;
Kin=1;
Kout=1;
EC50=200;
C2 = centr/V2;
C3 = peri/V3;
d/dt(depot) =-KA*depot;
d/dt(centr) = KA*depot - CL*C2 - Q*C2 + Q*C3;
d/dt(peri) = Q*C2 - Q*C3;
d/dt(eff) = Kin - Kout*(1-C2/(EC50+C2))*eff;

## These are making the more complex regimens of the RxODE tutorial

## bid for 5 days
bid <- et(timeUnits="hr") %>%
et(amt=10000,ii=12,until=set_units(5, "days"))

## qd for 5 days
qd <- et(timeUnits="hr") %>%
et(amt=20000,ii=24,until=set_units(5, "days"))

## bid for 5 days followed by qd for 5 days
et <- seq(bid,qd) %>% et(seq(0,11*24,length.out=100));
bidQd <- rxSolve(mod1, et)
plot(bidQd, C2)

## Now Infusion for 5 days followed by oral for 5 days

## note you can dose to a named compartment instead of using the compartment number
infusion <- et(timeUnits = "hr") %>%
et(amt=10000, rate=5000, ii=24, until=set_units(5, "days"), cmt="centr")

qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(5, "days"), cmt="depot")
et <- seq(infusion,qd)
infusionQd <- rxSolve(mod1, et)
plot(infusionQd, C2)

## 2wk-on, 1wk-off
qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")
et <- seq(qd, set_units(1, "weeks"), qd) %>%
    add.sampling(set_units(seq(0, 5.5, by=0.005), weeks))

wkOnOff <- rxSolve(mod1, et)

plot(wkOnOff, C2)

## You can also repeat the cycle easily with the rep function

qd <- et(timeUnits = "hr") %>%
    et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")

et <- etRep(qd, times=4, wait=set_units(1, "weeks")) %>%
    add.sampling(set_units(seq(0, 12.5, by=0.005), weeks))

repCycle4 <- rxSolve(mod1, et)

plot(repCycle4, C2)

---

**as.data.table.rxEt**

Convert event table to a data.table

**Description**

Convert an event table to a data.table

**Usage**

`as.data.table.rxEt(x, keep.rownames = FALSE, ...)`

**Arguments**

- `x` An R object.
- `keep.rownames` Default is FALSE. If TRUE, adds the input object’s names as a separate column named “rn”. `keep.rownames = "id"` names the column “id” instead.
- `...` Additional arguments to be passed to or from other methods.

**Value**

data.table of event table
as.et

Coerce object to data.frame

Description
Coerce object to data.frame

Usage

as.et(x, ...)

## Default S3 method:
as.et(x, ...)

Arguments

x Object to coerce to et.

... Other parameters

Value
An event table

as_tibble.rxEt

Convert to tbl

Description
Convert to tbl

Usage

## S3 method for class 'rxEt'
as_tibble(x, ...)

Arguments

x RxODE event table

... Other arguments to as_tibble

Value
tibble of event table
coef.RxODE

Return the RxODE coefficients

Description

This returns the parameters, state variables

Usage

## S3 method for class 'RxODE'
coef(object, ...)

Arguments

object is an RxODE object

... ignored arguments

Value

a rxCoef object with the following

- params is a list of strings for parameters for the RxODE object
- state is a list of strings for the names of each state in the RxODE object.
- ini is the model specified default values for the parameters.
- RxODE is the referring RxODE object

Author(s)

Matthew L. Fidler

cvPost

Sample a covariance Matrix from the Posterior Inverse Wishart distribution.

Description

Note this Inverse wishart rescaled to match the original scale of the covariance matrix.
Usage

```
cvPost(
  nu,
  omega,
  n = 1L,
  omegaIsChol = FALSE,
  returnChol = FALSE,
  type = c("invWishart", "lkj", "separation"),
  diagXformType = c("log", "identity", "variance", "nlmixrSqrt", "nlmixrLog",
                   "nlmixrIdentity")
)
```

Arguments

- **nu**: Degrees of Freedom (Number of Observations) for covariance matrix simulation.
- **omega**: Either the estimate of covariance matrix or the estimated standard deviations in matrix form each row forming the standard deviation simulated values
- **n**: Number of Matrices to sample. By default this is 1. This is only useful when omega is a matrix. Otherwise it is determined by the number of rows in the input omega matrix of standard deviations
- **omegaIsChol**: is an indicator of if the omega matrix is in the Cholesky decomposition. This is only used when codetype="invWishart"
- **returnChol**: Return the Cholesky decomposition of the covariance matrix sample. This is only used when codetype="invWishart"
- **type**: The type of covariance posterior that is being simulated. This can be:
  - *invWishart* The posterior is an inverse wishart; This allows for correlations between parameters to be modeled. All the uncertainty in the parameter is captured in the degrees of freedom parameter.
  - *lkj* The posterior separates the standard deviation estimates (modeled outside and provided in the omega argument) and the correlation estimates. The correlation estimate is simulated with the `rLKJ1()` function. This simulation uses the relationship \( \eta=(\nu-1)/2 \). This is relationship based on the proof of the relationship between the restricted LKJ-distribution and inverse wishart distribution (XXXXXX). Once the correlation posterior is calculated, the estimated standard deviations are then combined with the simulated correlation matrix to create the covariance matrix.
  - *separation* Like the lkj option, this separates out the estimation of the correlation and standard deviation. Instead of using the LKJ distribution to simulate the correlation, it simulates the inverse wishart of the identity matrix and converts the result to a correlation matrix. This correlation matrix is then used with the standard deviation to calculate the simulated covariance matrix.
- **diagXformType**: Diagonal transformation type. These could be:
  - *log* The standard deviations are log transformed, so the actual standard deviations are \( \exp(\omega) \)
• identity The standard deviations are not transformed. The standard deviations are not transformed; they should be positive.
• variance The variances are specified in the omega matrix; they are transformed into standard deviations.
• nlmixrSqrt These standard deviations come from an nlmixr omega matrix where diag(chol(inv(omega)))) = x^2
• nlmixrLog These standard deviations come from an nlmixr omega matrix omega matrix where diag(chol(solve(omega))) = exp(x)
• nlmixrIdentity These standard deviations come from an nlmixr omega matrix omega matrix where diag(chol(solve(omega))) = x

The nlmixr transformations only make sense when there is no off-diagonal correlations modeled.

Details

If your covariance matrix is a 1x1 matrix, this uses an scaled inverse chi-squared which is equivalent to the Inverse Wishart distribution in the uni-directional case.

In general, the separation strategy is preferred for diagonal matrices. If the dimension of the matrix is below 10, lkj is numerically faster than separation method. However, the lkj method has densities too close to zero (XXXX) when the dimension is above 10. In that case, though computationally more expensive separation method performs better.

For matrices with modeled covariances, the easiest method to use is the inverse Wishart which allows the simulation of correlation matrices (XXXX). This method is more well suited for well behaved matrices, that is the variance components are not too low or too high. When modeling non-linear mixed effects modeling matrices with too high or low variances are considered sub-optimal in describing a system. With these rules in mind, it is reasonable to use the inverse Wishart.

Value

a matrix (n=1) or a list of matrices (n > 1)

Author(s)

Matthew L. Fidler & Wenping Wang

References


Examples

```r
## Sample a single covariance.
draw1 <- cvPost(3, matrix(c(1, .3, .3, 1), 2, 2))
```
## Sample 3 covariances
set.seed(42)
draw3 <- cvPost(3, matrix(c(1, .3, .3, 1), 2, 2), n = 3)

## Sample 3 covariances, but return the cholesky decomposition
set.seed(42)
draw3c <- cvPost(3, matrix(c(1, .3, .3, 1), 2, 2), n = 3, returnChol = TRUE)

## Sample 3 covariances with lognormal standard deviations via LKJ
cvPost(3, sapply(1:3, function(...) { rnorm(10) }), type = "lkj")

## or return cholesky decomposition
cvPost(3, sapply(1:3, function(...) { rnorm(10) }),
type = "lkj",
returnChol = TRUE)

## Sample 3 covariances with lognormal standard deviations via separation
cvPost(3, sapply(1:3, function(...) { rnorm(10) }), type = "separation")

## or returning the cholesky decomposition
cvPost(3, sapply(1:3, function(...) { rnorm(10) }),
type = "separation",
returnChol = TRUE)

---

**et**  
*Event Table Function*

### Description

Event Table Function

### Usage

```r
et(x, ..., envir = parent.frame())
```

## S3 method for class 'RxODE'
et(x, ..., envir = parent.frame())
et

## S3 method for class 'rxSolve'
et(x, ..., envir = parent.frame())

## S3 method for class 'rxParams'
et(x, ..., envir = parent.frame())

## Default S3 method:
et(
  x,
  ..., 
  time,
  amt,
  evid,
  cmt,
  ii,
  addl,
  ss,
  rate,
  dur,
  until,
  id,
  amountUnits,
  timeUnits,
  addSampling,
  envir = parent.frame(),
  by = NULL,
  length.out = NULL
)

**Arguments**

- **x**
  This is the first argument supplied to the event table. This is named to allow et to be used in a pipe-line with arbitrary objects.

- **...**
  Times or event tables. They can also be one of the named arguments below.

- **envir**
  the environment in which expr is to be evaluated. May also be NULL, a list, a data frame, a pairlist or an integer as specified to `sys.call`.

- **time**
  Time is the time of the dose or the sampling times. This can also be unspecified and is determined by the object type (list or numeric/integer).

- **amt**
  Amount of the dose. If specified, this assumes a dosing record, instead of a sampling record.

- **evid**
  Event ID; This can be:

**Numeric Value** | **Description**
-----------------|-------------------
0                | An observation. This can also be specified as evid=obs
1                | A dose observation. This can also be specified as evid=dose
2                | A non-dose event. This can also be specified as evid=other
A reset event. This can also be specified as `evid=reset`.
Dose and reset event. This can also be specified as `evid=doseReset` or `evid=resetDose`

Note a reset event resets all the compartment values to zero and turns off all infusions.

### Compartment name or number
- If a number, this is an integer starting at 1. Negative compartments turn off a compartment.
- If the compartment is a name, the compartment name is changed to the correct state/compartment number before running the simulation. For a compartment named "-cmt" the compartment is turned off.

Can also specify `cmt` as `dosing.to`, `dose.to`, `doseTo`, `dosingTo`, and `state`.

### Inter-dose interval
- When specifying a dose, this is the inter-dose interval for `ss`, `addl` and `until` options (described below).

### Number of additional doses
- The number of additional doses at an inter-dose interval after one dose.

### Steady state flag
- It can be one of:

<table>
<thead>
<tr>
<th>Value</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>This dose is not a steady state dose</td>
</tr>
<tr>
<td>1</td>
<td>This dose is a steady state dose with the between/inter-dose interval of <code>ii</code></td>
</tr>
<tr>
<td>2</td>
<td>Superposition steady state</td>
</tr>
</tbody>
</table>

When `ss=2` the steady state dose that uses the super-position principle to allow more complex steady states, like 10 mg in the morning and 20 mg at night, or dosing at 8 am 12 pm and 8 pm instead of every 12 hours. Since it uses the super positioning principle, it only makes sense when you know the kinetics are linear.

All other values of `SS` are currently invalid.

### Rate
- When positive, this is the rate of infusion. Otherwise:

<table>
<thead>
<tr>
<th>Value</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>No infusion is on this record</td>
</tr>
<tr>
<td>-1</td>
<td>Modeled rate (in RxODE:rate(cmt) =); Can be et(rate=model).</td>
</tr>
<tr>
<td>-2</td>
<td>Modeled duration (in RxODE: dur(cmt) =); Can be et(dur=model) or et(rate=dur).</td>
</tr>
</tbody>
</table>

When a modeled bioavailability is applied to positive rates (`rate > 0`), the duration of infusion is changed. This is because the data specify the rate and amount, the only thing that modeled bioavailability can affect is duration.

If instead you want the modeled bioavailability to increase the rate of infusion instead of the duration of infusion, specify the `dur` instead or model the duration with `rate=2`. 
**dur**  
Duration of infusion. When *amt* and *dur* are specified the rate is calculated from the two data items. When *dur* is specified instead of *rate*, the bioavailability changes will increase rate instead of duration.

**until**  
This is the time until the dosing should end. It can be an easier way to figure out how many additional doses are needed over your sampling period.

**id**  
A integer vector of IDs to add or remove from the event table. If the event table is identical for each ID, then you may expand it to include all the IDs in this vector. All the negative IDs in this vector will be removed.

**amountUnits**  
The units for the dosing records (*amt*)

**timeUnits**  
The units for the time records (*time*)

**addSampling**  
This is a boolean indicating if a sampling time should be added at the same time as a dosing time. By default this is `FALSE`.

**by**  
When there are no observations in the event table, this is the amount to increment for the observations between `from` and `to`.

**length.out**  
The number of observations to create if there isn’t any observations in the event table. By default this is 200.

**Value**
A new event table

**Author(s)**
Matthew L Fidler, Wenping Wang

**References**

**See Also**
eventTable, add.sampling, add.dosing, et, etRep, etRbind, RxODE

**Examples**

```r
library(RxODE)
library(units)

## Model from RxODE tutorial
mod1 <- RxODE(
  KA=2.94E-01;
  CL=1.86E+01;
  V2=4.02E+01;
```

Q=1.05E+01;  
V3=2.97E+02;  
Kin=1;  
Kout=1;  
EC50=200;  
C2 = centr/V2;  
C3 = peri/V3;  
\[ \frac{d}{dt}(depot) = -KA \times depot; \]  
\[ \frac{d}{dt}(centr) = KA \times depot - CL \times C2 - Q \times C2 + Q \times C3; \]  
\[ \frac{d}{dt}(peri) = Q \times C2 - Q \times C3; \]  
\[ \frac{d}{dt}(eff) = Kin - Kout \times \left(1 - \frac{C2}{EC50 + C2}\right) \times eff; \]  
});

## These are making the more complex regimens of the RxODE tutorial

## bid for 5 days
bid <- et(timeUnits="hr") %>%  
et(amt=10000,ii=12,until=set_units(5, "days"))

## qd for 5 days
qd <- et(timeUnits="hr") %>%  
et(amt=20000,ii=24,until=set_units(5, "days"))

## bid for 5 days followed by qd for 5 days
et <- seq(bid,qd) %>% et(seq(0,11*24,length.out=100));

bidQd <- rxSolve(mod1, et)
plot(bidQd, C2)

## Now Infusion for 5 days followed by oral for 5 days

## note you can dose to a named compartment instead of using the compartment number
infusion <- et(timeUnits = "hr") %>%  
et(amt=10000, rate=5000, ii=24, until=set_units(5, "days"), cmt="centr")

qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(5, "days"), cmt="depot")

et <- seq(infusion,qd)
infusionQd <- rxSolve(mod1, et)
plot(infusionQd, C2)

## 2wk-on, 1wk-off
qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")
et <- seq(qd, set_units(1,"weeks"), qd) %>%  
add.sampling(set_units(seq(0, 5.5,by=0.005),weeks))
wkOnOff <- rxSolve(mod1, et)
plot(wkOnOff, C2)

## You can also repeat the cycle easily with the rep function
qd <- et(timeUnits = "hr") %>% et(amt = 10000, ii = 24, until = set_units(2, "weeks"), cmt = "depot")
et <- etRep(qd, times = 4, wait = set_units(1, "weeks")) %>%
    add.sampling(set_units(seq(0, 12.5, by = 0.005), weeks))
repCycle4 <- rxSolve(mod1, et)
plot(repCycle4, C2)

---

etExpand

*Expand additional doses*

**Description**
Expand additional doses

**Usage**
etExpand(et)

**Arguments**
et
Event table to expand additional doses for.

**Value**
New event table with adtl doses expanded

**Author(s)**
Matthew Fidler

**Examples**
ev <- et(amt = 3, ii = 24, until = 240)
print(ev)
etExpand(ev) # expands event table, but doesn't modify it
print(ev)
ev$expand() ## Expands the current event table and saves it in ev
etRbind

Combining event tables

Description

Combining event tables

Usage

etRbind(
  ..., 
  samples = c("use", "clear"), 
  waitII = c("smart", "+ii"), 
  id = c("merge", "unique")
)

## S3 method for class 'rxEt'
rbind(..., deparse.level = 1)

Arguments

... The event tables and optionally time between event tables, called waiting times in this help document.
samples How to handle samples when repeating an event table. The options are:
  • "clear" Clear sampling records before combining the datasets
  • "use" Use the sampling records when combining the datasets
waitII This determines how waiting times between events are handled. The options are:
  • "smart" This "smart" handling of waiting times is the default option. In this case, if the waiting time is above the last observed inter-dose interval in the first combined event table, then the actual time between doses is given by the wait time. If it is smaller than the last observed inter-dose interval, the time between event tables is given by the inter-dose interval + the waiting time between event tables.
  • "+ii" In this case, the wait time is added to the inter-dose interval no matter the length of the wait time or inter-dose interval
id This is how rbind will handle IDs. There are two different types of options:
  • merge with id="merge", the IDs are merged together, overlapping IDs would be merged into a single event table.
  • unique with id="unique", the IDs will be renumbered so that the IDs in all the event tables are not overlapping.
deparse.level The deparse.level of a traditional rbind is ignored.
Value

An event table

Author(s)

Matthew L Fidler
Matthew L Fidler, Wenping Wang

References


See Also
eventTable, add.sampling, add.dosing, et, etRep, etRbind, RxODE

Examples

library(RxODE)
library(units)

## Model from RxODE tutorial
mod1 <-RxODE({
  KA=2.94E-01;
  CL=1.86E+01;
  V2=4.02E+01;
  Q=1.05E+01;
  V3=2.97E+02;
  Kin=1;
  Kout=1;
  EC50=200;
  C2 = centr/V2;
  C3 = peri/V3;
  d/dt(depot) =-KA*depot;
  d/dt(centr) = KA*depot - CL*C2 - Q*C2 + Q*C3;
  d/dt(peri) = Q*C2 - Q*C3;
  d/dt(eff) = Kin - Kout*(1-C2/(EC50+C2))*eff;
})

## These are making the more complex regimens of the RxODE tutorial

## bid for 5 days
bid <- et(timeUnits="hr") %>%
et(amt=10000,ii=12,until=set_units(5, "days"))

## qd for 5 days
qd <- et(timeUnits="hr") %>%
et(amt=20000, ii=24, until=set_units(5, "days"))
## bid for 5 days followed by qd for 5 days
et <- seq(bid,qd) %>% et(seq(0,11*24,length.out=100));
bidQd <- rxSolve(mod1, et)
plot(bidQd, C2)

## Now Infusion for 5 days followed by oral for 5 days
## note you can dose to a named compartment instead of using the compartment number
infusion <- et(timeUnits = "hr") %>%
et(amt=10000, rate=5000, ii=24, until=set_units(5, "days"), cmt="centr")
qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(5, "days"), cmt="depot")
et <- seq(infusion,qd)
infusionQd <- rxSolve(mod1, et)
plot(infusionQd, C2)

## 2wk-on, 1wk-off
qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")
et <- seq(qd, set_units(1,"weeks"), qd) %>%
   add.sampling(set_units(seq(0, 5.5,by=0.005),weeks))
wkOnOff <- rxSolve(mod1, et)
plot(wkOnOff, C2)

## You can also repeat the cycle easily with the rep function
qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")
et <- etRep(qd, times=4, wait=set_units(1,"weeks")) %>%
   add.sampling(set_units(seq(0, 12.5,by=0.005),weeks))
repCycle4 <- rxSolve(mod1, et)
plot(repCycle4, C2)
Repeat an RxODE event table

Description
Repeat an RxODE event table

Usage
etRep(
  x,
  times = 1,
  length.out = NA,
  each = NA,
  n = NULL,
  wait = 0,
  id = integer(0),
  samples = c("clear", "use"),
  waitII = c("smart", "+ii"),
  ii = 24
)

## S3 method for class 'rxEt'
rep(x, ...)

Arguments

- **x**: An RxODE event table
- **times**: Number of times to repeat the event table
- **length.out**: Invalid with RxODE event tables, will throw an error if used.
- **each**: Invalid with RxODE event tables, will throw an error if used.
- **n**: The number of times to repeat the event table. Overrides times.
- **wait**: Waiting time between each repeated event table. By default there is no waiting, or wait=0
- **id**: A integer vector of IDs to add or remove from the event table. If the event table is identical for each ID, then you may expand it to include all the IDs in this vector. All the negative IDs in this vector will be removed.
- **samples**: How to handle samples when repeating an event table. The options are:
  - "clear" Clear sampling records before combining the datasets
  - "use" Use the sampling records when combining the datasets
- **waitII**: This determines how waiting times between events are handled. The options are:
• "smart" This "smart" handling of waiting times is the default option. In this case, if the waiting time is above the last observed inter-dose interval in the first combined event table, then the actual time between doses is given by the wait time. If it is smaller than the last observed inter-dose interval, the time between event tables is given by the inter-dose interval + the waiting time between event tables.

• "+ii" In this case, the wait time is added to the inter-dose interval no matter the length of the wait time or inter-dose interval

ii When specifying a dose, this is the inter-dose interval for ss, addl and until options (described below).

... Times or event tables. They can also be one of the named arguments below.

Value
An event table

Author(s)
Matthew L Fidler, Wenping Wang

References

See Also
eventTable, add.sampling, add.dosing, et, etRep, etRbind, RxODE

Examples

library(RxODE)
library(units)

## Model from RxODE tutorial
mod1 <- RxODE(
  KA=2.94E-01;
  CL=1.86E+01;
  V2=4.02E+01;
  Q=1.05E+01;
  V3=2.97E+02;
  Kin=1;
  Kout=1;
  EC50=200;
  C2 = centr/V2;
  C3 = peri/V3;
  C4 = time;
etRep

\[
\begin{align*}
\frac{d}{dt}(\text{depot}) &= -K_A \cdot \text{depot}; \\
\frac{d}{dt}(\text{centr}) &= K_A \cdot \text{depot} - CL \cdot C_2 - Q \cdot C_2 + Q \cdot C_3; \\
\frac{d}{dt}(\text{peri}) &= Q \cdot C_2 - Q \cdot C_3; \\
\frac{d}{dt}(\text{eff}) &= K_{in} - K_{out} \cdot \frac{1 - C_2/(EC_{50} + C_2)}{1 + C_2/(EC_{50} + C_2)} \cdot \text{eff};
\end{align*}
\]

## These are making the more complex regimens of the RxODE tutorial

### bid for 5 days

\[
bid <- \text{et}(\text{timeUnits} = \text{"hr"}) \%>\% \\
\text{et}(\text{amt}=10000, \text{ii}=12, \text{until}=\text{set_units}(5, \text{"days"}))
\]

### qd for 5 days

\[
qd <- \text{et}(\text{timeUnits} = \text{"hr"}) \%>\% \\
\text{et}(\text{amt}=20000, \text{ii}=24, \text{until}=\text{set_units}(5, \text{"days"}))
\]

### bid for 5 days followed by qd for 5 days

\[
et <- \text{seq}(\text{bid}, \text{qd}) \%>\% \text{et}(\text{seq}(0, 11 \times 24, \text{length.out}=100));
\]

\[
bidQd <- \text{rxSolve(mod1, et)}
\]

\[
\text{plot(bidQd, C2)}
\]

### Now Infusion for 5 days followed by oral for 5 days

### note you can dose to a named compartment instead of using the compartment number

\[
\text{infusion} <- \text{et}(\text{timeUnits} = \text{"hr"}) \%>\% \\
\text{et}(\text{amt}=10000, \text{rate}=5000, \text{ii}=24, \text{until}=\text{set_units}(5, \text{"days"}), \text{cmt}=\text{"centr"})
\]

\[
qd <- \text{et}(\text{timeUnits} = \text{"hr"}) \%>\% \text{et}(\text{amt}=10000, \text{ii}=24, \text{until}=\text{set_units}(5, \text{"days"}), \text{cmt}=\text{"depot"})
\]

\[
et <- \text{seq}(\text{infusion}, \text{qd})
\]

\[
\text{infusionQd} <- \text{rxSolve(mod1, et)}
\]

\[
\text{plot(infusionQd, C2)}
\]

### 2wk-on, 1wk-off

\[
qd <- \text{et}(\text{timeUnits} = \text{"hr"}) \%>\% \text{et}(\text{amt}=10000, \text{ii}=24, \text{until}=\text{set_units}(2, \text{"weeks"}), \text{cmt}=\text{"depot"})
\]

\[
et <- \text{seq}(\text{qd}, \text{set_units}(1, \text{"weeks"}), \text{qd}) \%>\% \\
\text{add.sampling(set_units(seq(0, 5.5, by=0.005), weeks))}
\]

\[
\text{wkOnOff} <- \text{rxSolve(mod1, et)}
\]

\[
\text{plot(wkOnOff, C2)}
\]

### You can also repeat the cycle easily with the rep function
qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")
et <- etRep(qd, times=4, wait=set_units(1,"weeks")) %>%
  add.sampling(set_units(seq(0, 12.5,by=0.005),weeks))
repCycle4 <- rxSolve(mod1, et)
plot(repCycle4, C2)

etSeq

Sequence of event tables

Description
This combines a sequence of event tables.

Usage
etSeq(..., samples = c("clear", "use"), waitII = c("smart", "+ii"), ii = 24)

## S3 method for class 'rxEt'
seq(...)

Arguments
...
  The event tables and optionally time between event tables, called waiting times in this help document.
samples
  How to handle samples when repeating an event table. The options are:
    • "clear" Clear sampling records before combining the datasets
    • "use" Use the sampling records when combining the datasets
waitII
  This determines how waiting times between events are handled. The options are:
    • "smart" This "smart" handling of waiting times is the default option. In this case, if the waiting time is above the last observed inter-dose interval in the first combined event table, then the actual time between doses is given by the wait time. If it is smaller than the last observed inter-dose interval, the time between event tables is given by the inter-dose interval + the waiting time between event tables.
    • "+ii" In this case, the wait time is added to the inter-dose interval no matter the length of the wait time or inter-dose interval
ii
  If there was no inter-dose intervals found in the event table, assume that the interdose interval is given by this ii value. By default this is 24.
Details
This sequences all the event tables in added in the argument list . . . . By default when combining the event tables the offset is at least by the last inter-dose interval in the prior event table (or ii). If you separate any of the event tables by a number, the event tables will be separated at least the wait time defined by that number or the last inter-dose interval.

Value
An event table

Author(s)
Matthew L Fidler, Wenping Wang

References

See Also
eventTable, add.sampling, add.dosing, et, etRep, etRbind, RxODE

Examples

```r
library(RxODE)
library(units)

## Model from RxODE tutorial
mod1 <- RxODE(
  KA=2.94E-01;
  CL=1.86E+01;
  V2=4.02E+01;
  Q=1.05E+01;
  V3=2.97E+02;
  Kin=1;
  Kout=1;
  EC50=200;
  C2 = centr/V2;
  C3 = peri/V3;
  d/dt(depot) =-KA*depot;
  d/dt(centr) = KA*depot - CL*C2 - Q*C2 + Q*C3;
  d/dt(peri) = Q*C2 - Q*C3;
  d/dt(eff) = Kin - Kout*(1-C2/(EC50+C2))*eff;
);

## These are making the more complex regimens of the RxODE tutorial
```
## bid for 5 days

bid <- et(timeUnits="hr") %>%
et(amt=10000, ii=12, until=set_units(5, "days"))

## qd for 5 days

qd <- et(timeUnits="hr") %>%
et(amt=20000, ii=24, until=set_units(5, "days"))

## bid for 5 days followed by qd for 5 days

et <- seq(bid, qd) %>% et(seq(0, 11*24, length.out=100));

bidQd <- rxSolve(mod1, et)

plot(bidQd, C2)

## Now Infusion for 5 days followed by oral for 5 days

## note you can dose to a named compartment instead of using the compartment number

infusion <- et(timeUnits = "hr") %>%
et(amt=10000, rate=5000, ii=24, until=set_units(5, "days"), cmt="centr")

qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(5, "days"), cmt="depot")

et <- seq(infusion, qd)

infusionQd <- rxSolve(mod1, et)

plot(infusionQd, C2)

## 2wk-on, 1wk-off

qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")

et <- seq(qd, set_units(1, "weeks"), qd) %>%
  add.sampling(set_units(seq(0, 5.5, by=0.005), weeks))

wkOnOff <- rxSolve(mod1, et)

plot(wkOnOff, C2)

## You can also repeat the cycle easily with the rep function

qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")

et <- etRep(qd, times=4, wait=set_units(1, "weeks")) %>%
  add.sampling(set_units(seq(0, 12.5, by=0.005), weeks))

repCycle4 <- rxSolve(mod1, et)
plot(repCycle4, C2)

---

Create an event table object

**Description**

Initializes an object of class ‘EventTable’ with methods for adding and querying dosing and observation records.

**Usage**

```r
eventTable(amount.units = NA, time.units = NA)
```

**Arguments**

- `amount.units`: string denoting the amount dosing units, e.g., “mg”, “ug”. Default to NA to denote unspecified units. It could also be a solved RxODE object. In that case, `eventTable(obj)` returns the eventTable that was used to solve the RxODE object.
- `time.units`: string denoting the time units, e.g., “hours”, “days”. Default to “hours”.

An `eventTable` is an object that consists of a data.frame storing ordered time-stamped events of an (unspecified) PK/PD dynamic system, units (strings) for dosing and time records, plus a list of functions to add and extract event records. Currently, events can be of two types: dosing events that represent inputs to the system and sampling time events that represent observations of the system with ‘amount.units’ and ‘time.units’, respectively.

**Value**

A modified data.frame with the following accessible functions:

- `get.EventTable()` returns the current event table.
- `add.dosing()` adds dosing records to the event table.
- `get.dosing()` returns a data.frame of dosing records.
- `clear.dosing()` clears or deletes all dosing from event table.
- `add.sampling()` adds sampling time observation records to the event table.
- `get.sampling()` returns a data.frame of sampled observation records.
- `clear.sampling()` removes all sampling from event table.
- `get.obs.rec()` returns a logical vector indicating whether each event record represents an observation or not.
- `get.nobs()` returns the number of observation (not dosing) records.
- `get.units()` returns a two-element character vector with the dosing and time units, respectively.
• `copy()` makes a copy of the current event table. To create a copy of an event table object use `qd2 <- qd$copy()`
• `expand()` Expands the event table for multi-subject solving. This is done by `qd$expand(400)` for a 400 subject data expansion

Author(s)
Matthew Fidler, Melissa Hallow and Wenping Wang

See Also
`et()`, `RxODE()`

Examples
```r
# create dosing and observation (sampling) events
# QD 50mg dosing, 5 days followed by 25mg 5 days
#
qd <- eventTable(amount.units = "mg", time.units = "days")
qd$add.dosing(dose = 50, nbr.doses = 5, dosing.interval = 1, do.sampling = FALSE)
# sample the system's drug amounts hourly the first day, then every 12 hours
# for the next 4 days
qd$add.sampling(seq(from = 0, to = 1, by = 1 / 24))
qd$add.sampling(seq(from = 1, to = 5, by = 12 / 24))
# print(qd$get.dosing()) # table of dosing records
print(qd$get.nobs()) # number of observation (not dosing) records
# BID dosing, 5 days
bid <- eventTable("mg", "days") # only dosing
bid$add.dosing(
  dose = 10000, nbr.doses = 2 * 5,
  dosing.interval = 12, do.sampling = FALSE)
#
# Use the copy() method to create a copy (clone) of an existing
# event table (simple assignments just create a new reference to
# the same event table object (closure)).
#
bid.ext <- bid$copy() # three-day extension for a 2nd cohort
bid.ext$add.dosing(
  dose = 5000, nbr.doses = 2 * 3,
  start.time = 120, dosing.interval = 12, do.sampling = FALSE)
#
# You can also use the Piping operator to create a table

qd2 <- eventTable(amount.units = "mg", time.units = "days") %>%
  add.dosing(dose = 50, nbr.doses = 5, dosing.interval = 1, do.sampling = FALSE) %>%
  add.sampling(seq(from = 0, to = 1, by = 1 / 24)) %>%
```

forderForceBase

add.sampling(seq(from = 1, to = 5, by = 12 / 24))
# print(qd2$get.dosing()) # table of dosing records
print(qd2$get.nobs()) # number of observation (not dosing) records

# Note that piping with %>% will update the original table.
qd3 <- qd2 %>% add.sampling(seq(from = 5, to = 10, by = 6 / 24))
print(qd2$get.nobs())
print(qd3$get.nobs())

forderForceBase  Force using base order for RxODE radix sorting

Description
Force using base order for RxODE radix sorting

Usage
forderForceBase(forceBase = FALSE)

Arguments
forceBase  boolean indicating if RxODE should use R’s order() for radix sorting instead of data.table’s parallel radix sorting.

Value
NILL; called for side effects

Examples
forderForceBase(TRUE) # Use base ‘order’ for RxODE sorts
forderForceBase(FALSE) # Use ‘data.table’ for RxODE sorts

gammap  Gammap: normalized lower incomplete gamma function

Description
This is the gamma_p from the boost library

Usage
gammap(a, z)
Arguments

a  The numeric 'a' parameter in the normalized lower incomplete gamma
z  The numeric 'z' parameter in the normalized lower incomplete gamma

Details

The gamma p function is given by:
gammap = lowergamma(a, z)/gamma(a)

Value

gammap results

Author(s)

Matthew L. Fidler

Examples

gammap(1, 3)
gammap(1:3, 3)
gammap(1, 1:3)

gammapDer(a, z)

Arguments

a  The numeric 'a' parameter in the upper incomplete gamma
z  The numeric 'z' parameter in the upper incomplete gamma

Value

lowergamma results

Author(s)

Matthew L. Fidler
Examples

gammapDer(1:3, 3)
gammapDer(1, 1:3)

gammapInv

Description

gammapInv and gammapInva: Inverses of normalized gammap function

Usage

gammapInv(a, p)
gammapInva(x, p)

Arguments

a The numeric 'a' parameter in the upper incomplete gamma
p The numeric 'p' parameter in the upper incomplete gamma
x The numeric 'x' parameter in the upper incomplete gamma

Details

With the equation:
p = gammap(a, x)

The 'gammapInv' function returns a value 'x' that satisfies the equation above
The 'gammapInva' function returns a value 'q' that satisfies the equation above

NOTE: gammapInva is slow

Value

inverse gammap results

Author(s)

Matthew L. Fidler
Examples

gammaInv(1:3, 0.5)
gammaInv(1, 1:3 / 3.1)
gammaInv(1:3, 1:3 / 3.1)
gammaInva(1:3, 1:3 / 3.1)

gammaq

Gammaq: normalized upper incomplete gamma function

Description
This is the gamma_q from the boost library

Usage
gammaq(a, z)

Arguments

a  The numeric ‘a’ parameter in the normalized upper incomplete gamma
z  The numeric ‘z’ parameter in the normalized upper incomplete gamma

Details
The gamma q function is given by:
gammaq = uppergamma(a, z)/gamma(a)

Value
gammaq results

Author(s)
Matthew L. Fidler

Examples

gammaq(1, 3)
gammaq(1:3, 3)
gammaq(1, 1:3)
**Description**

gammaqInv and gammaqInva: Inverses of normalized gammaq function

**Usage**

```r
gammaqInv(a, q)
gammaqInva(x, q)
```

**Arguments**

- `a`: The numeric ‘a’ parameter in the upper incomplete gamma
- `q`: The numeric ‘q’ parameter in the upper incomplete gamma
- `x`: The numeric ‘x’ parameter in the upper incomplete gamma

**Details**

With the equation:

\[ q = \gamma(a, x) \]

The ‘gammaInv’ function returns a value ‘x’ that satisfies the equation above

The ‘gammaInva’ function returns a value ‘a’ that satisfies the equation above

**Value**

inverse gammaq results

**Author(s)**

Matthew L. Fidler

**Examples**

```r
gammaqInv(1:3, 0.5)
gammaqInv(1, 1:3 / 3)
gammaqInv(1:3, 1:3 / 3.1)
gammaqInva(1:3, 1:3 / 3.1)
```
Generate an example (template) of a dosing regimen shiny app

Description
Create a complete shiny application for exploring dosing regimens given a (hardcoded) PK/PD model.

Usage
genShinyApp.template(appDir = "shinyExample", verbose = TRUE, ODE.config = list(ode = "model", params = c(KA = 0.294), inits = c(eff = 1), method = "lsoda", atol = 1e-08, rtol = 1e-06))
write.template.server(appDir)
write.template.ui(appDir, statevars)

Arguments
appDir a string with a directory where to store the shiny app, by default is "shinyExample". The directory appDir will be created if it does not exist.
verbose logical specifying whether to write messages as the shiny app is generated. Defaults to TRUE.
ODE.config model name compiled and list of parameters sent to rxSolve().
statevars List of statevars passed to the write.template.ui() function. This usually isn’t called directly.

A PK/PD model is defined using RxODE(), and a set of parameters and initial values are defined. Then the appropriate R scripts for the shiny’s user interface ui.R and the server logic server.R are created in the directory appDir.
The function evaluates the following PK/PD model by default:
C2 = centr/V2;
C3 = peri/V3;
d/dt(depot) = -KA*depot;
d/dt(centr) = KA*depot - CL*C2 - Q*C2 + Q*C3;
d/dt(peri) = Q*C2 - Q*C3;
d/dt(eff) = Kin - Kout*(1-C2/(EC50+C2))*eff;

This can be changed by the ODE.config parameter.
To launch the shiny app, simply issue the runApp(appDir) R command.

Value
None, these functions are used for their side effects.
Note

These functions create a simple, but working example of a dosing regimen simulation web application. Users may want to modify the code to experiment creating shiny applications for their specific RxODE models.

See Also

RxODE(), eventTable(), and the package shiny (https://shiny.rstudio.com).

Examples

```r
# create the shiny app example (template)
genShinyApp.template(appDir = "myapp")
# run the shiny app
library(shiny)
# runApp("myapp") # Won't launch in environments without browsers
unlink("myapp", recursive = TRUE, force = TRUE) # remove myapp
```

---

getRxThreads

Get/Set the number of threads that RxODE uses

Description

Get/Set the number of threads that RxODE uses

Usage

```r
getRxThreads(VERBOSE = FALSE)
setRxThreads(threads = NULL, percent = NULL, throttle = NULL)
rxCores(VERBOSE = FALSE)
```

Arguments

- **verbose**
  Display the value of relevant OpenMP settings

- **threads**
  NULL (default) rereads environment variables. 0 means to use all logical CPUs available. Otherwise a number >= 1

- **percent**
  If provided it should be a number between 2 and 100; the percentage of logical CPUs to use. By default on startup, 50 percent.

- **throttle**
  2 (default) means that, roughly speaking, a single thread will be used when number subjects solved for is <=2, 2 threads when the number of all points is <=4, etc. The throttle is to speed up small data tasks (especially when repeated many times) by not incurring the overhead of managing multiple threads.
The throttle will also suppress sorting which ID will be solved first when there are (nsubject solved)*throttle <= nthreads. In RxODE this sorting occurs to minimize the time for waiting for another thread to finish. If the last item solved is has a long solving time, all the other solving have to wait for that last costly solving to occur. If the items which are likely to take more time are solved first, this wait is less likely to have an impact on the overall solving time.

In RxODE the IDs are sorted by the individual number of solving points (largest first). It also has a C interface that allows these IDs to be resorted by total time spent solving the equation. This allows packages like nlmixr to sort by solving time if needed.

Overall the the number of threads is throttled (restricted) for small tasks and sorting for IDs are suppressed.

Value

number of threads that RxODE uses

guide_none

Empty Guide

Description

This empty guide draws nothing; It is included in RxODE for compatibility with ggplot 3.2

Usage

guide_none(title = waiver(), position = waiver())

Arguments

title A character string or expression indicating a title of guide. If NULL, the title is not shown. By default (waiver()), the name of the scale object or the name specified in labs() is used for the title.

position Where this guide should be drawn: one of top, bottom, left, or right.

Value

nothing, simply included to be compatible with ggplot 3.2
invWR1d  

One correlation sample from the Inverse Wishart distribution

Description

This correlation is constructed by transformation of the Inverse Wishart random covariate to a correlation.

Usage

invWR1d(d, nu, omegaIsChol = FALSE)

Arguments

d  The dimension of the correlation matrix
nu  Degrees of freedom of the Wishart distribution
omegaIsChol  is an indicator of if the omega matrix is in the Cholesky decomposition. This is only used when codetype="invWishart"

Value

One correlation sample from the inverse wishart

Author(s)

Matthew Fidler

is.rxEt  

Check to see if this is an rxEt object.

Description

Check to see if this is an rxEt object.

Usage

is.rxEt(x)

Arguments

x  object to check to see if it is rxEt

If this is an rxEt object that has expired strip all rxEt information.

Value

Boolean indicating if this is a RxODE event table
is.rxSolve  
Check to see if this is an rxSolve object.

Description
Check to see if this is an rxSolve object.

Usage
is.rxSolve(x)

Arguments
- x: object to check to see if it is rxSolve
  If this is an rxSolve object that has expired strip all rxSolve information.

Value
boolean indicating if this is a rxSolve object

Author(s)
Matthew L.Fidler

logit  
logit and inverse logit (expit) functions

Description
logit and inverse logit (expit) functions

Usage
logit(x, low = 0, high = 1)

expit(alpha, low = 0, high = 1)

logitNormInfo(mean = 0, sd = 1, low = 0, high = 1, abs.tol = 1e-06, ...)

probitNormInfo(mean = 0, sd = 1, low = 0, high = 1, abs.tol = 1e-06, ...)
**logit**

**Arguments**

- **x**: Input value(s) in range [low, high] to translate -Inf to Inf
- **low**: Lowest value in the range
- **high**: Highest value in the range
- **alpha**: Infinite value(s) to translate to range of [low, high]
- **mean**: Logit-scale mean
- **sd**: Logit-scale standard deviation
- **abs.tol**: Absolute accuracy requested.
- ...: Other parameters passed to `integrate()`

**Details**

Logit is given by:

\[
\text{logit}(p) = \log\left(\frac{1}{p} - 1\right)
\]

where:

\[
p = x - \text{low}/\text{high} - \text{low}
\]

Expit is given by:

\[
\text{expit}(p, \text{low}, \text{high}) = \frac{\text{high} - \text{low}}{1 + \exp(-\alpha)} + \text{low}
\]

The `logitNormInfo()` gives the mean, variance and coefficient of variability on the untransformed scale.

**Value**

Values from logit and expit

**Examples**

- `logit(0.25)`
- `expit(-1.09)`
- `logitNormInfo(logit(0.25), sd = 0.1)`
- `logitNormInfo(logit(1, 0, 10), sd = 1, low = 0, high = 10)`
lowergamma  

**lowergamma: upper incomplete gamma function**

**Description**

This is the tgamma_lower from the boost library

**Usage**

`lowergamma(a, z)`

**Arguments**

- `a` The numeric ‘a’ parameter in the upper incomplete gamma
- `z` The numeric ‘z’ parameter in the upper incomplete gamma

**Details**

The lowergamma function is given by:

\[
lowergamma(a, z) = \int_{0}^{z} t^{a-1} e^{-t} dt
\]

**Value**

lowergamma results

**Author(s)**

Matthew L. Fidler

**Examples**

- `lowergamma(1, 3)`
- `lowergamma(1:3, 3)`
- `lowergamma(1, 1:3)`
**Description**
Cumulative distribution of standard normal

**Usage**
\[ \phi(q) \]

**Arguments**
- \( q \) vector of quantiles.

**Value**
cumulative distribution of standard normal distribution

**Author(s)**
Matthew Fidler

**Examples**

```r
# phi is equivalent to pnorm(x)
phi(3)

# See
pnorm(3)

# This is provided for NONMEM-like compatibility in RxODE models
```

---

**print.RxODE**

Print information about the RxODE object.

**Description**
This prints the model name and its status for being able to be solved

**Usage**

```
## S3 method for class 'RxODE'
print(x, ...)
```
Arguments

- **x**  
  An rxode object

- **...**  
  Ignored parameters

Value

- **original object**

Author(s)

- Matthew L. Fidler

---

**probit**  
*probit and inverse probit functions*

Description

probit and inverse probit functions

Usage

```r
probit(x, low = 0, high = 1)
probitInv(x, low = 0, high = 1)
```

Arguments

- **x**  
  Input value(s) in range [low,high] to translate -Inf to Inf

- **low**  
  Lowest value in the range

- **high**  
  Highest value in the range

Value

values from probit, probitInv and probitNormInfo

Examples

```r
probit(0.25)
probitInv(-0.674)
probitNormInfo(probit(0.25), sd = 0.1)
probitNormInfo(probit(1, 0, 10), sd = 1, low = 0, high = 10)
```
Scaled Inverse Chi Squared distribution

Description
Scaled Inverse Chi Squared distribution

Usage
rinvchisq(n = 1L, nu = 1, scale = 1)

Arguments
n Number of random samples
nu degrees of freedom of inverse chi square
scale Scale of inverse chi squared distribution (default is 1).

Value
a vector of inverse chi squared deviates.

Examples
rinvchisq(3, 4, 1) ## Scale = 1, degrees of freedom = 4
rinvchisq(2, 4, 2) ## Scale = 2, degrees of freedom = 4

One correlation sample from the LKJ distribution

Description
One correlation sample from the LKJ distribution

Usage
rLKJ1(d, eta = 1, cholesky = FALSE)

Arguments
d The dimension of the correlation matrix
eta The scaling parameter of the LKJ distribution. Must be > 1. Also related to the
degrees of freedom nu. eta = (nu-1)/2.
cholesky boolean; If TRUE return the cholesky decomposition.
Value

A correlation sample from the LKJ distribution

Author(s)

Matthew Fidler (translated to RcppArmadillo) and Emma Schwager

----------

`rxAddReturn`  
*Add a return statement to a function.*

----------

Description

Add a return statement to a function.

Usage

`rxAddReturn(fn, ret = TRUE)`

Arguments

- `fn`  
  Function to deparse
- `ret`  
  boolean stating if a return statement will be added.

Value

Function with parens removed and add a return statement.

Author(s)

Matthew L. Fidler

----------

`rxAllowUnload`  
*Allow unloading of dlls*

----------

Description

Allow unloading of dlls

Usage

`rxAllowUnload(allow)`

Arguments

- `allow`  
  boolean indicating if garbage collection will unload of RxODE dlls.
**rxAssignPtr**

**Value**

Boolean allow; called for side effects

**Author(s)**

Matthew Fidler

**Examples**

```r
# Garbage collection will not unload un-used RxODE dlls
rxAllowUnload(FALSE);

# Garbage collection will unload unused RxODE dlls
rxAllowUnload(TRUE);
```

---

**rxAssignPtr**  
**Assign pointer based on model variables**

**Description**

Assign pointer based on model variables

**Usage**

`rxAssignPtr(object = NULL)`

**Arguments**

- **object**  
  RxODE family of objects

**Value**

nothing, called for side effects
rxbeta Simulate beta variable from threefry generator

Description

Care should be taken with this method not to encounter the birthday problem, described https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/. Since the sitmo threefry, this currently generates one random deviate from the uniform distribution to seed the engine threefry and then run the code.

Usage

rxbeta(shape1, shape2, n = 1L, ncores = 1L)

Arguments

- **shape1**: non-negative parameters of the Beta distribution.
- **shape2**: non-negative parameters of the Beta distribution.
- **n**: number of observations. If `length(n) > 1`, the length is taken to be the number required.
- **ncores**: Number of cores for the simulation

rxnorm simulates using the threefry sitmo generator; rxnormV uses the vander-corput generator

Details

Therefore, a simple call to the random number generated followed by a second call to random number generated may have identical seeds. As the number of random number generator calls are increased the probability that the birthday problem will increase.

The key to avoid this problem is to either run all simulations in the RxODE environment once (therefore one seed or series of seeds for the whole simulation), pre-generate all random variables used for the simulation, or seed the RxODE engine with rxSetSeed()

Also care should be made that the computer you will be running on can run the same number of cores as you are running so they can reproduce your results.

Value

beta random deviates

Examples

```r
## Use threefry engine
rxbeta(0.5, 0.5, n = 10) # with rxbeta you have to explicitly state n
```
rxbinom

rxbeta(5, 1, n = 10, ncores = 2) # You can parallelize the simulation using openMP
rxbeta(1, 3)

## This example uses `rxbeta` directly in the model

```r
rx <- RxODE({
a <- rxbeta(2, 2)
})
et <- et(1, id = 1:2)
s <- rxSolve(rx, et)
```

---

### Description

Care should be taken with this method not to encounter the birthday problem, described [https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/](https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/). Since the sitmo threefry, this currently generates one random deviate from the uniform distribution to seed the engine threefry and then run the code.

### Usage

```r
rxbinom(size, prob, n = 1L, ncores = 1L)
```

### Arguments

- **size**: number of trials (zero or more).
- **prob**: probability of success on each trial.
- **n**: number of observations. If `length(n) > 1`, the length is taken to be the number required.
- **ncores**: Number of cores for the simulation

*rxnorm* simulates using the threefry sitmo generator; *rxnormV* uses the vander-corput generator

### Details

Therefore, a simple call to the random number generated followed by a second call to random number generated may have identical seeds. As the number of random number generator calls are increased the probability that the birthday problem will increase.

The key to avoid this problem is to either run all simulations in the RxODE environment once (therefore one seed or series of seeds for the whole simulation), pre-generate all random variables used for the simulation, or seed the RxODE engine with `rxSetSeed()`
Also care should be made that the computer you will be running on can run the same number of cores as you are running so they can reproduce your results.

Value

binomial random deviates

Examples

## Use threefry engine

```r
rxbinom(10, 0.9, n = 10) # with rxbinom you have to explicitly state n
rxbinom(3, 0.5, n = 10, ncores = 2) # You can parallelize the simulation using openMP
rxbinom(4, 0.7)
```

## This example uses `rxbinom` directly in the model

```r
rx <- RxODE({
  a <- rxbinom(1, 0.5)
})

et <- et(1, id = 1:2)

s <- rxSolve(rx, et)
```

---

### rxcauchy

**Simulate Cauchy variable from threefry generator**

Description

Care should be taken with this method not to encounter the birthday problem, described [https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/](https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/). Since the sitmo threefry, this currently generates one random deviate from the uniform distribution to seed the engine threefry and then run the code.

Usage

```r
rxcauchy(location = 0, scale = 1, n = 1L, ncores = 1L)
```

Arguments

- `location`: location and scale parameters.
- `scale`: location and scale parameters.
The function `rxcauchy` simulates Cauchy random deviates.

**Usage**

```r
rxcauchy(0, 1, n = 10) # with rxcauchy you have to explicitly state n
rxcauchy(0.5, n = 10, ncores = 2) # You can parallelize the simulation using openMP
```

```r
rxcauchy(3)
```

**Examples**

```r
## This example uses `rxcauchy` directly in the model

```r
rx <- RxODE(
  a <- rxcauchy(2)
)
```r
et <- et(1, id = 1:2)
```r
s <- rxSolve(rx, et)
```
**rxCbindStudyIndividual**

*Bind the study parameters and individual parameters*

**Description**

Bind the study parameters and individual parameters

**Usage**

\[
rxCbindStudyIndividual(studyParameters, individualParameters)
\]

**Arguments**

- **studyParameters**
  
  These are the study parameters, often can be generated by sampling from a population. This can be either a matrix or a data frame

- **individualParameters**
  
  A data frame of individual parameters

**Value**

Data frame that can be used in RxODE simulations

**Author(s)**

Matthew Fidler

**Examples**

```r
# Function for converting coefficient of covariance into a variance
lognCv <- function(x){log((x/100)^2+1)}

set.seed(32)

nSub <- 100
nStud <- 10

# define theta
theta <- c(lka=log(0.5), # log ka
           lCl=log(5), # log Cl
           lV=log(300) # log V
)

# define theta Matrix
thetaMat <- lotri(lCl ~ lognCv(5),
                  lV ~ lognCv(5),
                  lka ~ lognCv(5))
```
nev <- nSub*nStud

ev1 <- data.frame(COV1=rnorm(nev,50,30), COV2=rnorm(nev,75,10),
                  COV3=sample(c(1.0,2.0),nev,replace=TRUE))

tmat <- rxRmvn(nStud, theta[dimnames(thetaMat)[[1]]], thetaMat)

rcBindStudyIndividual(tmat, ev1)

---

**rxChain**

**rxChain Chain or add item to solved system of equations**

**Description**

Add item to solved system of equations

**Usage**

rxChain(obj1, obj2)

```r
## S3 method for class 'solveRxDll'
obj1 + obj2
```

**Arguments**

- **obj1**  
  Solved object.

- **obj2**  
  New object to be added/piped/chained to solved object.

**Value**

When newObject is an event table, return a new solved object with the new event table.

**Author(s)**

Matthew L. Fidler
**rxchisq**

*Simulate chi-squared variable from threelfry generator*

**Description**

Care should be taken with this method not to encounter the birthday problem, described [https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/](https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/). Since the sitmo threelfry, this currently generates one random deviate from the uniform distribution to seed the engine threelfry and then run the code.

**Usage**

```r
taxchisq(df, n = 1L, ncores = 1L)
```

**Arguments**

- `df`: degrees of freedom (non-negative, but can be non-integer).
- `n`: number of observations. If `length(n) > 1`, the length is taken to be the number required.
- `ncores`: Number of cores for the simulation

In `rxnorm` simulates using the threelfry sitmo generator; `rxnormV` uses the vandercorput generator

**Details**

Therefore, a simple call to the random number generated followed by a second call to random number generated may have identical seeds. As the number of random number generator calls are increased the probability that the birthday problem will increase.

The key to avoid this problem is to either run all simulations in the RxODE environment once (therefore one seed or series of seeds for the whole simulation), pre-generate all random variables used for the simulation, or seed the RxODE engine with `rxSetSeed()`.

Also care should be made that the computer you will be running on can run the same number of cores as you are running so they can reproduce your results.

**Value**

chi squared random deviates

**Examples**

```r
## Use threelfry engine

rxchisq(0.5, n = 10) # with rxchisq you have to explicitly state n
rxchisq(5, n = 10, ncores = 2) # You can parallelize the simulation using openMP
```
### Example

```r
## This example uses `rxchisq` directly in the model
rx <- RxODE({
a <- rxchisq(2)
})
et <- et(1, id = 1:2)
s <- rxSolve(rx, et)
```

---

**rxClean**

Cleanup anonymous DLLs by unloading them

**Description**

This cleans up any RxODE loaded DLLs

**Usage**

```r
rxClean(wd)
```

**Arguments**

- `wd` What directory should be cleaned; (DEPRECATED), this no longer does anything. This unloads all RxODE anonymous dlls.

**Value**

TRUE if successful

**Author(s)**

Matthew L. Fidler
rxCompile

Compile a model if needed

Description

This is the compilation workhorse creating the RxODE model DLL files.

Usage

rxCompile(model, dir, prefix, force = FALSE, modName = NULL, package = NULL, ...)

## S3 method for class 'rxModelVars'
rxCompile(model, dir = NULL, prefix = NULL, force = FALSE, modName = NULL, package = NULL, ...)  

## S3 method for class 'character'
rxCompile(model, dir = NULL, prefix = NULL, force = FALSE, modName = NULL, package = NULL, ...)  

## S3 method for class 'rxDll'
rxCompile(model, ...)  

## S3 method for class 'RxODE'
rxCompile(model, ...)
Arguments

- **model**
  - This is the ODE model specification. It can be:
    - a string containing the set of ordinary differential equations (ODE) and other expressions defining the changes in the dynamic system.
    - a file name where the ODE system equation is contained
  - An ODE expression enclosed in `{}` (see also the filename argument). For details, see the sections “Details” and RxODE Syntax below.

- **dir**
  - This is the model directory where the C file will be stored for compiling.
  - If unspecified, the C code is stored in a temporary directory, then the model is compiled and moved to the current directory. Afterwards the C code is removed.
  - If specified, the C code is stored in the specified directory and then compiled in that directory. The C code is not removed after the DLL is created in the same directory. This can be useful to debug the c-code outputs.

- **prefix**
  - is a string indicating the prefix to use in the C based functions. If missing, it is calculated based on file name, or md5 of parsed model.

- **force**
  - is a boolean stating if the (re)compile should be forced if RxODE detects that the models are the same as already generated.

- **modName**
  - a string to be used as the model name. This string is used for naming various aspects of the computations, including generating C symbol names, dynamic libraries, etc. Therefore, it is necessary that modName consists of simple ASCII alphanumeric characters starting with a letter.

- **package**
  - Package name for pre-compiled binaries.

- **...**
  - Other arguments sent to the `rxTrans()` function.

Value

An rxDll object that has the following components

- dllDLL path
- modelmodel specification
- .cA function to call C code in the correct context from the DLL using the `.C()` function.
- .callA function to call C code in the correct context from the DLL using the `.Call()` function.
- argsA list of the arguments used to create the rxDll object.

Author(s)

Matthew L. Fidler

See Also

- RxODE()
**rxCreateCache**

This will create the cache directory for RxODE to save between sessions

**Description**

When run, if the `R_user_dir` for RxODE’s cache isn’t present, create the cache

**Usage**

```r
taxCreateCache()
```

**Value**

nothing

**Author(s)**

Matthew Fidler

---

**rxD**

Add to RxODE’s derivative tables

**Description**

Add to RxODE’s derivative tables

**Usage**

```r
taxD(name, derivatives)
```

**Arguments**

- **name**
  - Function Name

- **derivatives**
  - A list of functions. Each function takes the same number of arguments as the original function. The first function will construct the derivative with respect to the first argument; The second function will construct the derivative with respect to the second argument, and so on.

**Value**

nothing

**Author(s)**

Matthew Fidler
Examples

```r
## Add an arbitrary list of derivative functions
## In this case the fun(x,y) is assumed to be 0.5*x^2+0.5*y^2

rxD("fun", list(
  function(x, y) {
    return(x)
  },
  function(x, y) {
    return(y)
  }
))
```

```
rxDelete  Delete the DLL for the model
```

Description

This function deletes the DLL, but doesn’t delete the model information in the object.

Usage

```r
rxDelete(obj)
```

Arguments

| obj | RxODE family of objects |

Value

A boolean stating if the operation was successful.

Author(s)

Matthew L.Fidler

```
rxDerived  Calculate derived parameters for the 1-, 2-, and 3- compartment linear models.
```

Description

This calculates the derived parameters based on what is provided in a data frame or arguments

Usage

```r
rxDerived(..., verbose = FALSE, digits = 0)
```
Arguments

The input can be:

- A data frame with PK parameters in it; This should ideally be a data frame with one pk parameter per row since it will output a data frame with one PK parameter per row.
- PK parameters as either a vector or a scalar

verbose boolean that when TRUE provides a message about the detected pk parameters and the detected compartmental model. By default this is FALSE.

digits represents the number of significant digits for the output; If the number is zero or below (default), do not round.

Value

Return a data.frame of derived PK parameters for a 1-, 2-, or 3-compartment linear model given provided clearances and volumes based on the inferred model type.

The model parameters that will be provided in the data frame are:

- vc: Central Volume (for 1-, 2- and 3-compartment models)
- kel: First-order elimination rate (for 1-, 2-, and 3-compartment models)
- k12: First-order rate of transfer from central to first peripheral compartment; (for 2- and 3-compartment models)
- k21: First-order rate of transfer from first peripheral to central compartment, (for 2- and 3-compartment models)
- k13: First-order rate of transfer from central to second peripheral compartment; (3-compartment model)
- k31: First-order rate of transfer from second peripheral to central compartment (3-compartment model)
- vp: Peripheral Volume (for 2- and 3-compartment models)
- vp2: Peripheral Volume for 3rd compartment (3-compartment model)
- vss: Volume of distribution at steady state; (1-, 2-, and 3-compartment models)
- t12alpha: t1/2,α; (1-, 2-, and 3-compartment models)
- t12beta: t1/2,β; (2- and 3-compartment models)
- t12gamma: t1/2,γ; (3-compartment model)
- alpha: α; (1-, 2-, and 3-compartment models)
- beta: β; (2- and 3-compartment models)
- gamma: β; (3-compartment model)
- A: true A; (1-, 2-, and 3-compartment models)
- B: true B; (2- and 3-compartment models)
- C: true C; (3-compartment model)
- fracA: fractional A; (1-, 2-, and 3-compartment models)
- fracB: fractional B; (2- and 3-compartment models)
- fracC: fractional C; (3-compartment model)
**rxDfdy**

**Author(s)**

Matthew Fidler and documentation from Justin Wilkins, <justin.wilkins@occams.com>

**References**

Shafer S. L. *CONVERT.XLS*


**Examples**

```r
## Note that RxODE parses the names to figure out the best PK parameter
params <- rxDerived(cl = 29.4, v = 23.4, Vp = 114, vp2 = 4614, q = 270, q2 = 73)

## That is why this gives the same results as the value before
params <- rxDerived(CL = 29.4, V1 = 23.4, V2 = 114, V3 = 4614, Q2 = 270, Q3 = 73)

## You may also use micro-constants alpha/beta etc.
params <- rxDerived(k12 = 0.1, k21 = 0.2, k13 = 0.3, k31 = 0.4, kel = 10, v = 10)

## or you can mix vectors and scalars
params <- rxDerived(CL = 29.4, V = 1:3)

## If you want, you can round to a number of significant digits
## with the 'digits' argument:
params <- rxDerived(CL = 29.4, V = 1:3, digits = 2)
```

---

**rxDfdy**

*Jacobian and parameter derivatives*

**Description**

Return Jacobian and parameter derivatives

**Usage**

```r
rxDfdy(obj)
```

**Arguments**

- **obj**
  
  RxODE family of objects
Value
A list of the jacobian parameters defined in this RxODE object.

Author(s)
Matthew L. Fidler

Description
This is to make an EVID more readable by non pharmacometricians. It displays what each means and allows it to be displayed in a tibble.

Usage
rxEvid(x)

as.rxEvid(x)

## S3 method for class 'rxEvid'
c(x, ...)

## S3 method for class 'rxEvid'
x[...]

## S3 method for class 'rxEvid'
as.character(x, ...)

## S3 method for class 'rxEvid'
x[[...]]

## S3 method for class 'rxEvid'
type_sum(x)

pillar_shaft.rxEvid(x, ...)

## S3 method for class 'rxRateDur'
c(x, ...)

## S3 method for class 'rxEvid'
format(x, ...)

## S3 method for class 'rxRateDur'
format(x, ...)
## S3 method for class 'rxEvid'
print(x, ...)

**Arguments**

- **x**  
  Item to be converted to a RxODE EVID specification.
- **...**  
  Other parameters

**Value**

rxEvid specification

**Examples**

```r
rxEvid(1:7)
```

---

### rxexp

*Simulate exponential variable from threefry generator*

**Description**

Care should be taken with this method not to encounter the birthday problem, described [https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/](https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/). Since the sitmo threefry, this currently generates one random deviate from the uniform distribution to seed the engine threefry and then run the code.

**Usage**

```r
rxexp(rate, n = 1L, ncores = 1L)
```

**Arguments**

- **rate**  
  vector of rates.
- **n**  
  number of observations. If `length(n) > 1`, the length is taken to be the number required.
- **ncores**  
  Number of cores for the simulation

`rxnorm` simulates using the threefry sitmo generator; `rxnormV` uses the vander-corput generator
Details

Therefore, a simple call to the random number generated followed by a second call to random number generated may have identical seeds. As the number of random number generator calls are increased the probability that the birthday problem will increase.

The key to avoid this problem is to either run all simulations in the RxODE environment once (therefore one seed or series of seeds for the whole simulation), pre-generate all random variables used for the simulation, or seed the RxODE engine with rxSetSeed().

Also care should be made that the computer you will be running on can run the same number of cores as you are running so they can reproduce your results.

Value

exponential random deviates

Examples

```r
## Use threefry engine
rxexp(0.5, n = 10) # with rxexp you have to explicitly state n
rxexp(5, n = 10, ncores = 2) # You can parallelize the simulation using openMP
rxexp(1)

## This example uses `rxexp` directly in the model
rx <- RxODE({
  a <- rxexp(2)
})
et <- et(1, id = 1:2)
s <- rxSolve(rx, et)
```

---

**Simulate F variable from threefry generator**

Description

Care should be taken with this method not to encounter the birthday problem, described [https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/](https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/). Since the sitmo threefry, this currently generates one random deviate from the uniform distribution to seed the engine threefry and then run the code.
Usage

```r
rxf(df1, df2, n = 1L, ncores = 1L)
```

Arguments

- `df1`: degrees of freedom. Inf is allowed.
- `df2`: degrees of freedom. Inf is allowed.
- `n`: number of observations. If `length(n) > 1`, the length is taken to be the number required.
- `ncores`: Number of cores for the simulation

`rxnorm` simulates using the threefry sitmo generator; `rxnormV` uses the vandercorput generator

Details

Therefore, a simple call to the random number generated followed by a second call to random number generated may have identical seeds. As the number of random number generator calls are increased the probability that the birthday problem will increase.

The key to avoid this problem is to either run all simulations in the RxODE environment once (therefore one seed or series of seeds for the whole simulation), pre-generate all random variables used for the simulation, or seed the RxODE engine with `rxSetSeed()`.

Also care should be made that the computer you will be running on can run the same number of cores as you are running so they can reproduce your results.

Value

f random deviates

Examples

```r
## Use threefry engine

rxf(0.5, 0.5, n = 10) # with rxf you have to explicitly state n
rxf(5, 1, n = 10, ncores = 2) # You can parallelize the simulation using openMP

rxf(1, 3)

## This example uses `rxf` directly in the model

rx <- RxODE(
  a <- rxf(2, 2)
)

et <- et(1, id = 1:2)

s <- rxSolve(rx, et)
```
rxFun

Description

This adds a user function to RxODE that can be called. If needed, these functions can be differentiated by numerical differences or by adding the derivatives to RxODE’s internal derivative table with rxD().

Usage

```r
rxFun(name, args, cCode)
```

```r
rxRmFun(name)
```

Arguments

- `name`: This gives the name of the user function
- `args`: This gives the arguments of the user function
- `cCode`: This is the C-code for the new function

Value

nothing

Author(s)

Matthew L. Fidler

Examples

```r
## Right now RxODE is not aware of the function f
## Therefore it cannot translate it to symengine or
## Compile a model with it.

try(RxODE("a=fun(a,b,c)"))

## Note for this approach to work, it cannot interfere with C
## function names or reserved RxODE special terms. Therefore
## f(x) would not work since f is an alias for bioaviability.

fun <- "
double fun(double a, double b, double c) {
    return a*a+b*a+c;
```
## C-code for function

```r
rxFun("fun", c("a", "b", "c"), fun) ## Added function
```

## Now RxODE knows how to translate this function to symengine

```r
rxToSE("fun(a,b,c)"
```

## And will take a central difference when calculating derivatives

```r
rxFromSE("Derivative(fun(a,b,c),a)"
```

## Of course, you could specify the derivative table manually

```r
rxD("fun", list(
    function(a, b, c) {
      paste0("2\times", a, "+", b)
    },
    function(a, b, c) {
      return(a)
    },
    function(a, b, c) {
      return("0.0")
    }
  ))
```

```r
rxFromSE("Derivative(fun(a,b,c),a)"
```

# You can also remove the functions by `rxRmFun`

```r
rxRmFun("fun")
```

---

**rxgamma**

*Simulate gamma variable from threefry generator*

### Description

Care should be taken with this method not to encounter the birthday problem, described [here](https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/). Since the sitmo threefry, this currently generates one random deviate from the uniform distribution to seed the engine threefry and then run the code.

### Usage

```r
rxgamma(shape, rate = 1/scale, scale = 1, n = 1L, ncores = 1L)
```
Arguments

- **shape**: shape and scale parameters. Must be positive, scale strictly.
- **rate**: an alternative way to specify the scale.
- **scale**: shape and scale parameters. Must be positive, scale strictly.
- **n**: number of observations. If `length(n) > 1`, the length is taken to be the number required.
- **ncores**: Number of cores for the simulation

Details

Therefore, a simple call to the random number generated followed by a second call to random number generated may have identical seeds. As the number of random number generator calls are increased the probability that the birthday problem will increase.

The key to avoid this problem is to either run all simulations in the RxODE environment once (therefore one seed or series of seeds for the whole simulation), pre-generate all random variables used for the simulation, or seed the RxODE engine with `rxSetSeed()`.

Also care should be made that the computer you will be running on can run the same number of cores as you are running so they can reproduce your results.

Value

gamma random deviates

Examples

```r
## Use threefry engine
rxgamma(0.5, n = 10) # with rxgamma you have to explicitly state n
rxgamma(5, n = 10, ncores = 2) # You can parallelize the simulation using openMP
rxgamma(1)

## This example uses `rxbeta` directly in the model
rx <- RxODE(
  a <- rxgamma(2)
)

et <- et(1, id = 1:2)
s <- rxSolve(rx, et)
```
Generate pred-only SAEM RxODE model

Usage

```r
rxGenSaem(
  obj,
  predfn,
  pkpars = NULL,
  sum.prod = FALSE,
  optExpression = TRUE,
  loadSymengine = TRUE
)
```

Arguments

- `obj`: RxODE model (text or actual model)
- `predfn`: prediction function
- `pkpars`: PKpars function
- `sum.prod`: Change addition and deletion to sum() and prod() to stabilize round off errors
- `optExpression`: Boolean for optimizing expression to minimize more expensive calls
- `loadSymengine`: Boolean indicating if the model should be loaded into symengine. This cause all the ODEs to be collapsed into one expression that is eventually optimized if optExpression is TRUE.

Value

RxODE text

Author(s)

Matthew Fidler
**Description**

Care should be taken with this method not to encounter the birthday problem, described [https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/](https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/). Since the sitmo threefry, this currently generates one random deviate from the uniform distribution to seed the engine threefry and then run the code.

**Usage**

```r
rxgeom(prob, n = 1L, ncores = 1L)
```

**Arguments**

- `prob` probability of success in each trial. \(0 < \text{prob} \leq 1\).
- `n` number of observations. If `length(n) > 1`, the length is taken to be the number required.
- `ncores` Number of cores for the simulation
  
  - `rxnorm` simulates using the threefry sitmo generator; `rxnormV` uses the vander-corput generator

**Details**

Therefore, a simple call to the random number generated followed by a second call to random number generated may have identical seeds. As the number of random number generator calls are increased the probability that the birthday problem will increase.

The key to avoid this problem is to either run all simulations in the RxODE environment once (therefore one seed or series of seeds for the whole simulation), pre-generate all random variables used for the simulation, or seed the RxODE engine with `rxSetSeed()`

Also care should be made that the computer you will be running on can run the same number of cores as you are running so they can reproduce your results.

**Value**

geometric random deviates

**Examples**

```r
## Use threefry engine
	rxgeom(0.5, n = 10) # with rxgeom you have to explicitly state n
trxgeom(0.25, n = 10, ncores = 2) # You can parallelize the simulation using openMP
```
## This example uses ‘rxgeom’ directly in the model

```r
rx <- RxODE({
  a <- rxgeom(0.24)
})
et <- et(1, id = 1:2)
s <- rxSolve(rx, et)
```

### Description

Get the linear compartment model true function

### Usage

```r
rxGetLin(
  model,
  linCmtSens = c("linCmtA", "linCmtB", "linCmtC"),
  verbose = FALSE
)
```

### Arguments

- **model**: This is the ODE model specification. It can be:
  - a string containing the set of ordinary differential equations (ODE) and other expressions defining the changes in the dynamic system.
  - a file name where the ODE system equation is contained
  An ODE expression enclosed in `{}`
  (see also the filename argument). For details, see the sections “Details” and RxODE Syntax below.

- **linCmtSens**: The method to calculate the linCmt() solutions

- **verbose**: When TRUE be verbose with the linear compartmental model

### Value

model with linCmt() replaced with linCmtA()
**rxGetRxODE**  
*Get RxODE model from object*

**Description**  
Get RxODE model from object

**Usage**  
rxGetRxODE(obj)

**Arguments**  
obj RxODE family of objects

**Value**  
RxODE model

---

**rxHtml**  
*Format rxSolve and related objects as html.*

**Description**  
Format rxSolve and related objects as html.

**Usage**  
rxHtml(x, ...)

```r
class('rxSolve')

# S3 method for class 'rxSolve'
rxHtml(x, ...)
```

**Arguments**  
x RxODE object

... Extra arguments sent to kable

**Value**  
html code for rxSolve object

**Author(s)**  
Matthew L. Fidler
**rxIndLinState**

Set the preferred factoring by state

**Description**

Set the preferred factoring by state

**Usage**

```r
rxIndLinState(preferred = NULL)
```

**Arguments**

- `preferred`: A list of each state’s preferred factorization

**Value**

Nothing

**Author(s)**

Matthew Fidler

**rxIndLinStrategy**

This sets the inductive linearization strategy for matrix building

**Description**

When there is more than one state in a ODE that cannot be separated this specifies how it is incorporated into the matrix exponential.

**Usage**

```r
rxIndLinStrategy(strategy = c("curState", "split"))
```

**Arguments**

- `strategy`: The strategy for inductive linearization matrix building
  - `curState`: Prefer parameterizing in terms of the current state, followed by the first state observed in the term.
  - `split`: Split the parameterization between all states in the term by dividing each by the number of states in the term and then adding a matrix term for each state.
Value
Nothing

Author(s)
Matthew L. Fidler

---

**rxInv**

*Invert matrix using RcppArmadillo.*

---

**Description**

Invert matrix using RcppArmadillo.

**Usage**

```r
rxInv(matrix)
```

**Arguments**

- `matrix` matrix to be inverted.

**Value**

inverse or pseudo inverse of matrix.

---

**rxIsCurrent**

*Checks if the RxODE object was built with the current build*

---

**Description**

Checks if the RxODE object was built with the current build

**Usage**

```r
rxIsCurrent(obj)
```

**Arguments**

- `obj` RxODE family of objects

**Value**

boolean indicating if this was built with current RxODE
**rxLhs**

*Left handed Variables*

---

**Description**

This returns the model calculated variables

**Usage**

```r
rxLhs(obj)
```

**Arguments**

- `obj` RxODE family of objects

**Value**

A character vector listing the calculated parameters

**Author(s)**

Matthew L. Fidler

**See Also**

- `RxODE`

---

**rxLock**

*Lock/unlocking of RxODE dll file*

---

**Description**

Lock/unlocking of RxODE dll file

**Usage**

```r
rxLock(obj)

rxUnlock(obj)
```

**Arguments**

- `obj` A RxODE family of objects

**Value**

nothing; called for side effects
**rxNorm**

*Get the normalized model*

**Description**

This get the syntax preferred model for processing

**Usage**

```r
rxNorm(obj, condition = NULL, removeInis, removeJac, removeSens)
```

**Arguments**

- `obj`: RxODE family of objects
- `condition`: Character string of a logical condition to use for subsetting the normalized model. When missing, and a condition is not set via `rxCondition`, return the whole code with all the conditional settings intact. When a condition is set with `rxCondition`, use that condition.
- `removeInis`: A boolean indicating if parameter initialization will be removed from the model
- `removeJac`: A boolean indicating if the Jacobians will be removed.
- `removeSens`: A boolean indicating if the sensitivities will be removed.

**Value**

Normalized Normal syntax (no comments)

**Author(s)**

Matthew L. Fidler

---

**rxnorm**

*Simulate random normal variable from threefry/vandercorput generator*

**Description**

Simulate random normal variable from threefry/vandercorput generator

**Usage**

```r
rxnorm(mean = 0, sd = 1, n = 1L, ncores = 1L)
rxnormV(mean = 0, sd = 1, n = 1L, ncores = 1L)
```
Arguments

- **mean**: vector of means.
- **sd**: vector of standard deviations.
- **n**: number of observations
- **ncores**: Number of cores for the simulation

`rxnorm` simulates using the threefry sitmo generator; `rxnormV` uses the vandercorput generator

Value

normal random number deviates

Examples

```r
## Use threefry engine

rxnorm(n = 10) # with rxnorm you have to explicitly state n
rxnorm(n = 10, ncores = 2) # You can parallelize the simulation using openMP

rxnorm(2, 3) ## The first 2 arguments are the mean and standard deviation

## This example uses `rxnorm` directly in the model

rx <- RxODE(
  a <- rxnorm()
)

et <- et(1, id = 1:2)

s <- rxSolve(rx, et)

## Use vandercorput generator

rxnormV(n = 10) # with rxnorm you have to explicitly state n
rxnormV(n = 10, ncores = 2) # You can parallelize the simulation using openMP

rxnormV(2, 3) ## The first 2 arguments are the mean and standard deviation

## This example uses `rxnormV` directly in the model

rx <- RxODE(
  a <- rxnormV()
)

et <- et(1, id = 1:2)

s <- rxSolve(rx, et)
```
RxODE

Create an ODE-based model specification

Description
Create a dynamic ODE-based model object suitably for translation into fast C code

Usage

```r
RxODE(
  model,
  modName = basename(wd),
  wd = getwd(),
  filename = NULL,
  extraC = NULL,
  debug = FALSE,
  calcJac = NULL,
  calcSens = NULL,
  collapseModel = FALSE,
  package = NULL,
  ...
  linCmtSens = c("linCmtA", "linCmtB", "linCmtC"),
  indLin = FALSE,
  verbose = FALSE
)
```

Arguments

- **model**: This is the ODE model specification. It can be:
  - a string containing the set of ordinary differential equations (ODE) and other expressions defining the changes in the dynamic system.
  - a file name where the ODE system equation is contained

  An ODE expression enclosed in `\{\}`
  (see also the `filename` argument). For details, see the sections “Details” and `RxODE Syntax` below.

- **modName**: a string to be used as the model name. This string is used for naming various aspects of the computations, including generating C symbol names, dynamic libraries, etc. Therefore, it is necessary that `modName` consists of simple ASCII alphanumeric characters starting with a letter.

- **wd**: character string with a working directory where to create a subdirectory according to `modName`. When specified, a subdirectory named after the “`modName.d`” will be created and populated with a C file, a dynamic loading library, plus various other working files. If missing, the files are created (and removed) in the temporary directory, and the RxODE DLL for the model is created in the current directory named `rx_????_platform`, for example `rx_129f8f97fb94a87ca49ca8dafe691e1e_i386.dll`
filename A file name or connection object where the ODE-based model specification resides. Only one of model or filename may be specified.

extraC Extra C code to include in the model. This can be useful to specify functions in the model. These C functions should usually take double precision arguments, and return double precision values.

debug is a boolean indicating if the executable should be compiled with verbose debugging information turned on.

calcJac boolean indicating if RxODE will calculate the Jacobian according to the specified ODEs.

calcSens boolean indicating if RxODE will calculate the sensitivities according to the specified ODEs.

collapseModel boolean indicating if RxODE will remove all LHS variables when calculating sensitivities.

package Package name for pre-compiled binaries.

... ignored arguments.

linCmtSens The method to calculate the linCmt() solutions

indLin Calculate inductive linearization matrices and compile with inductive linearization support.

verbose When TRUE be verbose with the linear compartmental model

Details

The Rx in the name RxODE is meant to suggest the abbreviation Rx for a medical prescription, and thus to suggest the package emphasis on pharmacometrics modeling, including pharmacokinetics (PK), pharmacodynamics (PD), disease progression, drug-disease modeling, etc.

The ODE-based model specification may be coded inside a character string or in a text file, see Section RxODE Syntax below for coding details. An internal RxODE compilation manager object translates the ODE system into C, compiles it, and dynamically loads the object code into the current R session. The call to RxODE produces an object of class RxODE which consists of a list-like structure (environment) with various member functions (see Section Value below).

For evaluating RxODE models, two types of inputs may be provided: a required set of time points for querying the state of the ODE system and an optional set of doses (input amounts). These inputs are combined into a single event table object created with the function eventTable() or et().

An RxODE model specification consists of one or more statements optionally terminated by semicolons ; and optional comments (comments are delimited by # and an end-of-line).

A block of statements is a set of statements delimited by curly braces, { ... }.

Statements can be either assignments, conditional if/else if/else, while loops (can be exited by break), special statements, or printing statements (for debugging/testing)

Assignment statements can be:

- simple assignments, where the left hand is an identifier (i.e., variable)
- special time-derivative assignments, where the left hand specifies the change of the amount in the corresponding state variable (compartment) with respect to time e.g., d/dt(depot):
• special **initial-condition** assignments where the left hand specifies the compartment of the initial condition being specified, e.g. `depot(0) = 0`

• special model event changes including **bioavailability** (`f(depot)=1`), **lag time** (`alag(depot)=0`), **modeled rate** (`rate(depot)=2`) and **modeled duration** (`dur(depot)=2`). An example of these model features and the event specification for the modeled infusions the RxODE data specification is found in RxODE events vignette.

• special **change point syntax, or model times**. These model times are specified by `mtime(var)=time`

• special **Jacobian-derivative** assignments, where the left hand specifies the change in the compartment ode with respect to a variable. For example, if \( \frac{d}{dt}(y) = dy \), then a Jacobian for this compartment can be specified as `df(y)/dy(dy) = 1`. There may be some advantage to obtaining the solution or specifying the Jacobian for very stiff ODE systems. However, for the few stiff systems we tried with LSODA, this actually slightly slowed down the solving.

Note that assignment can be done by `=`, `<-` or `~`.

When assigning with the `~` operator, the **simple assignments** and **time-derivative** assignments will not be output.

Special statements can be:

• **Compartment declaration statements**, which can change the default dosing compartment and the assumed compartment number(s) as well as add extra compartment names at the end (useful for multiple-endpoint nlmixr models); These are specified by `cmt(compartmentName)`

• **Parameter declaration statements**, which can make sure the input parameters are in a certain order instead of ordering the parameters by the order they are parsed. This is useful for keeping the parameter order the same when using 2 different ODE models. These are specified by `param(par1,par2,...)`

An example model is shown below:

```r
# simple assignment
c2 = centr/V2;

# time-derivative assignment
d/dt(centr) = F*KA*depot - CL*C2 - Q*C2 + Q*C3;
```

Expressions in assignment and if statements can be numeric or logical, however, no character nor integer expressions are currently supported.

Numeric expressions can include the following numeric operators `+,-,*,/` and those mathematical functions defined in the C or the R math libraries (e.g., `fabs`, `exp`, `log`, `sin`, `abs`).

You may also access the R’s functions in the R math libraries, like `lgammafn` for the log gamma function.

The RxODE syntax is case-sensitive, i.e., `ABC` is different than `abc`, `Abc`, `ABc`, etc.

**Identifiers:**

Like R, Identifiers (variable names) may consist of one or more alphanumeric, underscore `_` or period `.`, characters, but the first character cannot be a digit or underscore `_`.

Identifiers in a model specification can refer to:

• State variables in the dynamic system (e.g., compartments in a pharmacokinetics model).
• Implied input variable, \( t \) (time), \( t_{\text{last}} \) (last time point), and \( \text{podo} \) (oral dose, in the undocumented case of absorption transit models).
• Special constants like \( \pi \) or R’s predefined constants.
• Model parameters (e.g., \( k_a \) rate of absorption, \( CL \) clearance, etc.)
• Others, as created by assignments as part of the model specification; these are referred as \textit{LHS} (left-hand side) variable.

Currently, the \texttt{RxODE} modeling language only recognizes system state variables and “parameters”. thus, any values that need to be passed from R to the ODE model (e.g., \textit{age}) should be either passed in the \texttt{params} argument of the integrator function \texttt{rxSolve()} or be in the supplied event data-set.

There are certain variable names that are in the \texttt{RxODE} event tables. To avoid confusion, the following event table-related items cannot be assigned, or used as a state but can be accessed in the \texttt{RxODE} code:

- \texttt{cmt}
- \texttt{dvid}
- \texttt{addl}
- \texttt{ss}
- \texttt{rate}
- \texttt{id}

However the following variables are cannot be used in a model specification:

- \texttt{evid}
- \texttt{ii}

Sometimes \texttt{RxODE} generates variables that are fed back to \texttt{RxODE}. Similarly, \texttt{nlmixr} generates some variables that are used in \texttt{nlmizx} estimation and simulation. These variables start with the either the \texttt{rx} or \texttt{nlmixr} prefixes. To avoid any problems, it is suggested to not use these variables starting with either the \texttt{rx} or \texttt{nlmixr} prefixes.

**Logical Operators:**

Logical operators support the standard R operators \( ==, !=, >=, <=, > \) and \( < \). Like R these can be in \texttt{if()} or \texttt{while()} statements, \texttt{ifelse()} expressions. Additionally they can be in a standard assignment. For instance, the following is valid:

\[
\text{cov1} = \text{covm*(sexf} == \text{“female”) + covm*(sexf} != \text{“female”)}
\]

Notice that you can also use character expressions in comparisons. This convenience comes at a cost since character comparisons are slower than numeric expressions. Unlike R, \texttt{as.numeric} or \texttt{as.integer} for these logical statements is not only not needed, but will cause an syntax error if you try to use the function.

**Value**

An object (environment) of class \texttt{RxODE} (see Chambers and Temple Lang (2001)) consisting of the following list of strings and functions:

- \texttt{‘model’} a character string holding the source model specification.
- \texttt{‘get.modelVars’} a function that returns a list with 3 character vectors, \texttt{‘params’}, \texttt{‘state’}, and \texttt{‘lhs’} of variable names used in the model
specification. These will be output when the model is computed (i.e., the ODE solved by integration).

* `solve` (this function solves (integrates) the ODE. This is done by passing the code to `rxSolve()`.
This is as if you called `rxSolve(RxODEobject, ...)`, but returns a matrix instead of a rxSolve object.

`params`: a numeric named vector with values for every parameter in the ODE system; the names must correspond to the parameter identifiers used in the ODE specification;

`events`: an `eventTable` object describing the input (e.g., doses) to the dynamic system and observation sampling time points (see `eventTable()`);

`inits`: a vector of initial values of the state variables (e.g., amounts in each compartment), and the order in this vector must be the same as the state variables (e.g., PK/PD compartments);

`stiff`: a logical (`TRUE` by default) indicating whether the ODE system is stiff or not.

For stiff ODE systems (`stiff = TRUE`), `RxODE` uses the LSODA (Livermore Solver for Ordinary Differential Equations) Fortran package, which implements an automatic method switching for stiff and non-stiff problems along the integration interval, authored by Hindmarsh and Petzold (2003).

For non-stiff systems (`stiff = FALSE`), `RxODE` uses `DOP853`, an explicit Runge-Kutta method of order 8(5, 3) of Dormand and Prince as implemented in C by Hairer and Wanner (1993).

`trans_abs`: a logical (`FALSE` by default) indicating whether to fit a transit absorption term (TODO: need further documentation and example);

`atol`: a numeric absolute tolerance (1e-08 by default);

`rtol`: a numeric relative tolerance (1e-06 by default).

The output of `solve` is a matrix with as many rows as there are sampled time points and as many columns as system variables (as defined by the ODEs and additional assignments in the RxODE model code).

* `isValid` a function that (naively) checks for model validity, namely that the C object code reflects the latest model
specification.
* `version` a string with the version of the `RxODE` object (not the package).
* `dynLoad` a function with one `force = FALSE` argument that dynamically loads the object code if needed.
* `dynUnload` a function with no argument that unloads the model object code.
* `delete` removes all created model files, including C and DLL files. The model object is no longer valid and should be removed, e.g., `rm(m1)`.
* `run` deprecated, use `solve`.
* `get.index` deprecated.
* `getObj` internal (not user callable) function.

**Author(s)**
Melissa Hallow, Wenping Wang and Matthew Fidler

**References**

**See Also**
eventTable(), et(), add.sampling(), add.dosing()

**Examples**

```r
# Step 1 - Create a model specification
ode <- 
  # A 4-compartment model, 3 PK and a PD (effect) compartment
  # (notice state variable names 'depot', 'centr', 'peri', 'eff')

  C2 = centr/V2;
  C3 = peri/V3;
  d/dt(depot) = -KA*depot;
  d/dt(centr) = KA*depot - CL*C2 - Q*C2 + Q*C3;
  d/dt(peri) = Q*C2 - Q*C3;
  d/dt(eff) = Kin - Kout*(1-C2/(EC50+C2))*eff;
```

m1 <- RxODE(model = ode)
print(m1)

# Step 2 - Create the model input as an EventTable, # including dosing and observation (sampling) events
# QD (once daily) dosing for 5 days.
qd <- eventTable(amount.units = "ug", time.units = "hours")
qd$add.dosing(dose = 10000, nbr.doses = 5, dosing.interval = 24)

# Sample the system hourly during the first day, every 8 hours # then after
qd$add.sampling(0:24)
qd$add.sampling(seq(from = 24 + 8, to = 5 * 24, by = 8))

# Step 3 - set starting parameter estimates and initial # values of the state
theta <-
  c(
    KA = .291, CL = 18.6,
    V2 = 40.2, Q = 10.5, V3 = 297.0,
    Kin = 1.0, Kout = 1.0, EC50 = 200.0
  )

# init state variable
inits <- c(0, 0, 0, 1)
# Step 4 - Fit the model to the data
qd.cp <- m1$solve(theta, events = qd, inits)

head(qd.cp)

# This returns a matrix. Note that you can also # solve using name initial values. For example:
inits <- c(eff = 1)
qd.cp <- solve(m1, theta, events = qd, inits)
print(qd.cp)
plot(qd.cp)
**rxParams**

**Description**

This optimizes RxODE code for computer evaluation by only calculating redundant expressions once.

**Usage**

```r
rxOptExpr(x, msg = "model")
```

**Arguments**

- **x**: RxODE model that can be accessed by rxNorm
- **msg**: This is the name of type of object that RxODE is optimizing that will in the message when optimizing. For example "model" will produce the following message while optimizing the model:
  
  finding duplicate expressions in model...

**Value**

Optimized RxODE model text. The order and type lhs and state variables is maintained while the evaluation is sped up. While parameters names are maintained, their order may be modified.

**Author(s)**

Matthew L. Fidler

---

**rxParams**

*Parameters specified by the model*

**Description**

This returns the model’s parameters that are required to solve the ODE system, and can be used to pipe parameters into an RxODE solve.

**Usage**

```r
rxParams(obj, ...)
```

```r
## S3 method for class 'RxODE'
rxParams(
    obj, 
    constants = TRUE,
    ..., 
    params = NULL, 
    inits = NULL, 
    iCov = NULL, 
    keep = NULL, 
    thetaMat = NULL, 
)```
omega = NULL,
dfSub = NULL,
sigma = NULL,
dfObs = NULL,
nSub = NULL,
nStud = NULL)

## S3 method for class 'rxSolve'
rxParams(
  obj,
  constants = TRUE,
  ...,
  params = NULL,
  inits = NULL,
iCov = NULL,
  keep = NULL,
  thetaMat = NULL,
  omega = NULL,
  dfSub = NULL,
  sigma = NULL,
  dfObs = NULL,
nSub = NULL,
nStud = NULL
)

## S3 method for class 'rxEt'
rxParams(
  obj,
  ...
  params = NULL,
  inits = NULL,
iCov = NULL,
  keep = NULL,
  thetaMat = NULL,
  omega = NULL,
  dfSub = NULL,
  sigma = NULL,
  dfObs = NULL,
nSub = NULL,
nStud = NULL
)

rxParam(obj, ...)

Arguments

obj  RxODE family of objects
... Other arguments including scaling factors for each compartment. This includes $S# = \text{numeric}$ will scale a compartment # by a dividing the compartment amount by the scale factor, like NONMEM.

`constants` is a boolean indicating if constants should be included in the list of parameters. Currently RxODE parses constants into variables in case you wish to change them without recompiling the RxODE model.

`params` is a numeric named vector with values for every parameter in the ODE system; the names must correspond to the parameter identifiers used in the ODE specification;

`inits` is a vector of initial values of the state variables (e.g., amounts in each compartment), and the order in this vector must be the same as the state variables (e.g., PK/PD compartments);

`iCov` is a data frame of individual non-time varying covariates to combine with the events dataset by merge.

`keep` is Columns to keep from either the input dataset or the iCov dataset. With the iCov dataset, the column is kept once per line. For the input dataset, if any records are added to the data LOCF (Last Observation Carried forward) imputation is performed.

`thetaMat` is a Named theta matrix.

`omega` is a Estimate of Covariance matrix. When omega is a list, assume it is a block matrix and convert it to a full matrix for simulations.

`dfSub` is Degrees of freedom to sample the between subject variability matrix from the inverse Wishart distribution (scaled) or scaled inverse chi squared distribution.

`sigma` is a Named sigma covariance or Cholesky decomposition of a covariance matrix. The names of the columns indicate parameters that are simulated. These are simulated for every observation in the solved system.

`dfObs` is Degrees of freedom to sample the unexplained variability matrix from the inverse Wishart distribution (scaled) or scaled inverse chi squared distribution.

`nSub` is Number between subject variabilities (ETAs) simulated for every realization of the parameters.

`nStud` is Number virtual studies to characterize uncertainty in estimated parameters.

Value

When extracting the parameters from an RxODE model, a character vector listing the parameters in the model.

Author(s)

Matthew L. Fidler
rxPkg

Creates a package from compiled RxODE models

Description

Creates a package from compiled RxODE models

Usage

```r
rxPkg(
  ..., package, wd = getwd(),
  action = c("install", "build", "binary", "create"),
  license = c("gpl3", "lgpl", "mit", "agpl3"),
  name = "Firstname Lastname",
  fields = list()
)
```

Arguments

- `...`: Models to build a package from
- `package`: String of the package name to create
- `wd`: character string with a working directory where to create a subdirectory according to `modName`. When specified, a subdirectory named after the "modName.d" will be created and populated with a C file, a dynamic loading library, plus various other working files. If missing, the files are created (and removed) in the temporary directory, and the RxODE DLL for the model is created in the current directory named `rx_????_platform`, for example `rx_129f8f97fb94a87ca49ca8dafe691e1e_i386.dll`
- `action`: Type of action to take after package is created
- `license`: is the type of license for the package.
- `name`: Full name of author
- `fields`: A named list of fields to add to DESCRIPTION, potentially overriding default values. See `use_description()` for how you can set personalized defaults using package options

Value

This function returns nothing and is used for its side effects

Author(s)

Matthew Fidler
Simulate random Poisson variable from threefry generator

Description

Care should be taken with this method not to encounter the birthday problem, described [https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/](https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/). Since the sitmo threefry, this currently generates one random deviate from the uniform distribution to seed the engine threefry and then run the code.

Usage

```
rxpois(lambda, n = 1L, ncores = 1L)
```

Arguments

- **lambda**: vector of (non-negative) means.
- **n**: number of random values to return.
- **ncores**: Number of cores for the simulation

**Details**

Therefore, a simple call to the random number generated followed by a second call to random number generated may have identical seeds. As the number of random number generator calls are increased the probability that the birthday problem will increase.

The key to avoid this problem is to either run all simulations in the RxODE environment once (therefore one seed or series of seeds for the whole simulation), pre-generate all random variables used for the simulation, or seed the RxODE engine with `rxSetSeed()`.

Also care should be made that the computer you will be running on can run the same number of cores as you are running so they can reproduce your results.

Value

poisson random number deviates

Examples

```
## Use threefry engine

rxpois(lambda = 3, n = 10) # with rxpois you have to explicitly state n
rxpois(lambda = 3, n = 10, ncores = 2) # You can parallelize the simulation using openMP
rxpois(4) ## The first arguments are the lambda parameter
```
## This example uses `rxpois` directly in the model

```r
rx <- RxODE(
  a <- rxpois(3)
)

et <- et(1, id = 1:2)

s <- rxSolve(rx, et)
```

---

### rxPp

Simulate a from a Poisson process

#### Description

Simulate a from a Poisson process

#### Usage

```r
rxPp(
  n,
  lambda,
  gamma = 1,
  prob = NULL,
  t0 = 0,
  tmax = Inf,
  randomOrder = FALSE
)
```

#### Arguments

- **n**
  Number of time points to simulate in the Poisson process

- **lambda**
  Rate of Poisson process

- **gamma**
  Asymmetry rate of Poisson process. When gamma=1.0, this simulates a homogenous Poisson process. When gamma<1.0, the Poisson process has more events early, when gamma > 1.0, the Poisson process has more events late in the process.
  When gamma is non-zero, the tmax should not be infinite but indicate the end of the Poisson process to be simulated. In most pharamcometric cases, this will be the end of the study. Internally this uses a rate of:
  \[ l(t) = \lambda \gamma \left(\frac{t}{t_{\max}}\right)^{\gamma-1} \]

- **prob**
  When specified, this is a probability function with one argument, time, that gives the probability that a Poisson time \( t \) is accepted as a rejection time.

- **t0**
  the starting time of the Poisson process
tmax the maximum time of the Poisson process
randomOrder when TRUE randomize the order of the Poisson events. By default (FALSE) it returns the Poisson process is in order of how the events occurred.

Value
This returns a vector of the Poisson process times; If the dropout is >= tmax, then all the rest of the times are = tmax to indicate the dropout is equal to or after tmax.

Author(s)
Matthew Fidler

Examples

```r
## Sample homogenous Poisson process of rate 1/10
rxPp(10, 1 / 10)

## Sample inhomogenous Poisson rate of 1/10
rxPp(10, 1 / 10, gamma = 2, tmax = 100)

## Typically the Poisson process times are in a sequential order, 
## using randomOrder gives the Poisson process in random order
rxPp(10, 1 / 10, gamma = 2, tmax = 10, randomOrder = TRUE)

## This uses an arbitrary function to sample a non-homogenous Poisson process
rxPp(10, 1 / 10, prob = function(x) {
  1 / x
})
```

Description

rxProgress sets up the progress bar

Usage

```r
rxProgress(num, core = 0L)
rxTick()
rxProgressStop(clear = TRUE)
rxProgressAbort(error = "Aborted calculation")
```
Arguments

num        Tot number of operations to track
core       Number of cores to show. If below 1, don’t show number of cores
clear      Boolean telling if you should clear the progress bar after completion (as if it
            wasn’t displayed). By default this is TRUE
error      With rxProgressAbort this is the error that is displayed

Details

rxTick is a progress bar tick
rxProgressStop stop progress bar

rxProgressAbort shows an abort if rxProgressStop wasn’t called.

Value

All return NULL invisibly.

Author(s)

Matthew L. Fidler

Examples

f <- function() {
  on.exit({
    rxProgressAbort()
  })
  rxProgress(100)
  for (i in 1:100) {
    rxTick()
    Sys.sleep(1 / 100)
  }
  rxProgressStop()
}

f()

---

rxRandNV Create a random "normal" matrix using vandercorput generator

Description

Create a random "normal" matrix using vandercorput generator
**rxRandNV**

**Usage**

```
rxRandNV(nrow = 1, ncol = 1)
```

**Arguments**

- `nrow` Number of rows
- `ncol` Number of Columns

**Value**

Matrix of random numbers

**Author(s)**

Matthew Fidler

**Examples**

```
rxRandNV(1, 1)
rxRandNV(3, 2)
```

---

**rxRateDur**

*Creates a rxRateDur object*

**Description**

This is primarily to display information about rate

**Usage**

```
rxRateDur(x)
```

`````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````````
Arguments

\[
\begin{align*}
x & \quad \text{rxRateDur data} \\
\ldots & \quad \text{Other parameters}
\end{align*}
\]

Value

rxRateDur object

---

rxReservedKeywords  
\textit{A list and description of Rode supported reserved keywords}

Description

A list and description of Rode supported reserved keywords

Usage

rxReservedKeywords

Format

A data frame with 3 columns and 98 or more rows

\begin{itemize}
  \item \textbf{Reserved Name}  Reserved Keyword Name
  \item \textbf{Meaning}  Reserved Keyword Meaning
  \item \textbf{Alias}  Keyword Alias
\end{itemize}

---

rxRmvn  
\textit{Simulate from a (truncated) multivariate normal}

Description

This is simulated with the fast, thread-safe threefry simulator and can use multiple cores to generate the random deviates.
Usage

```r
rxRmvn(
  n,
  mu = NULL,
  sigma,
  lower = -Inf,
  upper = Inf,
  ncores = 1,
  isChol = FALSE,
  keepNames = TRUE,
  a = 0.4,
  tol = 2.05,
  nlTol = 1e-10,
  nlMaxiter = 100L
)
```

Arguments

- **n** Number of random row vectors to be simulated OR the matrix to use for simulation (faster).
- **mu** mean vector
- **sigma** Covariance matrix for multivariate normal or a list of covariance matrices. If a list of covariance matrix, each matrix will simulate n matrices and combine them to a full matrix
- **lower** is a vector of the lower bound for the truncated multivariate norm
- **upper** is a vector of the upper bound for the truncated multivariate norm
- **ncores** Number of cores used in the simulation
- **isChol** A boolean indicating if sigma is a cholesky decomposition of the covariance matrix.
- **keepNames** Keep the names from either the mean or covariance matrix.
- **a** threshold for switching between methods; They can be tuned for maximum speed; There are three cases that are considered:
  - case 1: a < l < u
  - case 2: l < u < -a
  - case 3: otherwise
  where l=lower and u = upper
- **tol** When case 3 is used from the above possibilities, the tol value controls the acceptance rejection and inverse-transformation; When abs(u-l)>tol, uses accept-reject from randn
- **nlTol** Tolerance for newton line-search
- **nlMaxiter** Maximum iterations for newton line-search
Value

If \( n = \text{integer} \) (default) the output is an \((n \times d)\) matrix where the \(i\)-th row is the \(i\)-th simulated vector.

If \( \text{is.matrix}(n) \) then the random vector are store in \( n \), which is provided by the user, and the function returns NULL invisibly.

Author(s)

Matthew Fidler, Zdravko Botev and some from Matteo Fasiolo

References


The thread safe multivariate normal was inspired from the \texttt{mvnfast} package by Matteo Fasiolo

https://CRAN.R-project.org/package=mvnfast

The concept of the truncated multivariate normal was taken from Zdravko Botev (2017) doi: 10.1111/rssb.12162 and Botev and L’Ecuyer (2015) doi: 10.1109/WSC.2015.7408180 and converted to thread safe simulation;

Examples

## From \texttt{mvnfast}

## Unlike \texttt{mvnfast}, uses \texttt{threelfry} simulation

```r
d <- 5
mu <- 1:d

# Creating covariance matrix
tmp <- matrix(rnorm(d^2), d, d)
mcov <- tcrossprod(tmp, tmp)

set.seed(414)
rxRmvn(4, 1:d, mcov)
set.seed(414)
rxRmvn(4, 1:d, mcov)
set.seed(414)
rxRmvn(4, 1:d, mcov, ncores = 2) # r.v. generated on the second core are different

# Here we create the matrix that will hold the simulated
# random variables upfront.
A <- matrix(NA, 4, d)
class(A) <- "numeric" # This is important. We need the elements of A to be of class "numeric".

set.seed(414)
rxRmvn(A, 1:d, mcov, ncores = 2) # This returns NULL ...
```
## You can also simulate from a truncated normal:

```r
rxRmvn(10, 1:d, mcov, lower = 1:d - 1, upper = 1:d + 1)
```

# You can also simulate from different matrices (if they match dimensions) by using a list of matrices.

```r
matL <- lapply(1:4, function(...) {
  tmp <- matrix(rnorm(d^2), d, d)
  tcrossprod(tmp, tmp)
})

rxRmvn(4, setNames(1:d, paste0("a", 1:d)), matL)
```

---

**rxS**

*Load a model into a symengine environment*

### Description

Load a model into a symengine environment

### Usage

```r
rxS(x, doConst = TRUE, promoteLinSens = FALSE)
```

### Arguments

- **x**
  - RxODE object
- **doConst**
  - Load constants into the environment as well.
- **promoteLinSens**
  - Promote solved linear compartment systems to sensitivity-based solutions.

### Value

RxODE/symengine environment

### Author(s)

Matthew Fidler
### rxSetIni0

**Description**

Set Initial conditions to time zero instead of the first observed/dosed time

**Usage**

```r
rxSetIni0(ini0 = TRUE)
```

**Arguments**

- `ini0` When TRUE (default), set initial conditions to time zero. Otherwise the initial conditions are the first observed time.

**Value**

the boolean ini0, though this is called for its side effects

### rxSetProd

**Description**

Defunct setting of product

**Usage**

```r
rxSetProd(type = c("long double", "double", "logify"))
```

**Arguments**

- `type` used to be type of product

**Value**

nothing
**rxSetProgressBar**

*Set timing for progress bar*

**Description**

Set timing for progress bar

**Usage**

```r
rxSetProgressBar(seconds = 1)
```

**Arguments**

- **seconds**
  
  This sets the number of seconds that need to elapse before drawing the next segment of the progress bar. When this is zero or below this turns off the progress bar.

**Value**

nothing, used for side effects

**Author(s)**

Matthew Fidler

---

**rxSetSeed**

*Set the parallel seed for RxODE random number generation*

**Description**

This sets the seed for the RxODE parallel random number generation. If set, then whenever a seed is set for the threefry or vandercorput simulation engine, it will use this seed, increment for the number of seeds and continue with the sequence the next time the random number generator is called.

**Usage**

```r
rxSetSeed(seed)
```

**Arguments**

- **seed**

  An integer that represents the RxODE parallel and internal random number generator seed. When positive, use this seed for random number generation and increment and reseed any parallel or new engines that are being called. When negative, turn off the RxODE seed and generate a seed from the R’s uniform random number generator. Best practice is to set this seed.
Details

In contrast, when this is not called, the time that the vandercorput or threfry simulation engines are seeded it comes from a uniform random number generated from the standard R random seed. This may cause a duplicate seed based on the R seed state. This means that there could be correlations between simulations that do not exist. This will avoid the birthday problem picking exactly the same seed using the seed state of the R random number generator. The more times the seed is called, the more likely this becomes.

Value

Nothing, called for its side effects

Author(s)

Matthew Fidler

References


Examples

rxSetSeed(42)

# seed with generator 42
rxnorm()

# Use R's random number generator
rnorm(1)

rxSetSeed(42)

# reproduces the same number
rxnorm()

# But R's random number is not the same
rnorm(1)

# If we reset this to use the R's seed
# (internally RxODE uses a uniform random number to span seeds)
# This can lead to duplicate sequences and seeds
rxSetSeed(-1)

# Now set seed works for both.

# This is not recommended, but illustrates the different types of
# seeds that can be generated.
**rxSetSum**

set.seed(42)
rxnorm()
rnorm(1)
set.seed(42)
rxnorm()
rnorm(1)

---

**rxSetSum**  
*Defunct setting of sum*

**Description**
Defunct setting of sum

**Usage**

```r
rxSetSum(type = c("pairwise", "fsum", "kahan", "neumaier", "c"))
```

**Arguments**

- **type**  
  used to be type of product

**Value**
nothing

---

**rxShiny**  
*Use Shiny to help develop an RxODE model*

**Description**
Use Shiny to help develop an RxODE model

**Usage**

```r
rxShiny(  
  object,  
  params = NULL,  
  events = NULL,  
  inits = NULL,  
  ...,
  data = data.frame()
)```
## S3 method for class 'rxSolve'
rxShiny(
  object,
  params = NULL,
  events = NULL,
  inits = NULL,
  ...,
  data = data.frame()
)

## Default S3 method:
rxShiny(
  object = NULL,
  params = NULL,
  events = NULL,
  inits = NULL,
  ...,
  data = data.frame()
)

Arguments

object A RxODE family of objects. If not supplied a 2-compartment indirect effect model is used. If it is supplied, use the model associated with the RxODE object for the model exploration.

params Initial parameters for model

events Event information (currently ignored)

inits Initial estimates for model

... Other arguments passed to rxShiny. Currently doesn’t do anything.

data Any data that you would like to plot. If the data has a time variable as well as a compartment or calculated variable that matches the RxODE model, the data will be added to the plot of a specific compartment or calculated variable.

Value

Nothing: Starts a shiny server

Author(s)

Zufar Mulyukov and Matthew L. Fidler
Simulate Parameters from a Theta/Omega specification

Usage

```r
rxSimThetaOmega(
  params = NULL,
  omega = NULL,
  omegaDf = NULL,
  omegaLower = as.numeric(c(R_NegInf)),
  omegaUpper = as.numeric(c(R_PosInf)),
  omegaIsChol = FALSE,
  omegaSeparation = "auto",
  omegaXform = 1L,
  nSub = 1L,
  thetaMat = NULL,
  thetaLower = as.numeric(c(R_NegInf)),
  thetaUpper = as.numeric(c(R_PosInf)),
  thetaDf = NULL,
  thetaIsChol = FALSE,
  nStud = 1L,
  sigma = NULL,
  sigmaLower = as.numeric(c(R_NegInf)),
  sigmaUpper = as.numeric(c(R_PosInf)),
  sigmaDf = NULL,
  sigmaIsChol = FALSE,
  sigmaSeparation = "auto",
  sigmaXform = 1L,
  nCoresRV = 1L,
  nObs = 1L,
  dfSub = 0,
  dfObs = 0,
  simSubjects = TRUE
)
```

Arguments

- **params**: Named Vector of RxODE model parameters
- **omega**: Estimate of Covariance matrix. When omega is a list, assume it is a block matrix and convert it to a full matrix for simulations.
- **omegaDf**: The degrees of freedom of a t-distribution for simulation. By default this is NULL which is equivalent to Inf degrees, or to simulate from a normal distribution instead of a t-distribution.
omegaLower  Lower bounds for simulated ETAs (by default -Inf)
omegaUpper  Upper bounds for simulated ETAs (by default Inf)
omegaIsChol  Indicates if the omega supplied is a Cholesky decomposed matrix instead of the traditional symmetric matrix.

omegaSeparation
Omega separation strategy
Tells the type of separation strategy when simulating covariance with parameter uncertainty with standard deviations modeled in the thetaMat matrix.

- "lkj" simulates the correlation matrix from the rLKJ1 matrix with the distribution parameter eta equal to the degrees of freedom nu by (nu-1)/2
- "separation" simulates from the identity inverse Wishart covariance matrix with nu degrees of freedom. This is then converted to a covariance matrix and augmented with the modeled standard deviations. While computationally more complex than the "lkj" prior, it performs better when the covariance matrix size is greater or equal to 10
- "auto" chooses "lkj" when the dimension of the matrix is less than 10 and "separation" when greater than equal to 10.

omegaXform
When taking omega values from the thetaMat simulations (using the separation strategy for covariance simulation), how should the thetaMat values be turned into standard deviation values:

- identity This is when standard deviation values are directly modeled by the params and thetaMat matrix
- variance This is when the params and thetaMat simulates the variance that are directly modeled by the thetaMat matrix
- log This is when the params and thetaMat simulates log(sd)
- nlmixrSqrt This is when the params and thetaMat simulates the inverse cholesky decomposed matrix with the x^2 modeled along the diagonal. This only works with a diagonal matrix.
- nlmixrLog This is when the params and thetaMat simulates the inverse cholesky decomposed matrix with the exp(x^2) along the diagonal. This only works with a diagonal matrix.
- nlmixrIdentity This is when the params and thetaMat simulates the inverse cholesky decomposed matrix. This only works with a diagonal matrix.

nSub  Number between subject variabilities (ETAs) simulated for every realization of the parameters.
thetaMat  Named theta matrix.
thetaLower  Lower bounds for simulated population parameter variability (by default -Inf)
thetaUpper  Upper bounds for simulated population unexplained variability (by default Inf)
thetaDf  The degrees of freedom of a t-distribution for simulation. By default this is NULL which is equivalent to Inf degrees, or to simulate from a normal distribution instead of a t-distribution.
thetaIsChol  Indicates if the theta supplied is a Cholesky decomposed matrix instead of the traditional symmetric matrix.
nStud  Number of virtual studies to characterize uncertainty in estimated parameters.
sigma  Named sigma covariance or Cholesky decomposition of a covariance matrix. The names of the columns indicate parameters that are simulated. These are simulated for every observation in the solved system.
sigmaLower  Lower bounds for simulated unexplained variability (by default -Inf)
sigmaUpper  Upper bounds for simulated unexplained variability (by default Inf)
sigmaDf  Degrees of freedom of the sigma t-distribution. By default it is equivalent to Inf, or a normal distribution.
sigmaIsChol  Boolean indicating if the sigma is in the Cholesky decomposition instead of a symmetric covariance

**sigmaSeparation**
   separation strategy for sigma;
   Tells the type of separation strategy when simulating covariance with parameter uncertainty with standard deviations modeled in the thetaMat matrix.
   • "lkj" simulates the correlation matrix from the rLKJ1 matrix with the distribution parameter eta equal to the degrees of freedom nu by (nu-1)/2
   • "separation" simulates from the identity inverse Wishart covariance matrix with nu degrees of freedom. This is then converted to a covariance matrix and augmented with the modeled standard deviations. While computationally more complex than the "lkj" prior, it performs better when the covariance matrix size is greater or equal to 10
   • "auto" chooses "lkj" when the dimension of the matrix is less than 10 and "separation" when greater than equal to 10.

**sigmaXform**  When taking sigma values from the thetaMat simulations (using the separation strategy for covariance simulation), how should the thetaMat values be turned into standard deviation values:
   • identity This is when standard deviation values are directly modeled by the params and thetaMat matrix
   • variance This is when the params and thetaMat simulates the variance that are directly modeled by the thetaMat matrix
   • log This is when the params and thetaMat simulates log(sd)
   • nlmixrSqrt This is when the params and thetaMat simulates the inverse cholesky decomposed matrix with the x^2 modeled along the diagonal. This only works with a diagonal matrix.
   • nlmixrLog  This is when the params and thetaMat simulates the inverse cholesky decomposed matrix with the exp(x^2) along the diagonal. This only works with a diagonal matrix.
   • nlmixrIdentity  This is when the params and thetaMat simulates the inverse cholesky decomposed matrix. This only works with a diagonal matrix.

nCoresRV  Number of cores used for the simulation of the sigma variables. By default this is 1. To reproduce the results you need to run on the same platform with the same number of cores. This is the reason this is set to be one, regardless of what the number of cores are used in threaded ODE solving.
rxSolve

nObs

Number of observations to simulate (with sigma matrix)

dfSub

Degrees of freedom to sample the between subject variability matrix from the inverse Wishart distribution (scaled) or scaled inverse chi squared distribution.

dfObs

Degrees of freedom to sample the unexplained variability matrix from the inverse Wishart distribution (scaled) or scaled inverse chi squared distribution.

simSubjects

boolean indicated RxODE should simulate subjects in studies (TRUE, default) or studies (FALSE)

Value

a data frame with the simulated subjects

Author(s)

Matthew L.Fidler

rxSolve

Solving & Simulation of a ODE/solved system (and solving options)
equation

Description

This uses RxODE family of objects, file, or model specification to solve a ODE system. There are many options for a solved RxODE model, the first are the required object, and events with the some-times optional params and inits.

Usage

rxSolve(
  object,
  params = NULL,
  events = NULL,
  inits = NULL,
  scale = NULL,
  method = c("liblsoda", "lsoda", "dop853", "indLin"),
  transitAbs = NULL,
  atol = 1e-08,
  rtol = 1e-06,
  maxsteps = 70000L,
  hmin = 0,
  hmax = NA_real_,
  hmaxSd = 0,
  hini = 0,
  maxordn = 12L,
  maxords = 5L,
  ...,
cores,
covsInterpolation = c("locf", "linear", "nocb", "midpoint"),
addCov = FALSE,
matrix = FALSE,
sigma = NULL,
sigmaDf = NULL,
sigmaLower = -Inf,
sigmaUpper = Inf,
nCoresRV = 1L,
sigmaIsChol = FALSE,
sigmaSeparation = c("auto", "lkj", "separation"),
sigmaXform = c("identity", "variance", "log", "nlmixrSqrt", "nlmixrLog",
              "nlmixrIdentity"),
nDisplayProgress = 10000L,
amountUnits = NA_character_,
timeUnits = "hours",
stiff,
theta = NULL,
thetaLower = -Inf,
thetaUpper = Inf,
eta = NULL,
addDosing = FALSE,
stateTrim = Inf,
updateObject = FALSE,
omega = NULL,
omegaDf = NULL,
omegaIsChol = FALSE,
omegaSeparation = c("auto", "lkj", "separation"),
omegaXform = c("variance", "identity", "log", "nlmixrSqrt", "nlmixrLog",
               "nlmixrIdentity"),
omegaLower = -Inf,
omegaUpper = Inf,
nSub = 1L,
thetaMat = NULL,
thetaDf = NULL,
thetaIsChol = FALSE,
nStud = 1L,
dfSub = 0,
dfObs = 0,
returnType = c("rxSolve", "matrix", "data.frame", "data.frame.TBS", "data.table",
               "tbl", "tibble"),
seed = NULL,
nsim = NULL,
minSS = 10L,
maxSS = 1000L,
inSSstep = 12,
strictSS = TRUE,
istateReset = TRUE,
subsetNonmem = TRUE,
maxAtolRtolFactor = 0.1,
from = NULL,
to = NULL,
by = NULL,
length.out = NULL,
iCov = NULL,
keep = NULL,
indLinPhiTol = 1e-07,
indLinPhiM = 0L,
indLinMatExpType = c("expokit", "Al-Mohy", "arma"),
indLinMatExpOrder = 6L,
drop = NULL,
idFactor = TRUE,
mxhnil = 0,
hmxii = 0,
warnIdSort = TRUE,
warnDrop = TRUE,
ssAtol = 1e-08,
ssRtol = 1e-06,
safeZero = TRUE,
sumType = c("pairwise", "fsum", "kahan", "neumaier", "c"),
prodType = c("long double", "double", "logify"),
sensType = c("advan", "autodiff", "forward", "central"),
linDiff = c(tlag = 1.5e-05, f = 1.5e-05, rate = 1.5e-05, dur = 1.5e-05, tlag2 = 1.5e-05, f2 = 1.5e-05, rate2 = 1.5e-05, dur2 = 1.5e-05),
linDiffCentral = c(tlag = TRUE, f = TRUE, rate = TRUE, dur = TRUE, tlag2 = TRUE, f2 = TRUE, rate2 = TRUE, dur2 = TRUE),
resample = NULL,
resampleID = TRUE,
maxwhile = 1e+05
)

## Default S3 method:
rxSolve(
  object,
  params = NULL,
  events = NULL,
  inits = NULL,
  ...
)

## S3 method for class 'rxSolve'
update(object, ...)

## S3 method for class 'RxODE'
predict(object, ...)

## S3 method for class 'rxSolve'
predict(object, ...)

## S3 method for class 'rxEt'
predict(object, ...)

## S3 method for class 'rxParams'
predict(object, ...)

## S3 method for class 'RxODE'
simulate(object, nsim = 1L, seed = NULL, ...)

## S3 method for class 'rxSolve'
simulate(object, nsim = 1L, seed = NULL, ...)

## S3 method for class 'rxParams'
simulate(object, nsim = 1L, seed = NULL, ...)

## S3 method for class 'rxSolve'
solve(a, b, ...)

## S3 method for class 'RxODE'
solve(a, b, ...)

## S3 method for class 'rxParams'
solve(a, b, ...)

## S3 method for class 'rxEt'
solve(a, b, ...)

rxControl(..., params = NULL, events = NULL, inits = NULL)

Arguments

object is a either a RxODE family of objects, or a file-name with a RxODE model specification, or a string with a RxODE model specification.

params a numeric named vector with values for every parameter in the ODE system; the names must correspond to the parameter identifiers used in the ODE specification;

events an eventTable object describing the input (e.g., doses) to the dynamic system and observation sampling time points (see eventTable());

inits a vector of initial values of the state variables (e.g., amounts in each compartment), and the order in this vector must be the same as the state variables (e.g., PK/PD compartments);

scale a numeric named vector with scaling for ode parameters of the system. The names must correspond to the parameter identifiers in the ODE specification.
Each of the ODE variables will be divided by the scaling factor. For example `scale=c(center=2)` will divide the center ODE variable by 2.

**method**
The method for solving ODEs. Currently this supports:
- "liblsoda" thread safe lsoda. This supports parallel thread-based solving, and ignores user Jacobian specification.
- "lsoda" – LSODA solver. Does not support parallel thread-based solving, but allows user Jacobian specification.
- "dop853" – DOP853 solver. Does not support parallel thread-based solving nor user Jacobain specification
- "indLin" – Solving through inductive linearization. The RxODE dll must be setup specially to use this solving routine.

**transitAbs**
boolean indicating if this is a transit compartment absorption

**atol**
a numeric absolute tolerance (1e-8 by default) used by the ODE solver to determine if a good solution has been achieved; This is also used in the solved linear model to check if prior doses do not add anything to the solution.

**rtol**
a numeric relative tolerance (1e-6 by default) used by the ODE solver to determine if a good solution has been achieved. This is also used in the solved linear model to check if prior doses do not add anything to the solution.

**maxsteps**
maximum number of (internally defined) steps allowed during one call to the solver. (5000 by default)

**hmin**
The minimum absolute step size allowed. The default value is 0.

**hmax**
The maximum absolute step size allowed. When `hmax=NA` (default), uses the average difference + hmaxSd*sd in times and sampling events. The `hmaxSd` is a user specified parameter and which defaults to zero. When `hmax=NULL` RxODE uses the maximum difference in times in your sampling and events. The value 0 is equivalent to infinite maximum absolute step size.

**hmaxSd**
The number of standard deviations of the time difference to add to hmax. The default is 0

**hini**
The step size to be attempted on the first step. The default value is determined by the solver (when `hini = 0`)

**maxordn**
The maximum order to be allowed for the nonstiff (Adams) method. The default is 12. It can be between 1 and 12.

**maxords**
The maximum order to be allowed for the stiff (BDF) method. The default value is 5. This can be between 1 and 5.

...  
Other arguments including scaling factors for each compartment. This includes `S# = numeric` will scale a compartment # by a dividing the compartment amount by the scale factor, like NONMEM.

**cores**
Number of cores used in parallel ODE solving. This is equivalent to calling `setRxThreads()`

**covsInterpolation**
specifies the interpolation method for time-varying covariates. When solving ODEs it often samples times outside the sampling time specified in events. When this happens, the time varying covariates are interpolated. Currently this can be:
• "linear" interpolation, which interpolates the covariate by solving the line between the observed covariates and extrapolating the new covariate value.
• "constant" – Last observation carried forward (the default).
• "NOCB" – Next Observation Carried Backward. This is the same method that NONMEM uses.
• "midpoint" Last observation carried forward to midpoint; Next observation carried backward to midpoint.

addCov A boolean indicating if covariates should be added to the output matrix or data frame. By default this is disabled.

matrix A boolean indicating if a matrix should be returned instead of the RxODE’s solved object.

sigma Named sigma covariance or Cholesky decomposition of a covariance matrix. The names of the columns indicate parameters that are simulated. These are simulated for every observation in the solved system.

sigmaDf Degrees of freedom of the sigma t-distribution. By default it is equivalent to Inf, or a normal distribution.

sigmaLower Lower bounds for simulated unexplained variability (by default -Inf)

sigmaUpper Upper bounds for simulated unexplained variability (by default Inf)

nCoresRV Number of cores used for the simulation of the sigma variables. By default this is 1. To reproduce the results you need to run on the same platform with the same number of cores. This is the reason this is set to be one, regardless of what the number of cores are used in threaded ODE solving.

sigmaIsChol Boolean indicating if the sigma is in the Cholesky decomposition instead of a symmetric covariance

sigmaSeparation separation strategy for sigma;
   Tells the type of separation strategy when simulating covariance with parameter uncertainty with standard deviations modeled in the thetaMat matrix.

   • "lkj" simulates the correlation matrix from the rLKJ1 matrix with the distribution parameter eta equal to the degrees of freedom nu by \((nu-1)/2\)
   • "separation" simulates from the identity inverse Wishart covariance matrix with nu degrees of freedom. This is then converted to a covariance matrix and augmented with the modeled standard deviations. While computationally more complex than the "lkj" prior, it performs better when the covariance matrix size is greater or equal to 10
   • "auto" chooses "lkj" when the dimension of the matrix is less than 10 and "separation" when greater than equal to 10.

sigmaXform When taking sigma values from the thetaMat simulations (using the separation strategy for covariance simulation), how should the thetaMat values be turned into standard deviation values:

   • identity This is when standard deviation values are directly modeled by the params and thetaMat matrix
   • variance This is when the params and thetaMat simulates the variance that are directly modeled by the thetaMat matrix
• log This is when the params and thetaMat simulates log(sd)
• nlmixrSqrt This is when the params and thetaMat simulates the inverse cholesky decomposed matrix with the $x^2$ modeled along the diagonal. This only works with a diagonal matrix.
• nlmixrLog This is when the params and thetaMat simulates the inverse cholesky decomposed matrix with the $\exp(x^2)$ along the diagonal. This only works with a diagonal matrix.
• nlmixrIdentity This is when the params and thetaMat simulates the inverse cholesky decomposed matrix. This only works with a diagonal matrix.

nDisplayProgress
An integer indicating the minimum number of c-based solves before a progress bar is shown. By default this is 10,000.

discount
This supplies the dose units of a data frame supplied instead of an event table. This is for importing the data as an RxODE event table.

timeUnits
This supplies the time units of a data frame supplied instead of an event table. This is for importing the data as an RxODE event table.

stiff
a logical (TRUE by default) indicating whether the ODE system is stiff or not.

For stiff ODE systems (`stiff = TRUE`), `RxODE` uses the LSODA (Livermore Solver for Ordinary Differential Equations) Fortran package, which implements an automatic method switching for stiff and non-stiff problems along the integration interval, authored by Hindmarsh and Petzold (2003).

For non-stiff systems (`stiff = FALSE`), `RxODE` uses DOP853, an explicit Runge-Kutta method of order 8(5, 3) of Dormand and Prince as implemented in C by Hairer and Wanner (1993).

If stiff is not specified, the `method` argument is used instead.

theta
A vector of parameters that will be named THETA[#] and added to parameters

thetaLower
Lower bounds for simulated population parameter variability (by default -Inf)

thetaUpper
Upper bounds for simulated population unexplained variability (by default Inf)

eta
A vector of parameters that will be named ETA[#] and added to parameters

addDosing
Boolean indicating if the solve should add RxODE EVID and related columns. This will also include dosing information and estimates at the doses. Be default, RxODE only includes estimates at the observations. (default FALSE). When addDosing is NULL, only include EVID=0 on solve and exclude any model-times or EVID=2. If addDosing is NA the classic RxODE EVID events are returned. When addDosing is TRUE add the event information in NONMEM-style format; If subsetNonmem=FALSE RxODE will also include extra event types (EVID) for ending infusion and modeled times:

• EVID=-1 when the modeled rate infusions are turned off (matches rate=-1)
• EVID=-2 When the modeled duration infusions are turned off (matches rate=-2)
• EVID=-10 When the specified rate infusions are turned off (matches rate>0)
• EVID=-20 When the specified dur infusions are turned off (matches dur>0)
• EVID=101,102,103,... Modeled time where 101 is the first model time, 102 is the second etc.

**stateTrim**
When amounts/concentrations in one of the states are above this value, trim them to be this value. By default Inf. Also trims to -stateTrim for large negative amounts/concentrations. If you want to trim between a range say c(0,200000) you may specify 2 values with a lower and upper range to make sure all state values are in the reasonable range.

**updateObject**
This is an internally used flag to update the RxODE solved object (when supplying an RxODE solved object) as well as returning a new object. You probably should not modify it's FALSE default unless you are willing to have unexpected results.

**omega**
Estimate of Covariance matrix. When omega is a list, assume it is a block matrix and convert it to a full matrix for simulations.

**omegaDf**
The degrees of freedom of a t-distribution for simulation. By default this is NULL which is equivalent to Inf degrees, or to simulate from a normal distribution instead of a t-distribution.

**omegaIsChol**
Indicates if the omega supplied is a Cholesky decomposed matrix instead of the traditional symmetric matrix.

**omegaSeparation**
Omega separation strategy
Tells the type of separation strategy when simulating covariance with parameter uncertainty with standard deviations modeled in the thetaMat matrix.

• "lkj" simulates the correlation matrix from the rLKJ1 matrix with the distribution parameter eta equal to the degrees of freedom nu by (nu-1)/2
• "separation" simulates from the identity inverse Wishart covariance matrix with nu degrees of freedom. This is then converted to a covariance matrix and augmented with the modeled standard deviations. While computationally more complex than the "lkj" prior, it performs better when the covariance matrix size is greater or equal to 10
• "auto" chooses "lkj" when the dimension of the matrix is less than 10 and "separation" when greater than equal to 10.

**omegaXform**
When taking omega values from the thetaMat simulations (using the separation strategy for covariance simulation), how should the thetaMat values be turned into standard deviation values:

• identity This is when standard deviation values are directly modeled by the params and thetaMat matrix
• variance This is when the params and thetaMat simulates the variance that are directly modeled by the thetaMat matrix
• log This is when the params and thetaMat simulates log(sd)
• nlmixrSqrt This is when the params and thetaMat simulates the inverse cholesky decomposed matrix with the x^2 modeled along the diagonal. This only works with a diagonal matrix.
• \texttt{nlmixrLog} This is when the \texttt{params} and \texttt{thetaMat} simulates the inverse cholesky decomposed matrix with the \(\exp(x^2)\) along the diagonal. This only works with a diagonal matrix.

• \texttt{nlmixrIdentity} This is when the \texttt{params} and \texttt{thetaMat} simulates the inverse cholesky decomposed matrix. This only works with a diagonal matrix.

\[
\omega_{\text{Lower}} \quad \text{Lower bounds for simulated ETAs (by default -Inf)}
\]

\[
\omega_{\text{Upper}} \quad \text{Upper bounds for simulated ETAs (by default Inf)}
\]

\[
n_{\text{Sub}} \quad \text{Number between subject variabilities (ETAs) simulated for every realization of the parameters.}
\]

\[
\texttt{thetaMat} \quad \text{Named theta matrix.}
\]

\[
\texttt{thetaDf} \quad \text{The degrees of freedom of a t-distribution for simulation. By default this is NULL which is equivalent to Inf degrees, or to simulate from a normal distribution instead of a t-distribution.}
\]

\[
\texttt{thetaIsChol} \quad \text{Indicates if the theta supplied is a Cholesky decomposed matrix instead of the traditional symmetric matrix.}
\]

\[
n_{\text{Stud}} \quad \text{Number virtual studies to characterize uncertainty in estimated parameters.}
\]

\[
\texttt{df_{Sub}} \quad \text{Degrees of freedom to sample the between subject variability matrix from the inverse Wishart distribution (scaled) or scaled inverse chi squared distribution.}
\]

\[
\texttt{df_{Obs}} \quad \text{Degrees of freedom to sample the unexplained variability matrix from the inverse Wishart distribution (scaled) or scaled inverse chi squared distribution.}
\]

\[
\texttt{returnType} \quad \text{This tells what type of object is returned. The currently supported types are:}
\]

\[
\begin{array}{l}
\quad \text{• "rxSolve" (default) will return a reactive data frame that can change easily change different pieces of the solve and update the data frame. This is the currently standard solving method in RxODE, is used for \texttt{rxSolve(object,...)}, solve(object,...),}
\quad \text{• "data.frame" – returns a plain, non-reactive data frame; Currently very slightly faster than returnType="matrix"}
\quad \text{• "matrix" – returns a plain matrix with column names attached to the solved object. This is what is used object$run as well as object$solve}
\quad \text{• "data.table" – returns a data.table: The data.table is created by reference (ie setDt()), which should be fast.}
\quad \text{• "tbl" or "tibble" returns a tibble format.}
\end{array}
\]

\[
\texttt{seed} \quad \text{an object specifying if and how the random number generator should be initialized}
\]

\[
\texttt{nsim} \quad \text{represents the number of simulations. For RxODE, if you supply single subject event tables (created with [eventTable()])}
\]

\[
\text{\texttt{minSS}} \quad \text{Minimum number of iterations for a steady-state dose}
\]

\[
\text{\texttt{maxSS}} \quad \text{Maximum number of iterations for a steady-state dose}
\]

\[
\text{\texttt{infSSstep}} \quad \text{Step size for determining if a constant infusion has reached steady state. By default this is large value, 420.}
\]
strictSS: Boolean indicating if a strict steady-state is required. If a strict steady-state is (TRUE) required then at least minSS doses are administered and the total number of steady states doses will continue until maxSS is reached, or atol and rtol for every compartment have been reached. However, if ODE solving problems occur after the minSS has been reached the whole subject is considered an invalid solve. If strictSS is FALSE then as long as minSS has been reached the last good solve before ODE solving problems occur is considered the steady state, even though either atol, rtol or maxSS have not been achieved.

istateReset: When TRUE, reset the ISTATE variable to 1 for lsoda and liblsoda with doses, like deSolve; When FALSE, do not reset the ISTATE variable with doses.

subsetNonmem: subset to NONMEM compatible EVIDs only. By default TRUE.

maxAtolRtolFactor: The maximum atol/rtol that FOCEi and other routines may adjust to. By default 0.1

from: When there is no observations in the event table, start observations at this value. By default this is zero.

to: When there is no observations in the event table, end observations at this value. By default this is 24 + maximum dose time.

by: When there are no observations in the event table, this is the amount to increment for the observations between from and to.

length.out: The number of observations to create if there isn’t any observations in the event table. By default this is 200.

iCov: A data frame of individual non-time varying covariates to combine with the events dataset by merge.

keep: Columns to keep from either the input dataset or the iCov dataset. With the iCov dataset, the column is kept once per line. For the input dataset, if any records are added to the data LOCF (Last Observation Carried forward) imputation is performed.

indLinPhiTol: the requested accuracy tolerance on exponential matrix.

indLinPhiM: the maximum size for the Krylov basis

indLinMatExpType: This is them matrix exponential type that is use for RxODE. Currently the following are supported:
- Al-Mohy Uses the exponential matrix method of Al-Mohy Higham (2009)
- arma Use the exponential matrix from RcppArmadillo
- expokit Use the exponential matrix from Roger B. Sidje (1998)

indLinMatExpOrder: an integer, the order of approximation to be used, for the Al-Mohy and expokit values. The best value for this depends on machine precision (and slightly on the matrix). We use 6 as a default.

drop: Columns to drop from the output

idfactor: This boolean indicates if original ID values should be maintained. This changes the default sequentially ordered ID to a factor with the original ID values in the original dataset. By default this is enabled.
mxhnil  maximum number of messages printed (per problem) warning that $T + H = T$ on a step ($H$ = step size). This must be positive to result in a non-default value. The default value is 0 (or infinite).

hmxi  inverse of the maximum absolute value of $H$ to be used. $hmxi = 0.0$ is allowed and corresponds to an infinite $hmax1$ (default). $hmin$ and $hmxi$ may be changed at any time, but will not take effect until the next change of $H$ is considered. This option is only considered with method="liblsoda".

warnIdSort  Warn if the ID is not present and RxODE assumes the order of the parameters/iCov are the same as the order of the parameters in the input dataset.

warnDrop  Warn if column(s) were supposed to be dropped, but were not present.

ssAtol  Steady state atol convergence factor. Can be a vector based on each state.

ssRtol  Steady state rtol convergence factor. Can be a vector based on each state.

safeZero  Use safe zero divide and log routines. By default this is turned on but you may turn it off if you wish.

sumType  Sum type to use for sum() in RxODE code blocks.

pairwise uses the pairwise sum (fast, default)

fsum uses Python’s fsum function (most accurate)

kahan uses Kahan correction

neumaier uses Neumaier correction

c uses no correction: default/native summing

prodType  Product to use for prod() in RxODE blocks

long double converts to long double, performs the multiplication and then converts back.

double uses the standard double scale for multiplication.

sensType  Sensitivity type for linCmt() model:

advan Use the direct advan solutions

autodiff Use the autodiff advan solutions

forward Use forward difference solutions

central Use central differences

linDiff  This gives the linear difference amount for all the types of linear compartment model parameters where sensitivities are not calculated. The named components of this numeric vector are:

- "lag" Central compartment lag
- "f" Central compartment bioavailability
- "rate" Central compartment modeled rate
- "dur" Central compartment modeled duration
- "lag2" Depot compartment lag
- "f2" Depot compartment bioavailability
- "rate2" Depot compartment modeled rate
- "dur2" Depot compartment modeled duration

linDiffCentral  This gives the which parameters use central differences for the linear compartment model parameters. The are the same components as linDiff
resample

A character vector of model variables to resample from the input dataset; This sampling is done with replacement. When NULL or FALSE no resampling is done. When TRUE resampling is done on all covariates in the input dataset.

resampleID

boolean representing if the resampling should be done on an individual basis TRUE (ie. a whole patient is selected) or each covariate is resampled independent of the subject identifier FALSE. When resampleID=TRUE correlations of parameters are retained, whereas when resampleID=FALSE ignores patient covariate correlations. Hence the default is resampleID=TRUE.

maxwhile

represents the maximum times a while loop is evaluated before exiting. By default this is 100000

a

when using solve(), this is equivalent to the object argument. If you specify object later in the argument list it overwrites this parameter.

b

when using solve(), this is equivalent to the params argument. If you specify params as a named argument, this overwrites the output.

Details

The rest of the document focuses on the different ODE solving methods, followed by the core solving method’s options, RxODE event handling options, RxODE’s numerical stability options, RxODE’s output options, and finally internal RxODE options or compatibility options.

Value

An “rxSolve” solve object that stores the solved value in a special data.frame or other type as determined by returnType. By default this has as many rows as there are sampled time points and as many columns as system variables (as defined by the ODEs and additional assignments in the RxODE model code). It also stores information about the call to allow dynamic updating of the solved object.

The operations for the object are similar to a data-frame, but expand the $ and [[""""""]] access operators and assignment operators to resolve based on different parameter values, initial conditions, solver parameters, or events (by updating the time variable).

You can call the eventTable() methods on the solved object to update the event table and resolve the system of equations.

Author(s)

Matthew Fidler, Melissa Hallow and Wenping Wang

References

"New Scaling and Squaring Algorithm for the Matrix Exponential", by Awad H. Al-Mohy and Nicholas J. Higham, August 2009


**See Also**

RxODE()

---

### rxStack

*Stack a solved object for things like ggplot*

**Description**

Stack a solved object for things like ggplot

**Usage**

```r
rxStack(Data, vars = NULL)
```

**Arguments**

- **Data**
  - is a RxODE object to be stacked.
- **vars**
  - Variables to include in stacked data; By default this is all the variables when vars is NULL.

**Value**

Stacked data with `value` and `trt`, where `value` is the values and `trt` is the state and `lhs` variables.

**Author(s)**

Matthew Fidler

---

### rxState

*State variables*

**Description**

This returns the model’s compartments or states.

**Usage**

```r
rxState(obj = NULL, state = NULL)
```
**Arguments**

- **obj**  
  RxODE family of objects
- **state**  
  is a string indicating the state or compartment that you would like to lookup.

**Value**

If state is missing, return a character vector of all the states.
If state is a string, return the compartment number of the named state.

**Author(s)**

Matthew L. Fidler

**See Also**

RxODE()

---

**rxSumProdModel**  
*Recast model in terms of sum/prod*

**Description**

Recast model in terms of sum/prod

**Usage**

```r
rxSumProdModel(model, expand = FALSE, sum = TRUE, prod = TRUE)
```

**Arguments**

- **model**  
  RxODE model
- **expand**  
  Boolean indicating if the expression is expanded.
- **sum**  
  Use `sum(...)`
- **prod**  
  Use `prod(...)`

**Value**

model string with prod(.) and sum(.) for all these operations.

**Author(s)**

Matthew L. Fidler
**rxSupportedFuns**

*Get list of supported functions*

**Description**

Get list of supported functions

**Usage**

```r
rxSupportedFuns()
```

**Value**

list of supported functions in RxODE

**Examples**

```r
rxSupportedFuns()
```

---

**rxSuppressMsg**

*Respect suppress messages*

**Description**

This turns on the silent REprintf in C when suppressMessages() is turned on. This makes the REprintf act like messages in R, they can be suppressed with suppressMessages()

**Usage**

```r
rxSuppressMsg()
```

**Value**

Nothing

**Author(s)**

Matthew Fidler
Examples

```r
# rxSupressMsg() is called with RxODE()

# Note the errors are output to the console
try(RxODE("d/dt(matt)=3"), silent = TRUE)

# When using suppressMessages, the output is suppressed
suppressMessages(try(RxODE("d/dt(matt)=3"), silent = TRUE))

# In RxODE, we use REprintf so that interrupted threads do not crash R
# if there is a user interrupt. This isn't captured by R's messages, but
# This interface allows the `suppressMessages()` to suppress the C printing
# as well

# If you want to suppress messages from RxODE in other packages, you can use
# this function
```

---

**rxSymInvChol**  
*Get Omega^-1 and derivatives*

**Description**

Get Omega^-1 and derivatives

**Usage**

```r
rxSymInvChol(
  invObjOrMatrix,
  theta = NULL,
  type = "cholOmegaInv",
  thetaNumber = 0L
)
```

**Arguments**

- **invObjOrMatrix**  
  Object for inverse-type calculations. If this is a matrix, setup the object for inversion `rxSymInvCholCreate()` with the default arguments and return a reactive s3 object. Otherwise, use the inversion object to calculate the requested derivative/inverse.

- **theta**  
  Thetas to be used for calculation. If missing (NULL), a special s3 class is created and returned to access Omega^-1 objects as needed and cache them based on the theta that is used.

- **type**  
  The type of object. Currently the following types are supported:
  - `cholOmegaInv` gives the Cholesky decomposition of the Omega Inverse matrix.
• `omegaInv` gives the Omega Inverse matrix.
• `d(omegaInv)` gives the `d(Omega^{-1})` with respect to the theta parameter specified in `thetaNumber`.
• `d(D)` gives the `d(diagonal(Omega^{-1}))` with respect to the theta parameter specified in the `thetaNumber` parameter.

`thetaNumber` For types `d(omegaInv)` and `d(D)`, the theta number that the derivative is taken against. This must be positive from 1 to the number of thetas defining the Omega matrix.

Value
Matrix based on parameters or environment with all the matrixes calculated in variables `omega`, `omegaInv`, `dOmega`, `dOmegaInv`.

Author(s)
Matthew L. Fidler

---

**rxSyncOptions**

`Sync options with RxODE variables`

Description
Accessing RxODE options via `getOption` slows down solving. This allows the options to be synced with variables.

Usage
```
rxSyncOptions(setDefaults = c("none", "permissive", "strict"))
```

Arguments
- `setDefaults` This will setup RxODE’s default solving options with the following options:
  - "none" leave the options alone
  - "permissive" This is a permissive option set similar to R language specifications.
  - "strict" This is a strict option set similar to the original RxODE(). It requires semicolons at the end of lines and equals for assignment.

Value
nothing; called for side effects

Author(s)
Matthew L. Fidler
# rxSyntaxFunctions

**Description**

A list and description of Rode supported syntax functions

**Usage**

```r
rxSyntaxFunctions
```

**Format**

A data frame with 3 columns and 98 or more rows

<table>
<thead>
<tr>
<th>Function</th>
<th>Reserved function Name</th>
</tr>
</thead>
<tbody>
<tr>
<td>Description</td>
<td>Description of function</td>
</tr>
<tr>
<td>Aliases</td>
<td>Function Aliases</td>
</tr>
</tbody>
</table>

---

```r
rxt(df, n = 1L, ncores = 1L)
```

**Description**

Care should be taken with this method not to encounter the birthday problem, described [https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/](https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/). Since the sitmo threefry, this currently generates one random deviate from the uniform distribution to seed the engine threefry and then run the code.

**Usage**

```r
rxt(df, n = 1L, ncores = 1L)
```

**Arguments**

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>df</td>
<td>degrees of freedom (&gt; 0, maybe non-integer). df = Inf is allowed.</td>
</tr>
<tr>
<td>n</td>
<td>number of observations. If length(n) &gt; 1, the length is taken to be the number required.</td>
</tr>
<tr>
<td>ncores</td>
<td>Number of cores for the simulation</td>
</tr>
</tbody>
</table>

`rxxnorm` simulates using the threefry sitmo generator; `rxxnormV` uses the vandercorput generator
Details

Therefore, a simple call to the random number generated followed by a second call to random number generated may have identical seeds. As the number of random number generator calls are increased the probability that the birthday problem will increase.

The key to avoid this problem is to either run all simulations in the RxODE environment once (therefore one seed or series of seeds for the whole simulation), pre-generate all random variables used for the simulation, or seed the RxODE engine with \texttt{rxSetSeed()}

Also care should be made that the computer you will be running on can run the same number of cores as you are running so they can reproduce your results.

Value

t-distribution random numbers

Examples

```r
## Use threefry engine
rxt(df = 3, n = 10) # with rxt you have to explicitly state n
rxt(df = 3, n = 10, ncores = 2) # You can parallelize the simulation using openMP

rxt(4) ## The first argument is the df parameter

## This example uses `rxt` directly in the model
rx <- RxODE(
  a <- rxt(3)
)

et <- et(1, id = 1:2)

s <- rxSolve(rx, et)
```

**rxTempDir**

Get the RxODE temporary directory

Description

Get the RxODE temporary directory

Usage

\texttt{rxTempDir()}
**rxTheme**

**Value**

RxODE temporary directory.

---

**rxTheme**

*rxTheme is the RxODE theme for plots*

---

**Description**

rxTheme is the RxODE theme for plots

**Usage**

```r
rxTheme(
  base_size = 11,
  base_family = "",
  base_line_size = base_size/22,
  base_rect_size = base_size/22,
  grid = TRUE
)
```

**Arguments**

- `base_size` base font size, given in pts.
- `base_family` base font family
- `base_line_size` base size for line elements
- `base_rect_size` base size for rect elements
- `grid` a Boolean indicating if the grid is on (TRUE) or off (FALSE). This could also be a character indicating x or y.

**Value**

ggplot2 theme used in RxODE

---

**rxToSE**

*RxODE to symengine environment*

---

**Description**

RxODE to symengine environment
Usage

```r
rxToSE(x, envir = NULL, progress = FALSE, promoteLinSens = TRUE)
.rxToSE(x, envir = NULL, progress = FALSE)
rxFromSE(x, unknownDerivatives = c("forward", "central", "error"))
.rxFromSE(x)
```

Arguments

- `x` expression
- `envir` default is `NULL`; Environment to put symengine variables in.
- `progress` shows progress bar if true.
- `promoteLinSens` Promote solved linear compartment systems to sensitivity-based solutions.
- `unknownDerivatives` When handling derivatives from unknown functions, the translator will translate into different types of numeric derivatives. The currently supported methods are:
  - `forward` for forward differences
  - `central` for central differences
  - `error` for throwing an error for unknown derivatives

Value

An rxode symengine environment

Author(s)

Matthew L. Fidler

Description

This function translates the model to C code, if needed

Usage

```r
rxTrans(
  model,
  modelPrefix = "", 
  md5 = "",
  modName = NULL,
  rxClearOutput = FALSE
)
```
...}

## Default S3 method:
rxTrans(
  model,
  modelPrefix = "",
  md5 = "",
  modName = NULL,
  modVars = FALSE,
...
)

## S3 method for class 'character'
rxTrans(
  model,
  modelPrefix = "",
  md5 = "",
  modName = NULL,
  modVars = FALSE,
...
)

Arguments

- **model**
  - This is the ODE model specification. It can be:
    - a string containing the set of ordinary differential equations (ODE) and
      other expressions defining the changes in the dynamic system.
    - a file name where the ODE system equation is contained
      An ODE expression enclosed in \{\}
      (see also the filename argument). For details, see the sections “Details” and
      RxODE Syntax below.
- **modelPrefix**
  - Prefix of the model functions that will be compiled to make sure that multiple
    RxODE objects can coexist in the same R session.
- **md5**
  - Is the md5 of the model before parsing, and is used to embed the md5 into DLL,
    and then provide for functions like `rxModelVars()`.
- **modName**
  - a string to be used as the model name. This string is used for naming various
    aspects of the computations, including generating C symbol names, dynamic
    libraries, etc. Therefore, it is necessary that `modName` consists of simple ASCII
    alphanumeric characters starting with a letter.
- **modVars**
  - returns the model variables instead of the named vector of translated properties.
- **...**
  - Ignored parameters.

Value

- a named vector of translated model properties including what type of jacobian is specified, the C
  function prefixes, as well as the C functions names to be called through the compiled model.
rxunif

Simulate uniform variable from threefry generator

Description

Care should be taken with this method not to encounter the birthday problem, described [https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/](https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/). Since the sitmo threefry, this currently generates one random deviate from the uniform distribution to seed the engine threefry and then run the code.

Usage

`rxunif(min = 0, max = 1, n = 1L, ncores = 1L)`

Arguments

- `min` lower and upper limits of the distribution. Must be finite.
- `max` lower and upper limits of the distribution. Must be finite.
- `n` number of observations. If `length(n) > 1`, the length is taken to be the number required.
- `ncores` Number of cores for the simulation

Details

Therefore, a simple call to the random number generated followed by a second call to random number generated may have identical seeds. As the number of random number generator calls are increased the probability that the birthday problem will increase.

The key to avoid this problem is to either run all simulations in the RxODE environment once (therefore one seed or series of seeds for the whole simulation), pre-generate all random variables used for the simulation, or seed the RxODE engine with `rxSetSeed()`.

Also care should be made that the computer you will be running on can run the same number of cores as you are running so they can reproduce your results.

Value

uniform random numbers
Examples

```r
## Use threefry engine

rxunif(min = 0, max = 4, n = 10) # with rxunif you have to explicitly state n
rxunif(min = 0, max = 4, n = 10, ncores = 2) # You can parallelize the simulation using openMP

rxunif()

## This example uses `rxunif` directly in the model

rx <- RxODE(
  a <- rxunif(0, 3)
)

et <- et(1, id = 1:2)
s <- rxSolve(rx, et)
```

### Description

Unloads all RxODE compiled DLLs

### Usage

```r
rxUnloadAll()
```

### Value

List of RxODE dlls still loaded

boolean of if all RxODE dlls have been unloaded

### Examples

```r
print(rxUnloadAll())
```
**rxUse**

*Use model object in your package*

**Description**

Use model object in your package

**Usage**

```
rxUse(obj, overwrite = TRUE, compress = "bzip2", internal = FALSE)
```

**Arguments**

- `obj`: model to save.
- `overwrite`: By default, `use_data()` will not overwrite existing files. If you really want to do so, set this to TRUE.
- `compress`: Choose the type of compression used by `save()`. Should be one of "gzip", "bzip2", or "xz".
- `internal`: If this is run internally. By default this is FALSE

**Value**

Nothing; This is used for its side effects and shouldn’t be called by a user

---

**rxValidate**

*Validate RxODE This allows easy validation/qualification of nlmixr by running the testing suite on your system.*

**Description**

Validate RxODE This allows easy validation/qualification of nlmixr by running the testing suite on your system.

**Usage**

```
rxValidate(type = NULL)
rxTest(type = NULL)
```

**Arguments**

- `type`: Type of test or filter of test type

**Value**

nothing
rxweibull

Simulate Weibull variable from the threefry generator

**Author(s)**

Matthew L. Fidler

---

**Description**

Care should be taken with this method not to encounter the birthday problem, described [here](https://www.johndcook.com/blog/2016/01/29/random-number-generator-seed-mistakes/). Since the sitmo threefry, this currently generates one random deviate from the uniform distribution to seed the engine threefry and then run the code.

**Usage**

```r
rxweibull(shape, scale = 1, n = 1L, ncores = 1L)
```

**Arguments**

- `shape`  
  shape and scale parameters, the latter defaulting to 1.
- `scale`  
  shape and scale parameters, the latter defaulting to 1.
- `n`  
  number of observations. If `length(n) > 1`, the length is taken to be the number required.
- `ncores`  
  Number of cores for the simulation

**Details**

Therefore, a simple call to the random number generated followed by a second call to random number generated may have identical seeds. As the number of random number generator calls are increased the probability that the birthday problem will increase.

The key to avoid this problem is to either run all simulations in the RxODE environment once (therefore one seed or series of seeds for the whole simulation), pre-generate all random variables used for the simulation, or seed the RxODE engine with `rxSetSeed()`

Also care should be made that the computer you will be running on can run the same number of cores as you are running so they can reproduce your results.

**Value**

Weibull random deviates
Examples

```r
## Use threefry engine

# with rxweibull you have to explicitly state n
rxweibull(shape = 1, scale = 4, n = 10)

# You can parallelize the simulation using openMP
rxweibull(shape = 1, scale = 4, n = 10, ncores = 2)

rxweibull(3)

## This example uses `rxweibull` directly in the model

rx <- RxODE({
  a <- rxweibull(1, 3)
})

et <- et(1, id = 1:2)

s <- rxSolve(rx, et)
```

---

**rxWinSetup**

*Setup Windows components for RxODE*

**Description**

Setup Windows components for RxODE

**Usage**

```r
rxWinSetup(rm.rtools = TRUE)
```

**Arguments**

- `rm.rtools` Remove the Rtools from the current path specs.

**Value**

nothing, used for its side effects

**Author(s)**

Matthew L. Fidler
Description

This is a dosing geom that shows the vertical lines where a dose occurs.

Usage

```r
stat_amt(
  mapping = NULL,
  data = NULL,
  position = "identity",
  show.legend = NA,
  inherit.aes = TRUE,
  ...
)
```

```r
geom_amt(
  mapping = NULL,
  data = NULL,
  position = "identity",
  show.legend = NA,
  inherit.aes = TRUE,
  ...
)
```

Arguments

- **mapping**: Set of aesthetic mappings created by `aes()` or `aes()`. If specified and `inherit.aes = TRUE` (the default), it is combined with the default mapping at the top level of the plot. You must supply `mapping` if there is no plot mapping.

- **data**: The data to be displayed in this layer. There are three options:
  - If `NULL`, the default, the data is inherited from the plot data as specified in the call to `ggplot()`.
  - A `data.frame`, or other object, will override the plot data. All objects will be fortified to produce a data frame. See `fortify()` for which variables will be created.
  - A function will be called with a single argument, the plot data. The return value must be a `data.frame`, and will be used as the layer data. A function can be created from a formula (e.g. `~ head(.x, 10)`).

- **position**: Position adjustment, either as a string, or the result of a call to a position adjustment function.

- **show.legend**: logical. Should this layer be included in the legends? `NA`, the default, includes if any aesthetics are mapped. `FALSE` never includes, and `TRUE` always includes. It can also be a named logical vector to finely select the aesthetics to display.
In `stat_cens`:

- `inherit.aes` If `FALSE`, overrides the default aesthetics, rather than combining with them. This is most useful for helper functions that define both data and aesthetics and shouldn’t inherit behaviour from the default plot specification, e.g. `borders()`.

- `...` Other arguments passed on to `layer()`. These are often aesthetics, used to set an aesthetic to a fixed value, like `colour = "red"` or `size = 3`. They may also be parameters to the paired geom/stat.

**Details**

Requires the following aesthetics:

- `x` representing the x values, usually time
- `amt` representing the dosing values; They are missing or zero when no dose is given

**Value**

This returns a stat_amt in context of a ggplot2 plot

---

**stat_cens**

*Censoring geom/stat*

**Description**

This is a censoring geom that shows the left or right censoring specified in the nlmixr input data-set or fit

**Usage**

```r
stat_cens(
  mapping = NULL,
  data = NULL,
  position = "identity",
  show.legend = NA,
  inherit.aes = TRUE,
  width = 0.01,
  ...
)
```

```r
gem_cens(
  mapping = NULL,
  data = NULL,
  position = "identity",
  show.legend = NA,
  inherit.aes = TRUE,
  width = 0.01,
  ...
)
```
Arguments

mapping  Set of aesthetic mappings created by `aes()` or `aes_()`. If specified and `inherit.aes = TRUE` (the default), it is combined with the default mapping at the top level of the plot. You must supply `mapping` if there is no plot mapping.

data     The data to be displayed in this layer. There are three options:
          If NULL, the default, the data is inherited from the plot data as specified in the call to `ggplot()`.
          A data.frame, or other object, will override the plot data. All objects will be fortified to produce a data frame. See `fortify()` for which variables will be created.
          A function will be called with a single argument, the plot data. The return value must be a data.frame, and will be used as the layer data. A function can be created from a formula (e.g. `~ head(.x,10)`).

position Position adjustment, either as a string, or the result of a call to a position adjustment function.

show.legend logical. Should this layer be included in the legends? `NA`, the default, includes if any aesthetics are mapped. `FALSE` never includes, and `TRUE` always includes. It can also be a named logical vector to finely select the aesthetics to display.

inherit.aes If `FALSE`, overrides the default aesthetics, rather than combining with them. This is most useful for helper functions that define both data and aesthetics and shouldn’t inherit behaviour from the default plot specification, e.g. `borders()`.

width     represents the width (in censoring box

...      Other arguments passed on to `layer()`. These are often aesthetics, used to set an aesthetic to a fixed value, like `colour = "red"` or `size = 3`. They may also be parameters to the paired geom/stat.

Details

Requires the following aesthetics:

- `x` Represents the independent variable, often the time scale
- `y` represents the dependent variable
- `CENS` for the censoring information; (-1 right censored, 0 no censoring or 1 left censoring)
- `LIMIT` which represents the corresponding limit ()

Will add boxes representing the areas of the fit that were censored.

Value

This returns a ggplot2 stat
summary.RxODE  

Print expanded information about the RxODE object.

Description

This prints the expanded information about the RxODE object.

Usage

## S3 method for class 'RxODE'
summary(object, ...)

Arguments

object  
RxODE object

...  
Ignored parameters

Value

object is returned

Author(s)

Matthew L. Fidler

uppergamma

uppergamma: upper incomplete gamma function

Description

This is the tgamma from the boost library

Usage

uppergamma(a, z)

Arguments

a  
The numeric 'a' parameter in the upper incomplete gamma

z  
The numeric 'z' parameter in the upper incomplete gamma

Details

The uppergamma function is given by:

\[ \text{uppergamma}(a, z) = \int_z^\infty t^{a-1} e^{-t} \, dt \]
**Value**

uppergamma results

**Author(s)**

Matthew L. Fidler

**Examples**

uppergamma(1, 3)

uppergamma(1:3, 3)

uppergamma(1, 1:3)
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