

Package ‘Qindex’

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Description Select optimal quantile-based predictors for survival outcome, to include the means to handle dichotomizing the quantiles, mean-signal-intensity or other continuous markers, and to obtain estimates for dichotomized predictors by Bootstrap-based bias correction.

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'BBC_dichotom.R' 'FRindex.R'

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Qindex-package	<i>Qindex: Continuous and dichotomized index predictors based on distribution quantiles</i>
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Description

Select optimal functional or dichotomized quantile-based predictors for survival/logistic/numeric outcome and perform optimal dichotomization with optimistic bias correction for any continuous predictors

Details

The package provides tools to

1. use [sampleQp](#) to calculate user-selected sample quantiles in each independent cluster of observations. This function is simply a wrapper of [aggregate](#) for [quantile](#) function.
2. use [eval_dichotom](#) to estimate the effect size for dichotomized user-selected predictors of interest (e.g. sample quantiles) (absolute value of the corresponding hazard ratio, odds ratio or regression coefficient) using repeated random split train/test sampling. In this function, we first use [rpart](#) to identify optimal cutoff in each training set and use this cutoff to dichotomize each predictor of interest in the corresponding independent test set. The effect size for dichotomized predictor is estimated in the test set by fitting [coxph](#), [glm](#) or [lm](#) to fit a Cox proportional hazard model, logistic regression, or linear regression for [Surv](#), [logical](#), or [numeric](#) endpoint.
3. use [optQp](#) to select the set of optimal quantiles that has the largest effect size (absolute value of the corresponding hazard ratio, odds ratio or regression coefficient) for a given [Surv](#), [logical](#), or [numeric](#) endpoint. [optQp](#) is a wrapper of [summary.eval_dichotom](#).
4. use [BBC_dichotom](#) to dichotomize predictors of interest and to obtain bootstrap-based optimism corrected effect size from Cox model, logistic regression, or linear regression. Internally, [BBC_dichotom](#) calls [dichotom_int](#) to dichotomize each predictor in `contX` based on univariate model setting and [model_dichotom](#) to fit Cox proportional hazard model, logistic

regression, or linear regression for [Surv](#), [logical](#), or [numeric](#) endpoint with the dichotomized predictors from [dichotom_int](#).

5. use [FRindex](#) to derive a scalar functional regression index as a predictor in the functional regression model with any response supported by [gam](#) package. Function [FRindex](#) calls [gam](#) to fit a generalized additive model(GAM) to the training set and makes use of [plot.gam](#) to extract functional coefficient tabulated on the same grid as functional predictor(s) in the training and test set (if the test set is provided).

References

Selection of optimal quantile protein biomarkers based on cell-level immunohistochemistry data, Misung Yi, Tingting Zhan , Amy P. Peck, Jeffrey A. Hooke, Albert J. Kovatich, Craig D. Shriver, Hai Hu, Yunguang Sun, Hallgeir Rui and Inna Chervoneva, under review

Quantile index biomarkers based on single-cell expression data, Misung Yi, Tingting Zhan , Amy P. Peck, Jeffrey A. Hooke, Albert J. Kovatich, Craig D. Shriver, Hai Hu, Yunguang Sun, Hallgeir Rui and Inna Chervoneva, under review

BBC_dichotom

Bootstrap Bias Correction for Dichotomization

Description

Bootstrap-based optimism correction for dichotomizing selected continuous predictor(s)

Usage

```
BBC_dichotom(formula, data, contX, R = 200L, ...)
```

Arguments

formula	formula , with a Surv , logical , or numeric endpoint and covariates to be included as is (i.e., without dichotomization).
data	data.frame
contX	character scalar, name of the matrix containing the continuous predictor(s) to be dichotomized optimally
R	integer scalar, number of bootstrap samples, see boot . Default 200L.
...	additional parameters, currently not in use

Details

The bootstrap optimism correction procedure is performed as described for a general model selection. First, R bootstrap samples are drawn with replacement from the main sample. In each bootstrap sample, the recursive partitioning tree model is used to establish an objective data-driven optimal cut-point for selected continuous predictors The cut-point from the current bootstrap sample is used to compute the effect size (log hazards ratio (HR), odds ratio (OR), or coefficient) for each dichotomized predictor in the current bootstrap sample (**bootstrap performance**) and in the main

sample (**test performance**), and the optimism in log OR/HR or coefficient estimation is computed as the difference between log OR/HR or coefficient for "Bootstrap performance" and for "Test performance". The median optimism estimate is computed as the median of optimism estimates over all bootstrap samples. The cutpoint for dichotomizing each selected continuous predictor is also established in the main sample and its "apparent performance" is computed as the log OR/HR or coefficient for dichotomized quantile in the univariate Cox models, logistic regression, or linear regression. Finally, the optimism-corrected performance estimate is computed by subtracting the median optimism estimate from the apparent performance estimate.

Value

`BBC_dichotom` returns a **numeric** bootstrap-based bias adjusted coefficients of model, corresponding median optimism, R bootstrap resampling based thresholds, and a single apparent performance threshold

References

Ewout W. Steyerberg (2009) Clinical Prediction Models. [doi:10.1007/9780387772448](https://doi.org/10.1007/9780387772448)

Frank E. Harrell Jr., Kerry L. Lee, Daniel B. Mark. (1996) Multivariable prognostic models: issues in developing models, evaluating assumptions and adequacy, and measuring and reducing errors. [doi:10.1002/\(SICI\)10970258\(19960229\)15:4<361::AIDSIM168>3.0.CO;24](https://doi.org/10.1002/(SICI)10970258(19960229)15:4<361::AIDSIM168>3.0.CO;24)

Examples

```
# see ?FRindex
# see ?eval_dichotom
```

celldata

Ki67 Data

Description

Ki67 cell data containing 622 patients

Usage

Ki67

Format

PATIENT_ID **factor**, unique patient identifier
 tiissueID **factor**, ID of the TMA core
 RECURRENCE **integer**, recurrence indicator, 1 = Recurred, 0 = not Recurred
 RECFREESURV_MO **integer**, recurrence-free survival time in months
 Marker **numeric**, cell signal intensity of the protein immunofloerscence signal

inner_x [integer](#), x -coordinate in the cell centroid in the TMA core
 inner_y [integer](#), y -coordinate in the cell centroid in the TMA core
 AGE_AT_DX [integer](#), age at diagnosis
 Tstage [integer](#), tumor stage
 NodeSt [integer](#), node stage, -1 = unknown, 0 = Node Negative, 1 = Node Positive
 HRpos [integer](#), indicator of hormone positive status (ER+ or PR+), 1 = positive, 0 = negative
 HistologicalGrade [integer](#), histology grade
 Her2_path_qIF [integer](#), Her2 status, 1 = positive, 0 = negative
 RACE [character](#), race, White, Black, Asian, Native Hawaiian or Other Pacific Islander, American Indian or Alaska Native, Unknown
 RadjCHEMO [integer](#), adjuvant chemo treatment, 0 = unknown, 1 = done, 2 = NOT done
 RadjRAD [integer](#), adjuvant radiation treatment, 0 = unknown, 1 = done, 2 = NOT done
 HORM_4cat [integer](#), hormone treatment, 0 = unknown, 1 = not indicated, 2 = done, 3 = recommended, but not done
 MSI [numeric](#), mean signal intensity (mean over all cells in the TMA core)

dichotom_int

Dichotomizing

Description

Generate dichotomized variables by a threshold calculated from [rpart](#) using [Surv](#), [logical](#), or [numeric](#) endpoint and each quantile

Usage

```
dichotom_int(edp, contX)
```

Arguments

edp [Surv](#), [logical](#), or [numeric](#) object, the endpoint
 contX see [BBC_dichotom](#)

Value

[dichotom_int](#) returns a [logical matrix](#) of the same dimension and dimension names as argument contX, as dichotomized using the first node of [rpart](#) as threshold. The thresholds (per column) are returned as an [numeric](#) vector in attribute 'thres'.

eval_dichotom	<i>Optimal Quantile Predictor</i>
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Description

From a given set of sample quantiles, this function selects the optimal quantile with the largest effect size for predicting given [Surv](#), [logical](#), or [numeric](#) outcome

Usage

```
eval_dichotom(formula, data, seeds, pct_train = 0.8, ...)
```

Arguments

formula	formula , supports Surv , logical , or numeric endpoint and one matrix predictor of the descriptive statistics of markers per cluster. See details of parameter data.
data	data.frame , with at least <ul style="list-style-type: none"> • two Surv columns including time-to-event and event indicator, one logical column, or one numeric column as the outcome • one matrix column as the descriptive statistics of markers per cluster. Currently only a quantile sequence is supported.
seeds	integer vector of random seeds for generating repeated random split samples, see set.seed
pct_train	numeric scalar, proportion of the training set, default .8
...	additional parameters, currently not in use

Details

Optimal quantile selection algorithm For a sample $x_i, 1 \leq i \leq n$ of repeated measures of independent variable X and probability $p, 0 < p < 1$, the empirical quantile function $Q_n(p)$ is defined as the k th order statistic of the sample, where k is such that $(k - 1)/n < p < k/n$. For $p = .01, \dots, .99$ and $k = 100p$, $Q_n(p)$ is also known as k th percentile. The following algorithm is proposed to identify the optimal $Q(p)$ predictor of survival outcome in a screening data set:

1. Select the set of quantiles to be evaluated as predictors and the desired ratio for training/test sets.
2. Split the data into training and test sets
 - In case of [Surv](#) outcomes, split the group of subjects with event into a training set and a test set randomly with desired ratio. Similarly, split the group of subjects without event into a training set and a test set randomly with desired ratio. Combine training sets with and without event and test sets with and without event.
 - In case of [logical](#) outcomes, split the group of subjects with one level of outcome into a training set and a test set randomly with desired ratio. Similarly, split the group of subjects with the other level of outcome into a training set and a test set randomly with desired ratio. Combine training sets with both levels of logical outcome and test sets with both levels of logical outcome.

- In case of **numeric** outcomes, split the entire subjects into a training set and a test set randomly with desired ratio.
3. For each training/test set pair and each considered quantile,
 - Determine the optimal cutoff (e.g. using the R package rpart) in the combined training set.
 - Apply the optimal cutoff to the combined test set and estimate the effect size (hazard ratio, odds ratio, or exponentiated coefficient).
 4. Repeat steps 2 and 3 for 100 training/test splits, compute the median log effect size (log hazard ratio(HR), log odds ratio(OR), or coefficient) for each quantile.
 5. Rank the effect sizes (absolute value of log OR, HR, or coefficient) for all considered quantiles and select the optimal quantile with the highest effect size.
 6. Perform bootstrap-based optimism correction for the selected optimal quantile(s). In this work, for each random split, 80% of subjects were assigned to the training set and 20% of subjects were assigned to the test set. We considered every fifth quantile starting from the 5th quantile to the 95th quantile plus 99th quantile as candidate predictors of PFS (a total of 20 quantile predictors). Also, we identified quantiles with the second and third highest effect sizes to compare them to the optimal ones.

Value

`eval_dichotom` returns a `eval_dichotom` object, which is a `list` with elements `thresholds` and `coefs`.

`thresholds` `matrix`, data set with all cut points for all candidate quantiles and `Np` columns and `Nsplit` rows

`coefs` `matrix`, coefficients corresponding to thresholds

`data` `data.frame`, this is the unmodified input data

References

Selection of optimal quantile protein biomarkers based on cell-level immunohistochemistry data, Misung Yi, Tingting Zhan, Amy P. Peck, Jeffrey A. Hooke, Albert J. Kovatich, Craig D. Shriver, Hai Hu, Yunguang Sun, Hallgeir Rui and Inna Chervoneva, under review

Examples

```
if (FALSE) { # masked to save time
library(survival)

Ki67_Qps = sampleQp(data = Ki67, subjID = 'PATIENT_ID',
  exclude = c('tissueID', 'inner_x', 'inner_y'), Qpredictor = 'Marker')
Ki67c = eval_dichotom(Surv(RECFREESURV_MO, RECURRENCE) ~ Marker, data = Ki67_Qps,
  seeds = 1:20)
# summary(Ki67c) # works, but not needed
head(Ki67_opt <- optQp(Ki67c, n = 2L))

set.seed(1)
```

```

mod_c = BBC_dichotom(Surv(RECFREESURV_MO, RECURRENCE) ~ NodeSt + Tstage,
  data = optQp(Ki67c, n = 2L), contX = 'Marker', R = 100)
summary(mod_c)
}

if (FALSE) { # mask to save time
Ki67a = eval_dichotom(RECFREESURV_MO ~ Marker, data = Ki67_Qps, seeds = 1:20)
set.seed(1)
mod_a = BBC_dichotom(RECFREESURV_MO ~ NodeSt + Tstage, data = optQp(Ki67a, n = 2L),
  contX = 'Marker', R = 100)
summary(mod_a)

Ki67b = eval_dichotom(RECURRENCE ~ Marker, data = Ki67_Qps, seeds = 1:20)
set.seed(1)
mod_b = BBC_dichotom(RECURRENCE ~ NodeSt + Tstage, data = optQp(Ki67b, n = 2L),
  contX = 'Marker', R = 100)
summary(mod_b)
}

```

FRindex

Functional Regression Index (FRindex)

Description

Computes a scalar functional index as a predictor in the functional regression model. The training data are used to estimate the functional coefficient by fitting functional regression model with any response supported by **gam** package. The functional regression index is computed for training and (optional) test data using the functional coefficient from the model fitted to the training data.

Usage

```

FRindex(
  trainData,
  testData,
  trainArgs = attr(trainData, which = "xarg", exact = TRUE),
  testArgs = if (!missing(testData)) attr(testData, which = "xarg", exact = TRUE) else
    trainArgs,
  response,
  family,
  predictor = "Marker",
  log = TRUE,
  knot_pct = 0.4,
  knot.value = ceiling(ncol(trainData[[predictor]]) * knot_pct),
  ...
)

```

Arguments

trainData	data.frame , training data including a response variable and a matrix of tabulated functional predictor. If the functional predictor is the quantile function, then trainData is preferably the returned object of sampleQp .
testData	data.frame , test data including a response variable and a matrix of tabulated functional predictor. The number of values in the grid for tabulating functional predictor does not need to be the same as that number for the training data. If the functional predictor is the quantile function, then testData is preferably the returned object of sampleQp .
trainArgs	numeric vector
testArgs	numeric vector
response	language of the response variable. All response variable types supported by gam package are allowed, including continuous, binary or survival outcome.
family	family object specifying the distribution and link to use in gam
predictor	character scalar, name of the matrix column variable in trainData and testData, with each row representing the tabulated, on a common grid, functional predictor values for each subject. If the functional predictor is the quantile function, then the number of columns in the matrix column identified by predictor equals to the number of quantiles used.
log	logical scalar, whether to perform log transformation on quantiles (default FALSE).
knot_pct	numeric scalar, percentage of the column dimension of the matrix column predictor, to be used as <code>knot.value</code> . Default is 40%. If <code>knot.value</code> is provided by the end-user, then <code>knot_pct</code> is not used.
knot.value	integer scalar, number of knots (i.e., parameter <code>k</code> in the spline smooth function <code>s</code>) used in gam function. Default is the ceiling of <code>knot_pct</code> of the column dimension of the matrix column predictor.
...	additional parameters, currently not in use

Details

The Functional Regression Index ([FRindex](#)) is defined as the integral of the functional predictor multiplied by common weight function. The weight function is proportional to the functional coefficient in the functional regression model for a given response variable fitted to training data. The [FRindex](#) values computed for the training data and test data, if provided.

Value

[FRindex](#) returns a [data.frame](#) with the training data and [FRindex](#), and the vectors of weights on the grid of the functional predictor

References

- J. O. Ramsay, B. W. Silverman (2005). Functional Data Analysis, ed 2. Springer New York, NY [doi:10.1007/b98888](#)
- Cui, E., Crainiceanu, C. M., & Leroux, A. (2021). Additive Functional Cox Model. Journal of Computational and Graphical Statistics. 2021;30(3):780-793. [doi:10.1080/10618600.2020.1853550](#)

Gellar, J. E., Colantuoni, E., Needham, D. M., & Crainiceanu, C. M. (2015). Cox regression models with functional covariates for survival data. *Statistical Modelling*, 15(3), 256-278. doi:10.1177/1471082X14565526

Examples

```
pt = unique(Ki67$PATIENT_ID)
length(pt) # 622
train = subset(Ki67, PATIENT_ID %in% pt[1:500])
test = subset(Ki67, PATIENT_ID %in% pt[501:622])
train_Qps = sampleQp(data = train, subjID = 'PATIENT_ID',
  exclude = c('tissueID', 'inner_x', 'inner_y'), Qpredictor = 'Marker')
test_Qps = sampleQp(data = test, subjID = 'PATIENT_ID',
  exclude = c('tissueID', 'inner_x', 'inner_y'), Qpredictor = 'Marker')

if (FALSE) { # masked to save time
FRQI = FRindex(trainData = train_Qps, testData = test_Qps,
  response = Surv(RECFREESURV_MO, RECURRENCE), predictor = 'Marker', log = TRUE)
set.seed(1)
mod = BBC_dichotom(Surv(RECFREESURV_MO, RECURRENCE) ~ NodeSt + Tstage, data = FRQI$trainData,
  contX = 'FRindex_std', R = 100)
}

if (FALSE) { # masked to save time
FRindex(trainData = train_Qps, testData = test_Qps, response = RECURRENCE)
FRindex(trainData = train_Qps, testData = test_Qps, response = RECFREESURV_MO)

set.seed(1)
mod = BBC_dichotom(Surv(RECFREESURV_MO, RECURRENCE) ~ 1, data = FRQI$trainData,
  contX = 'FRindex_std', R = 200)
summary(mod)
names(attributes(mod))
attr(mod, 'median_optimism')
}
```

model_dichotom

Model with Dichotomized Predictors

Description

fit Cox proportional hazard model, logistic regression, or linear regression with dichotomized marker and/or other covariates built as formula in [BBC_dichotom](#)

Usage

```
model_dichotom(
  formula,
  data,
  contX,
```

```
dQ = dichotom_int(edp = eval(formula[[2L]], envir = data), contX = data[[contX]])
)
```

Arguments

formula	same formula in BBC_dichotom
data	data.frame
contX	see BBC_dichotom
dQ	returned object from dichotom_int

Value

[model_dichotom](#) returns a [numeric](#) vector of the concatenated coefficients.

optQp	<i>Optimal Quantile Predictor</i>
-------	-----------------------------------

Description

Wrapper of [eval_dichotom](#).

To create a [data.frame](#) with designated number of optimal quantiles returned from [eval_dichotom](#) in addition to subjID, [Surv](#) endpoint and [numeric](#), [character](#), or [logical](#) predictors

Usage

```
optQp(x, n = 3L, ...)
```

Arguments

x	eval_dichotom object
n	integer scalar, number of optimal quantiles to be printed
...	additional parameters, not currently in use

Value

[optQp](#) returns a [data.frame](#) with designated number of optimal quantiles

Examples

```
# see ?BBC_dichotom
```

`print.eval_dichotom` *Print [eval_dichotom](#) Object*

Description

To print the optimal quantile that has the largest absolute value of coefficients for a given [Surv](#) outcome, [logical](#), or [numeric](#) outcome

Usage

```
## S3 method for class 'eval_dichotom'
print(x, n = 3L, ...)
```

Arguments

`x` [eval_dichotom](#) object
`n` [integer](#) scalar, number of optimal quantiles to be printed
`...` additional parameters, currently not in use

Details

[print.eval_dichotom](#) simply calls [summary.eval_dichotom](#) and print three largest absolute value of coefficients, hazard ratio or odds ratio, and threshold

Value

[print.eval_dichotom](#) does not have a returned value.

`sampleQp` *Cluster-Specific Sample Quantiles*

Description

This function calculates vectors of sample quantiles in each independent cluster of observations (sample or subject) Equidistant probabilities between user provided `p_min` and `p_max` are used (including both ends).

Usage

```
sampleQp(
  data,
  subjID = "PATIENT_ID",
  Qpredictor = "Marker",
  include = setdiff(names(data), y = c(Qpredictor)),
  exclude = NULL,
  from = 0.01,
  to = 0.99,
  by = 0.01,
  type = 7,
  ...
)
```

Arguments

data	data.frame
subjID	character scalar, column name of the subject/patient index in data
Qpredictor	character scalar, column name of the predictor variable
include	character vector , predictors to be included in output data set
exclude	character vector , predictors to be excluded from output data set
from, to, by	see seq a sequence of probabilities with starting, ending values, and interval to calculate corresponding quantiles
type	integer scalar, type of the formula for quantiles, see quantile
...	additional parameters, currently not in use

Details

[sampleQp](#) calculates N_p sample quantiles in each independent cluster of observations defined by `subjID` or `sampleID`. Sample quantiles are stored in N_p columns for $Q(p)$, $p = 1, \dots, N_p$. Additional subject-level predictors may be designated to be kept in the output data set. Observation-level predictors should be excluded from the input data set.

Value

[sampleQp](#) returns a [data.frame](#), with aggregated [Surv](#) endpoint and [numeric](#), [character](#), or [logical](#) predictors, in addition to a [matrix](#) of quantiles.

Examples

```
Ki67_Qps = sampleQp(data = Ki67, subjID = 'PATIENT_ID',
  exclude = c('tissueID', 'inner_x', 'inner_y'), Qpredictor = 'Marker')
head(Ki67_Qps)
sapply(Ki67_Qps, FUN = class)
head(Ki67_Qps$Marker)
```

summary.eval_dichotom *Summary Information of [eval_dichotom](#) Object*

Description

To summarize [eval_dichotom](#) object

Usage

```
## S3 method for class 'eval_dichotom'  
summary(object, FUN = median, ...)
```

Arguments

object	eval_dichotom object
FUN	summarizing function , either median (default) or mean
...	additional parameters, currently not in use

Details

[summary.eval_dichotom](#) present effect sizes (absolute value of log hazard ratio, log odds ratio, or coefficient), corresponding hazard ratio or odds ratio, and threshold to dichotomize quantile. And then sort by `abs.coef`

Value

[summary.eval_dichotom](#) returns a [data.frame](#) with three (3) columns

`abs.coef` effect sizes (absolute value of log hazard ratio, log odds ratio, or coefficient)

`HR` corresponding hazard ratio or odds ratio

`threshold` threshold for dichotomizing the quantile

See Also

[summary](#)

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